

bradscholars

Cross-talk and regulation between glutamate and GABAB receptors

Item Type	Article
Authors	Kantamneni, Sriharsha
Citation	Kantamneni S (2015) Cross-talk and regulation between glutamate and GABAB receptors. <i>Frontiers in Cellular Neuroscience</i> Apr 10th, vol.9, article135. doi: http://journal.frontiersin.org/article/10.3389/fncel.2015.00135/full
DOI	https://doi.org/10.3389/fncel.2015.00135
Rights	© 2016 The Author. First published by Frontiers Media. Reproduced in accordance with the publisher's self-archiving policy, under a Creative Commons attribution licence. http://creativecommons.org/licenses/by-nc-nd/4.0/
Download date	2026-03-05 10:40:12
Link to Item	http://hdl.handle.net/10454/8122

The University of Bradford Institutional Repository

<http://bradscholars.brad.ac.uk>

This work is made available online in accordance with publisher policies. Please refer to the repository record for this item and our Policy Document available from the repository home page for further information.

To see the final version of this work please visit the publisher's website. Available access to the published online version may require a subscription.

Link to publisher's version:

<http://journal.frontiersin.org/article/10.3389/fncel.2015.00135/full>

Citation: Kantamneni S (2015) Cross-talk and regulation between glutamate and GABAB receptors. *Frontiers in Cellular Neuroscience* Apr 10th, vol.9, article135. doi:
<http://journal.frontiersin.org/article/10.3389/fncel.2015.00135/full>

Copyright statement: © 2016 The Author. First published by Frontiers Media. Reproduced in accordance with the publisher's self-archiving policy, under a Creative Commons attribution licence.



Cross-talk and regulation between glutamate and GABA_B receptors

Sriharsha Kantamneni *

Bradford School of Pharmacy, School of Life Sciences, University of Bradford, Bradford, West Yorkshire, UK

Brain function depends on co-ordinated transmission of signals from both excitatory and inhibitory neurotransmitters acting upon target neurons. NMDA, AMPA and mGluR receptors are the major subclasses of glutamate receptors that are involved in excitatory transmission at synapses, mechanisms of activity dependent synaptic plasticity, brain development and many neurological diseases. In addition to canonical role of regulating presynaptic release and activating postsynaptic potassium channels, GABA_B receptors also regulate glutamate receptors. There is increasing evidence that metabotropic GABA_B receptors are now known to play an important role in modulating the excitability of circuits throughout the brain by directly influencing different types of postsynaptic glutamate receptors. Specifically, GABA_B receptors affect the expression, activity and signaling of glutamate receptors under physiological and pathological conditions. Conversely, NMDA receptor activity differentially regulates GABA_B receptor subunit expression, signaling and function. In this review I will describe how GABA_B receptor activity influence glutamate receptor function and *vice versa*. Such a modulation has widespread implications for the control of neurotransmission, calcium-dependent neuronal function, pain pathways and in various psychiatric and neurodegenerative diseases.

OPEN ACCESS

Edited by:

Milos Petrovic,
University of Belgrade, Serbia

Reviewed by:

Rostislav Turecek,
Academy of Sciences of Czech
Republic, Czech Republic
William Martin Connelly,
Cardiff University, UK

*Correspondence:

Sriharsha Kantamneni,
Bradford School of Pharmacy,
School of Life Sciences, University of
Bradford, Norcroft Building 2.12,
Richmond Road, Bradford,
West Yorkshire - BD7 1DP, UK
Tel: 0044 (0) 1274 236072
s.kantamneni@bradford.ac.uk

Received: 17 October 2014

Accepted: 23 March 2015

Published: 10 April 2015

Citation:

Kantamneni S (2015) Cross-talk and regulation between glutamate and GABA_B receptors. *Front. Cell. Neurosci.* 9:135. doi: 10.3389/fncel.2015.00135

Keywords: glutamate receptor, NMDAR, GABA_BR, AMPAR, AKAP, receptor regulation, receptor trafficking and mGluR

Introduction

Most excitatory signals that a neuron receives are mediated via glutamate receptors whereas most inhibitory signals are mediated via γ -aminobutyric acid (GABA) receptors (Cherubini et al., 1991; Hollmann and Heinemann, 1994). Many factors influence the regulation of excitatory and inhibitory synaptic inputs on a given neuron. One important factor is the subtype of neurotransmitter receptors present at not only the correct location to receive the appropriate signals but also their abundance at synapses (Dingledine et al., 1999; Sheng and Kim, 2011). Thus the molecular mechanisms that regulate receptor expression and localization at specific sites are of considerable importance. This review will describe the recent advances in our understanding of the molecular mechanisms underlying glutamate and GABA_B receptors cross-talk and discuss the roles of specific proteins that might control these processes.

Glutamate receptors are the major excitatory neurotransmitter receptors in the brain and play an important role in neural plasticity and development. Improper function of glutamate receptors is involved in various psychiatric and neurodegenerative diseases (Mattson, 2008; Musazzi et al., 2013). N-methyl-D-aspartate receptors (NMDARs), α -amino-3-hydroxy-5-methyl-4-isoxazolepropionate receptors (AMPA) and kainate receptors are

glutamate-gated ion channels, whereas metabotropic glutamate receptors (mGluRs) are G-protein-coupled receptors (GPCRs) that signal downstream via interaction with heterotrimeric G-proteins. Pharmacological and molecular biological studies have revealed that glutamate receptors exist as different subclasses, where receptor subtypes comprise multiple subunits such as NMDA receptors (GluN1 to GluN3), AMPA receptors (GluA1 to GluA4), kainate receptors (GluK1 to GluK5) and mGlu receptors (mGluR1 to mGluR8) (for reviews, see Nakanishi et al., 1998; Lodge, 2009; Nicoletti et al., 2011).

Conversely, GABA receptors are the primary proteins responsible for inhibitory responses in the brain. Metabotropic GABA receptors (GABA_BRs) are GPCRs that can mediate slow inhibitory neurotransmission in the CNS. GABA_BRs are located at both presynaptic and postsynaptic compartments and changes in their number, localization and activity affect the level of synaptic inhibition. Presynaptic GABA_BRs inhibit release of neurotransmitter by inhibiting Ca²⁺ channels (Wu and Saggau, 1995; Takahashi et al., 1998). Activation of postsynaptic GABA_BRs activates inwardly rectifying K⁺ channels (GIRK) to generate slow inhibitory postsynaptic potentials (reviewed in Marshall et al., 1999; Bowery et al., 2002; Gainetdinov et al., 2004). The GABA_BR is a heteromeric GPCR consisting of GABA_{B1} and GABA_{B2} subunits that exert much longer lasting synaptic inhibition compared to GABA_A ion channels (Marshall et al., 1999; Watanabe et al., 2002). The ligand-binding domain (Malitschek et al., 1999) is present in GABA_{B1} subunit and G-proteins interact with GABA_{B2} to regulate adenylate cyclase, GIRK channels and Ca²⁺ channels (Robbins et al., 2001). A large body of work over the last 20 years has demonstrated that GABA_B receptors are regulated via mechanisms distinct from those utilized by many classical GPCRs such as the β₂-adrenergic receptor (Bettler and Tiao, 2006). For example, following agonist exposure most GPCRs are phosphorylated and endocytosed from the cell surface into intracellular compartments and then either down-regulated via lysosomal or proteasomal degradation or recycled back to the cell surface following agonist removal. In contrast, cell surface GABA_B receptor levels are not significantly altered upon receptor stimulation in cultured cortical and hippocampal neurons (Fairfax et al., 2004; Bettler and Tiao, 2006). GABA_B receptors are very stable at the plasma membrane even after agonist exposure with little internalization in cultured neurons. The absence of receptor endocytosis correlates with lack of arrestin recruitment and agonist-induced phosphorylation (Fairfax et al., 2004). Surprisingly, increased phosphorylation at serine 892 in GABA_{B2} subunit decreased degradation rates and stabilizes surface GABA_BRs in neurons (Couve et al., 2004; Fairfax et al., 2004).

The main regulatory sites on both glutamate receptors and GABA_BRs are their intracellular C-terminal tails. Depending on the activity or stimulation received by the receptors, the C-terminal domains bind to various proteins including enzymes, scaffolds, and trafficking and signaling proteins (De La Rue and Henley, 2002). These sites sometimes also mediate complex formation during a cross-talk between the receptors. Many immunocytochemical and electron microscopy studies have

demonstrated that glutamatergic synapses are enriched with GABA_BRs (Fritschy et al., 1999; Luján and Shigemoto, 2006). There is also increasing evidence that NMDARs, AMPARs and mGluRs are modulated directly and sometimes indirectly by GABA_BRs (Morrisett et al., 1991; Hirono et al., 2001; Otmakhova and Lisman, 2004; Tabata et al., 2004; Sun et al., 2006; Chalifoux and Carter, 2010; Gandal et al., 2012; Terunuma et al., 2014). Conversely, GABA_BR subunits are differentially regulated by glutamate receptor subtypes under various stimulation protocols (Vargas et al., 2008; Cimarosti et al., 2009; Guetg et al., 2010; Maier et al., 2010; Terunuma et al., 2010; Kantamneni et al., 2014). The sections below in this review will follow this theme of regulation or modulation between GABA_B and glutamate receptors. This cross-talk provides important regulatory mechanisms, for example, in altering presynaptic release or changes to membrane potential, but also alters the function of glutamate receptors, which may prove useful in a therapeutic context.

GABA_BR-Mediated Regulation of Glutamate Receptor Function

GABA_BR Regulation of NMDAR-Dependent Post-Synaptic Calcium Signals

The major synaptic Ca²⁺ signals in the brain are mediated via NMDARs, which are crucial for activity-dependent changes in synaptic plasticity (Bliss and Collingridge, 1993; Mainen et al., 1999; Malenka and Bear, 2004). These Ca²⁺ signals are thought to be inhibited by GABA_B receptors via modulation of K⁺ channels, resulting in a hyperpolarization that decreases the Ca²⁺ influx and overall current by enhancing Mg²⁺ blockade of NMDARs. (Morrisett et al., 1991; Otmakhova and Lisman, 2004; Deng et al., 2009). Interestingly, it has also been demonstrated recently that Ca²⁺ influx via NMDARs is inhibited by GABA_B receptor activation (Chalifoux and Carter, 2010). This effect on NMDARs is independent of K⁺ channel and voltage sensitive Ca²⁺ channel activation, Gβγ subunits and internal Ca²⁺ stores. Via coupling to G_{α_i}/G_{α_o} G proteins, GABA_BRs inhibit adenylate cyclase to reduce PKA activity by decreasing cAMP levels. The Ca²⁺ influx via NMDA receptors is normally increased by PKA activity and reduction of PKA activity by GABA_BRs inhibits Ca²⁺ signals (Chalifoux and Carter, 2010). GABA_BR-mediated postsynaptic modulation through the PKA pathway does not affect synaptic currents mediated by NMDA or AMPA receptors (Chalifoux and Carter, 2010). As outlined below, protein kinases such as PKA and phosphatases such as PP1/2 and calcineurin (CaN) are regulated via AKAPs (A Kinase Anchoring Proteins) and mediate signaling where they act as scaffold molecules (see below for further insights).

NMDAR and GABA_BR Cross-Talk in Disease

Recently it has been demonstrated that, there is clear interplay between GABA_B and NMDA receptors not only in physiological functions but also in pathological situations. Altered NMDAR activity is observed in models of pain and neuropsychiatric disorders, but an interesting phenomenon is that these phenotypes can be rescued with GABA_BR ligands. For example,

in diabetic neuropathy, NMDAR expression is increased in spinal cord dorsal horn, while GABA_B receptors are down regulated at protein level (Wang et al., 2011). Using streptozotocin (STZ)-induced diabetic neuropathy rat models (STZ), it has been found that intrathecal injection of the GABA_BR agonist baclofen significantly increased paw withdrawal threshold. This effect was blocked with pre-treatment of CGP55845, a GABA_BR—selective antagonist (Bai et al., 2014). In STZ rats, changes in expression were observed in both cyclic AMP response element-binding protein (CREB) and GluN2B, which were significantly increased at the protein (CREB and GluN2B) and mRNA level (GluN2B) in spinal cord. The higher expression levels of both GluN2B and phosphorylated CREB proteins were significantly reduced by administration of baclofen (Liu et al., 2014). Importantly, baclofen-induced reduction of GluN2B and CREB expression was abolished when CGP55845 was pre-administered, suggesting that GABA_BR activation in the spinal cord dorsal horn can normalize NMDAR expression levels in diabetic neuropathic pain (Wang et al., 2011; Bai et al., 2014; Liu et al., 2014).

In contrast, reduced NMDA receptor functionality has been observed in neuropsychiatric disorders like intellectual disability, autism and schizophrenia (Gonda, 2012). For example, a mouse model expressing a reduced amount of GluN1 subunit (NR1^{neo-/-} mice) was characterized to mimic schizophrenic-like behavior (Mohn et al., 1999). These mice have increased power in the gamma (30–80 Hz) EEG range during rest, but show a reduced auditory-stimulus evoked gamma power (reduced gamma signal-to-noise), causing changes in excitatory/inhibitory balance, and express treatment resistant symptoms of autism and schizophrenia (Gandal et al., 2012). Treating NR1^{neo-/-} mice with baclofen restored excitatory/inhibitory balance, neural synchrony and also improved social function and spatial memory deficits (Gandal et al., 2012). To summarize, diseases characterized by NMDA receptor dysfunction, have the additional possibility of using GABA_B receptors as an appropriate target for therapy that could possibly pave the way to restore abnormalities in many other neurological diseases.

GABA_BR Cross-Talk with AMPARs

Surface expression of AMPA receptors was increased in a knock-in mouse model in which wild-type GABA_{B2}R was replaced with a S783A-mutated version which cannot be phosphorylated (Terunuma et al., 2014). The S783 on GABA_{B2} subunit is phosphorylated by AMP-dependent protein kinase (AMPK), which in-turn enhances receptor coupling to GIRKs (Kuramoto et al., 2007). Activating NMDARs transiently results in increased phosphorylation whereas prolonged activation results in dephosphorylation of GABA_BRs by protein phosphatase 2A (PP2A). GABA_BRs stability at cell surface is due to high constitutive phosphorylation of GABA_{B2}R and dephosphorylation of this subunit selectively targets the receptors for lysosomal degradation (Fairfax et al., 2004; Terunuma et al., 2010). The expression of GABA_BR was increased with the mutation due to reduced degradation, leading to decreased level of Arc/Arg3.1 protein necessary for memory consolidation. This, in turn, increased the number of

excitatory synapses, PSD95 protein expression and cell surface AMPA receptors. This cross-talk demonstrates a crucial role for GABA_BRs in regulating excitatory synaptic transmission and neuronal architecture (Terunuma et al., 2014).

GABA_BR Cross-Talk with mGluRs

Long-term depression (LTD) at cerebellar parallel fiber Purkinje cell synapses is a form of synaptic plasticity critical for cerebellar motor learning and requires the activation of the metabotropic glutamate receptor mGluR1 (Ichise et al., 2000; Ito, 2001). GABA_BRs are concentrated at cerebellar parallel fiber Purkinje cell synapses and have many functions that are both dependent and independent of GABA. GABA_BRs and mGluR1 are highly co-expressed in cerebellar Purkinje cells, and display very similar subcellular localizations throughout development (Ige et al., 2000; Luján and Shigemoto, 2006; Rives et al., 2009). Electrophysiological studies have shown that at Purkinje cell synapses, GABA_BR activation inhibits neurotransmitter release by inhibiting calcium channels as well as affecting release processes (Dittman and Regehr, 1996, 1997; Vigot and Batini, 1997). Extracellular Ca²⁺ interacts with GABA_BR in cerebellar Purkinje cells, leading to an increase in the glutamate sensitivity of mGluR1. This sensitization of mGluR1 to glutamate is specifically mediated by GABA_BRs as it is absent in cells from GABA_{B1}^{-/-} animals. It has also been shown that both GPCRs form a complex in cerebellum and that extracellular Ca²⁺-mediated crosstalk is not mediated via G_{i/o} proteins (Tabata et al., 2004). Activity-dependent GABA_BR inhibition by selective antagonists reduces the magnitude of LTD at parallel fiber Purkinje cell synapses (Kamikubo et al., 2007; Rives et al., 2009). In summary GABA_BRs not only mediate classical synaptic GABAergic neurotransmission but also regulate mGluR signaling and cerebellar synaptic plasticity.

NMDAR-Mediated Regulation of GABA_BR Function

GABA_BRs are very stable at cell surface in terms of agonist stimulation and the number of cell surface GABA_BRs is primarily controlled by glutamate and not GABA in central neurons (Fairfax et al., 2004; Vargas et al., 2008). Sustained application of glutamate leads to GABA_BR endocytosis, trafficking to lysosomes and subsequent degradation, resulting in a decrease in receptor expression at the cell membrane (Vargas et al., 2008; Maier et al., 2010). Further dissection of the effect of glutamate indicated that activation of AMPA and NMDA receptors is required for the down-regulation of GABA_BRs and that this effect is enhanced by activation of the group I mGluRs (mGlu1 and mGlu5) (Maier et al., 2010). Activation of NMDARs alone leads to down-regulation and degradation of GABA_{B1} and GABA_{B2} subunits, thereby reducing cell surface expression (Guetg et al., 2010; Terunuma et al., 2010; Kantamneni et al., 2014). Mechanistically, NMDAR activation triggers GABA_{B1} subunit phosphorylation on Ser867 by CaMKII, causing a CaMKII-dependent down regulation (Guetg et al., 2010). In both hippocampal and cortical cultured neurons NMDAR activation also alters the

phosphorylation state of GABA_{B2} subunit on Ser783, resulting in endocytosis and lysosomal degradation of the receptor complex (Terunuma et al., 2010). The GABA_{B2} subunit is also rapidly phosphorylated by AMPK upon NMDAR activation. Prolonged NMDAR activation subsequently results in GABA_{B2} subunit dephosphorylation by PP2A, which decreases the number of cell surface receptors (Terunuma et al., 2010).

Recently it has been shown that selective activation of synaptic NMDARs using chemically induced LTP (long-term potentiation) protocol (chem-LTP) leads to an increase in surface GABA_B receptors (Kantamneni et al., 2014). In the chem-LTP protocol, glycine (along with strychnine and bicuculline—to block glycine and GABA_A receptors, respectively) was used to specifically activate synaptic NMDARs, leading to significant increase in surface expression of AMPARs (Lu et al., 2001; Park et al., 2004). Prolonged activation of extrasynaptic NMDARs promotes cell death, whereas activation of synaptic NMDARs mediates synaptic plasticity and is thought to be involved in neuroprotection via modulation of nuclear Ca²⁺ signaling (Hardingham and Bading, 2010). Using the chem-LTP method, both GABA_{B1} and GABA_{B2} receptor subunit expression on the cell surface were increased in cultured rat hippocampal neurons due to enhanced receptor recycling from intracellular pools (Kantamneni et al., 2014).

GABA_BR subunits are differentially regulated under oxygen/glucose deprivation (OGD) conditions, which stimulates release of excess glutamate resulting in excitotoxic activation of NMDARs (Papadia and Hardingham, 2007; Cimarosti et al., 2009; Kantamneni et al., 2014). After OGD, expression of GABA_{B1} subunits at the cell surface is increased via enhanced recycling, while total cellular and cell surface expression levels of GABA_{B2} subunits are decreased due to reduced recycling (Cimarosti et al., 2009; Kantamneni et al., 2014; Maier et al., 2014). Removing GABA_{B2} subunit will decrease the number of functional GABA_BRs, as both subunits are required for normal signaling. In conclusion, the above findings demonstrate that the expression and regulation of GABA_BR subunits are dynamically regulated in response to synaptic and prolonged/global stimulation of NMDARs. Moreover, NMDAR regulation of GABA_BRs may be important under conditions of neurological disease, such as epilepsy or ischemia.

Anchoring and Scaffold Proteins as Possible Mediators of GABA/Glutamate Receptor Cross-Talk

Both GABAergic and glutamatergic receptor complexes are regulated and orchestrated by anchoring and scaffold proteins, which are increasingly being implicated in the cross-talk between the two systems. Components of receptor signalosome are typically localized together via scaffold proteins, which co-assemble receptors with regulatory proteins such as protein kinases and phosphatases. AKAPs are typical examples of this class of scaffold proteins (Wong and Scott, 2004). For example, AKAP5 (or AKAP79/150) is thought to localize PKA, protein kinase C (PKC) and the calmodulin-activated protein

phosphatase calcineurin (CaN) at specific synaptic sites to regulate excitatory synaptic strength (Gomez et al., 2002; Smith et al., 2006; Robertson et al., 2009; Jurado et al., 2010). AKAP5 is linked to NMDARs via PSD-95 (Colledge et al., 2000). AKAP5 is known to be a master scaffolding protein that links many proteins including kinases, phosphatases, cadherins, F-actin, MAGUKs and PIP₂ together with ion channels and receptors to regulate activity dependent signaling processes at synapses (Tunquist et al., 2008; Sanderson and Dell'Acqua, 2011). Many of the proteins binding to AKAP5 (such as PKA, PP2B) also regulate GABA_BRs and perhaps there is possibility that AKAP5 scaffolding function may be required for glutamate/GABA receptors cross-talk.

Yotiao is another AKAP protein derived from alternative splicing of AKAP9 (also known as AKAP350/450) and plays a major role in regulating NMDARs. Yotiao was first identified as a binding partner of the GluN1 subunit and later found to be an AKAP via its ability to bind PKA-RII subunits *in vitro* (Lin et al., 1998; Westphal et al., 1999). Yotiao binds both protein phosphatase 1 (PP1) and PKA to form a phosphatase-kinase signaling complex with the GluN1A receptor splice variant. The Yotiao-PP1-PKA complex functions as dual switch, in that activation of anchored PKA enhances NMDAR currents while activation of PP1 exerts an inhibitory effect on NMDAR activity (Westphal et al., 1999; Colledge et al., 2000).

GABA_{B1}Rs were previously shown to interact with a scaffold protein, GISP that enhances cell surface expression of heteromeric complex GABA_{B1}/GABA_{B2} (Kantamneni et al., 2007). GISP is an AKAP9 C-terminal splice variant with more than 90% similarity to AKAP9 but lacking any RII domain, which are PKA binding sites (Kantamneni et al., 2007). As mentioned previously, the NMDAR binding protein Yotiao is also an AKAP9 splice variant, but within the N-terminal region. Therefore, theoretically, AKAP9 could interact simultaneously with NMDARs and GABA_BRs as well as regulatory protein kinases and phosphatases. Thus, while speculative, it is tempting to suggest that AKAP9 functions to assemble the signaling complex responsible for mediating the observed cross-talk between the NMDARs and GABA_BRs. From expression studies it is known that AKAP9 is expressed in the brain and localized to synapses (Collado-Hilly and Coquil, 2009). In similarity to the AKAP5-CaN_PP2B-PKA complex, the AKAP9-PKA-PP1 complex might exist as one large macromolecular complex held together with receptor proteins such as GABA_BRs and NMDARs. At least in yeast-two hybrid assay it has been confirmed that GISP does not interact with NMDAR sub-type 1 (Kantamneni et al., 2007). GISP binding to other subtypes of NMDARs or Yotiao binding to GABA_BRs has not been tested, and that this warrants further work. Another protein that may potentially mediate direct crosstalk between GABA_BR signaling and glutamate receptor signaling is CaMKII. CaMKII is a Ca²⁺ calmodulin dependent protein kinase, previously been shown to interact with both GABA_B and NMDA receptors and regulate NMDAR mediated plasticity (Bayer et al., 2001; Guetg et al., 2010; El Gaamouch et al., 2012). Unlike the earlier examples of indirect receptor modulation, AKAPs and other signaling molecules like CaMKII potentially function as direct links between glutamate

and GABA_B receptors. If further characterized these complexes may eventually serve as potential drug targets.

Conclusions

Taken together we can conclude that there is very tight regulation between glutamate and GABA_B receptors. Regulation of NMDAR-mediated synaptic signals by GABA_BRs comprises a powerful mechanism for controlling the major excitatory systems in brain. Conversely, NMDAR-mediated control of GABA_BRs is clearly an important emerging concept in dictating the balance of excitability in the brain. Studying the trafficking and signaling pathways utilized by these excitatory and inhibitory receptors in an integrated manner will undoubtedly provide more understanding of these critical regulatory mechanisms and will ultimately shed light on how the balance between

excitatory and inhibitory neurotransmission is dictated in the brain. While many examples of interactions between glutamate and GABA_B receptors have been discovered, importantly, the molecular players involved in mediating this cross-talk are only just beginning to be discovered. With this in mind, investigation of the potential players in these processes, such as the AKAPs, is an exciting future avenue of study. Ultimately, targeting these specific regulatory pathways may form the basis of new therapies to treat a number of neurological disorders that are characterized by aberrant balance between excitatory and inhibitory neurotransmitter systems in the brain.

Acknowledgments

I thank Dr. Kevin Wilkinson, Dr. Daniel Rocca, Prof. Tim Palmer and Dr. Sonia Correa, for critical comments on manuscript.

References

- Bai, H., Liu, P., Wu, Y., Guo, W., Guo, Y., and Wang, X. (2014). Activation of spinal GABA_B receptors normalizes N-methyl-D-aspartate receptor in diabetic neuropathy. *J. Neurol. Sci.* 341, 68–72. doi: 10.1016/j.jns.2014.04.002
- Bayer, K., De Koninck, P., Leonard, A., Hell, J., and Schulman, H. (2001). Interaction with the NMDA receptor locks CaMKII in an active conformation. *Nature* 411, 801–805. doi: 10.1038/35081080
- Bettler, B., and Tiao, J. (2006). Molecular diversity, trafficking and subcellular localization of GABA_B receptors. *Pharmacol. Ther.* 110, 533–543. doi: 10.1016/j.pharmthera.2006.03.006
- Bliss, T., and Collingridge, G. (1993). A synaptic model of memory: long-term potentiation in the hippocampus. *Nature* 361, 31–39. doi: 10.1038/361031a0
- Bowery, N., Bettler, B., Froestl, W., Gallagher, J., Marshall, F., Raiteri, M., et al. (2002). International union of pharmacology. XXXIII. Mammalian γ -aminobutyric acid(B) receptors: structure and function. *Pharmacol. Rev.* 54, 247–264. doi: 10.1124/pr.54.2.247
- Chalifoux, J., and Carter, A. (2010). GABA_B receptors modulate NMDA receptor calcium signals in dendritic spines. *Neuron* 66, 101–113. doi: 10.1016/j.neuron.2010.03.012
- Cherubini, E., Giaresia, J., and Ben-Ari, Y. (1991). GABA: an excitatory transmitter in early postnatal life. *Trends Neurosci.* 14, 515–519. doi: 10.1016/0166-2236(91)90003-d
- Cimarosti, H., Kantamneni, S., and Henley, J. (2009). Ischaemia differentially regulates GABA(B) receptor subunits in organotypic hippocampal slice cultures. *Neuropharmacology* 56, 1088–1096. doi: 10.1016/j.neuropharm.2009.03.007
- Collado-Hilly, M., and Coquil, J. (2009). Ins(1,4,5)P₃ receptor type 1 associates with AKAP9 (AKAP450 variant) and protein kinase A type IIbeta in the Golgi apparatus in cerebellar granule cells. *Biol. Cell* 101, 469–480. doi: 10.1042/BC20080184
- Colledge, M., Dean, R., Scott, G., Langeberg, L., Haganir, R., and Scott, J. (2000). Targeting of PKA to glutamate receptors through a MAGUK-AKAP complex. *Neuron* 27, 107–119. doi: 10.1016/s0896-6273(00)00013-1
- Couve, A., Calver, A., Fairfax, B., Moss, S., and Pangalos, M. (2004). Unravelling the unusual signalling properties of the GABA(B) receptor. *Biochem. Pharmacol.* 68, 1527–1536. doi: 10.1016/j.bcp.2004.06.036
- De La Rue, S. A., and Henley, J. M. (2002). Proteins involved in the trafficking and functional synaptic expression of AMPA and KA receptors. *ScientificWorldJournal* 2, 461–482. doi: 10.1100/tsw.2002.97
- Deng, P.-Y., Xiao, Z., Yang, C., Rojanathammanee, L., Grisanti, L., Watt, J., et al. (2009). GABA(B) receptor activation inhibits neuronal excitability and spatial learning in the entorhinal cortex by activating TREK-2 K⁺ channels. *Neuron* 63, 230–243. doi: 10.1016/j.neuron.2009.06.022
- Dingledine, R., Borges, K., Bowie, D., and Traynelis, S. (1999). The glutamate receptor ion channels. *Pharmacol. Rev.* 51, 7–61.
- Dittman, J. S., and Regehr, W. G. (1996). Contributions of calcium-dependent and calcium-independent mechanisms to presynaptic inhibition at a cerebellar synapse. *J. Neurosci.* 16, 1623–1633.
- Dittman, J., and Regehr, W. (1997). Mechanism and kinetics of heterosynaptic depression at a cerebellar synapse. *J. Neurosci.* 17, 9048–9059.
- El Gaamouch, F., Buisson, A., Moustié, O., Lemieux, M., Labrecque, S., Bontempi, B., et al. (2012). Interaction between α CaMKII and GluN2B controls ERK-dependent plasticity. *J. Neurosci.* 32, 10767–10779. doi: 10.1523/JNEUROSCI.5622-11.2012
- Fairfax, B., Pitcher, J., Scott, M., Calver, A., Pangalos, M., Moss, S., et al. (2004). Phosphorylation and chronic agonist treatment atypically modulate GABA_B receptor cell surface stability. *J. Biol. Chem.* 279, 12565–12573. doi: 10.1074/jbc.m311389200
- Fritschy, J., Meskenaite, V., Weinmann, O., Honer, M., Benke, D., and Mohler, H. (1999). GABA_B-receptor splice variants GB1a and GB1b in rat brain: developmental, regulation, cellular distribution and extrasynaptic localization. *Eur. J. Neurosci.* 11, 761–768. doi: 10.1046/j.1460-9568.1999.00481.x
- Gainetdinov, R., Premont, R., Bohn, L., Lefkowitz, R., and Caron, M. (2004). Desensitization of G protein-coupled receptors and neuronal functions. *Annu. Rev. Neurosci.* 27, 107–144. doi: 10.1146/annurev.neuro.27.070203.144206
- Gandal, M., Sisti, J., Klook, K., Ortinski, P., Leitman, V., Liang, Y., et al. (2012). GABA_B-mediated rescue of altered excitatory-inhibitory balance, gamma synchrony and behavioral deficits following constitutive NMDAR-hypofunction. *Transl. Psychiatry* 2:e142. doi: 10.1038/tp.2012.69
- Gomez, L., Alam, S., Smith, K., Horne, E., and Dell'Acqua, M. (2002). Regulation of A-kinase anchoring protein 79/150-cAMP-dependent protein kinase postsynaptic targeting by NMDA receptor activation of calcineurin and remodeling of dendritic actin. *J. Neurosci.* 22, 7027–7044.
- Gonda, X. (2012). Basic pharmacology of NMDA receptors. *Curr. Pharm. Des.* 18, 1558–1567. doi: 10.2174/138161212799958521
- Guett, N., Abdel Aziz, S., Holbro, N., Turecek, R., Rose, T., Seddik, R., et al. (2010). NMDA receptor-dependent GABA_B receptor internalization via CaMKII phosphorylation of serine 867 in GABA_B1. *Proc. Natl. Acad. Sci. U S A* 107, 13924–13929. doi: 10.1073/pnas.1000909107
- Hardingham, G. E., and Bading, H. (2010). Synaptic versus extrasynaptic NMDA receptor signalling: implications for neurodegenerative disorders. *Nat. Rev. Neurosci.* 11, 682–696. doi: 10.1038/nrn2911
- Hirono, M., Yoshioka, T., and Konishi, S. (2001). GABA(B) receptor activation enhances mGluR-mediated responses at cerebellar excitatory synapses. *Nat. Neurosci.* 4, 1207–1216. doi: 10.1038/nn764
- Hollmann, M., and Heinemann, S. (1994). Cloned glutamate receptors. *Annu. Rev. Neurosci.* 17, 31–108. doi: 10.1146/annurev.neuro.17.1.31
- Ichise, T., Kano, M., Hashimoto, K., Yanagihara, D., Nakao, K., Shigemoto, R., et al. (2000). mGluR1 in cerebellar Purkinje cells essential for long-term depression, synapse elimination and motor coordination. *Science* 288, 1832–1835. doi: 10.1126/science.288.5472.1832

- Ige, A., Bolam, J., Billinton, A., White, J., Marshall, F., and Emson, P. (2000). Cellular and sub-cellular localisation of GABA(B1) and GABA(B2) receptor proteins in the rat cerebellum. *Brain Res. Mol. Brain Res.* 83, 72–80. doi: 10.1016/S0169-328X(00)00199-6
- Ito, M. (2001). Cerebellar long-term depression: characterization, signal transduction and functional roles. *Physiol. Rev.* 81, 1143–1195.
- Jurado, S., Biou, V., and Malenka, R. (2010). A calcineurin/AKAP complex is required for NMDA receptor-dependent long-term depression. *Nat. Neurosci.* 13, 1053–1055. doi: 10.1038/nn.2613
- Kamikubo, Y., Tabata, T., Kakizawa, S., Kawakami, D., Watanabe, M., Ogura, A., et al. (2007). Postsynaptic GABA_B receptor signalling enhances LTD in mouse cerebellar Purkinje cells. *J. Physiol.* 585, 549–563. doi: 10.1111/jphysiol.2007.141010
- Kantamneni, S., Corrêa, S., Hodgkinson, G., Meyer, G., Vinh, N., Henley, J., et al. (2007). GISP: a novel brain-specific protein that promotes surface expression and function of GABA(B) receptors. *J. Neurochem.* 100, 1003–1017. doi: 10.1111/j.1471-4159.2006.04271.x
- Kantamneni, S., González-González, I., Luo, J., Cimarosti, H., Jacobs, S., Jaafari, N., et al. (2014). Differential regulation of GABA_B receptor trafficking by different modes of N-methyl-D-aspartate (NMDA) receptor signaling. *J. Biol. Chem.* 289, 6681–6694. doi: 10.1074/jbc.M113.487348
- Kuramoto, N., Wilkins, M., Fairfax, B., Revilla-Sanchez, R., Terunuma, M., Tamaki, K., et al. (2007). Phospho-dependent functional modulation of GABA(B) receptors by the metabolic sensor AMP-dependent protein kinase. *Neuron* 53, 233–247. doi: 10.1016/j.neuron.2006.12.015
- Lin, J., Wyszynski, M., Madhavan, R., Sealock, R., Kim, J., and Sheng, M. (1998). Yotiao, a novel protein of neuromuscular junction and brain that interacts with specific splice variants of NMDA receptor subunit NR1. *J. Neurosci.* 18, 2017–2027.
- Liu, P., Guo, W., Zhao, X., Bai, H., Wang, Q., Wang, X., et al. (2014). Intrathecal baclofen, a GABA_B receptor agonist, inhibits the expression of p-CREB and NR2B in the spinal dorsal horn in rats with diabetic neuropathic pain. *Can. J. Physiol. Pharmacol.* 92, 655–660. doi: 10.1139/cjpp-2013-0463
- Lodge, D. (2009). The history of the pharmacology and cloning of ionotropic glutamate receptors and the development of idiosyncratic nomenclature. *Neuropharmacology* 56, 6–21. doi: 10.1016/j.neuropharm.2008.08.006
- Lu, W., Man, H., Ju, W., Trimble, W., MacDonald, J. F., and Wang, Y. T. (2001). Activation of synaptic NMDA receptors induces membrane insertion of new AMPA receptors and LTP in cultured hippocampal neurons. *Neuron* 29, 243–254. doi: 10.1016/S0896-6273(01)00194-5
- Luján, R., and Shigemoto, R. (2006). Localization of metabotropic GABA receptor subunits GABA_B1 and GABA_B2 relative to synaptic sites in the rat developing cerebellum. *Eur. J. Neurosci.* 23, 1479–1490. doi: 10.1111/j.1460-9568.2006.04669.x
- Maier, P., Marin, I., Grampp, T., Sommer, A., and Benke, D. (2010). Sustained glutamate receptor activation down-regulates GABA_B receptors by shifting the balance from recycling to lysosomal degradation. *J. Biol. Chem.* 285, 35606–35614. doi: 10.1074/jbc.M110.142406
- Maier, P., Zemoura, K., Acuña, M., Yévenes, G., Zeilhofer, H., and Benke, D. (2014). Ischemia-like oxygen and glucose deprivation mediates down-regulation of cell surface γ -aminobutyric acid_B receptors via the endoplasmic reticulum (ER) stress-induced transcription factor CCAAT/enhancer-binding protein (C/EBP)-homologous protein (CHOP). *J. Biol. Chem.* 289, 12896–12907. doi: 10.1074/jbc.M114.550517
- Mainen, Z., Malinow, R., and Svoboda, K. (1999). Synaptic calcium transients in single spines indicate that NMDA receptors are not saturated. *Nature* 399, 151–155. doi: 10.1038/20187
- Malenka, R. C., and Bear, M. F. (2004). LTP and LTD: an embarrassment of riches. *Neuron* 44, 5–21. doi: 10.1016/j.neuron.2004.09.012
- Malitschek, B., Schweizer, C., Keir, M., Heid, J., Froestl, W., Mosbacher, J., et al. (1999). The N-terminal domain of γ -aminobutyric Acid(B) receptors is sufficient to specify agonist and antagonist binding. *Mol. Pharmacol.* 56, 448–454.
- Marshall, F. H., Jones, K. A., Kaupmann, K., and Bettler, B. (1999). GABA_B receptors—the first 7TM heterodimers. *Trends Pharmacol. Sci.* 20, 396–399. doi: 10.1016/S0165-6147(99)01383-8
- Mattson, M. (2008). Glutamate and neurotrophic factors in neuronal plasticity and disease. *Ann. N Y Acad. Sci.* 1144, 97–112. doi: 10.1196/annals.1418.005
- Mohn, A., Gainetdinov, R., Caron, M., and Koller, B. (1999). Mice with reduced NMDA receptor expression display behaviours related to schizophrenia. *Cell* 98, 427–436. doi: 10.1016/S0092-8674(00)81972-8
- Morrisett, R., Mott, D., Lewis, D., Swartzwelder, H., and Wilson, W. (1991). GABA_B-receptor-mediated inhibition of the N-methyl-D-aspartate component of synaptic transmission in the rat hippocampus. *J. Neurosci.* 11, 203–209.
- Musazzi, L., Treccani, G., Mallei, A., and Popoli, M. (2013). The action of antidepressants on the glutamate system: regulation of glutamate release and glutamate receptors. *Biol. Psychiatry* 12, 1180–1188. doi: 10.1016/j.biopsych.2012.11.009
- Nakanishi, S., Nakajima, Y., Masu, M., Ueda, Y., Nakahara, K., Watanabe, D., et al. (1998). Glutamate receptors: brain function and signal transduction. *Brain Res. Brain Res. Rev.* 26, 230–235. doi: 10.1016/S0165-0173(97)00033-7
- Nicoletti, F., Bockaert, J., Collingridge, G., Conn, P., Ferraguti, F., Schoepp, D., et al. (2011). Metabotropic glutamate receptors: from the workbench to the bedside. *Neuropharmacology* 60, 1017–1041. doi: 10.1016/j.neuropharm.2010.10.022
- Otmakhova, N., and Lisman, J. (2004). Contribution of I_h and GABA_B to synaptically induced afterhyperpolarizations in CA1: a brake on the NMDA response. *J. Neurophysiol.* 92, 2027–2039. doi: 10.1152/jn.00427.2004
- Papadia, S., and Hardingham, G. E. (2007). The dichotomy of NMDA receptor signaling. *Neuroscientist* 13, 572–579. doi: 10.1177/1073858407305833
- Park, M., Penick, E., Edwards, J., Kauer, J., and Ehlers, M. (2004). Recycling endosomes supply AMPA receptors for LTP. *Science* 305, 1972–1975. doi: 10.1126/science.1102026
- Rives, M., Vol, C., Fukazawa, Y., Tinel, N., Trinquet, E., Ayoub, M., et al. (2009). Crosstalk between GABA_B and mGlu_{1a} receptors reveals new insight into GPCR signal integration. *EMBO J.* 28, 2195–2208. doi: 10.1038/emboj.2009.177
- Robbins, M., Calver, A., Filippov, A., Hirst, W., Russell, R., Wood, M., et al. (2001). GABA(B₂) is essential for g-protein coupling of the GABA(B) receptor heterodimer. *J. Neurosci.* 21, 8043–8052.
- Robertson, H., Gibson, E., Benke, T., and Dell'Acqua, M. (2009). Regulation of postsynaptic structure and function by an A-kinase anchoring protein-membrane-associated guanylate kinase scaffolding complex. *J. Neurosci.* 29, 7929–7943. doi: 10.1523/JNEUROSCI.6093-08.2009
- Sanderson, J. L., and Dell'Acqua, M. L. (2011). AKAP signaling complexes in regulation of excitatory synaptic plasticity. *Neuroscientist* 3, 321–336. doi: 10.1177/1073858410384740
- Sheng, M., and Kim, E. (2011). The postsynaptic organization of synapses. *Cold Spring Harb. Perspect. Biol.* 3:a005678. doi: 10.1101/cshperspect.a005678
- Smith, K., Gibson, E., and Dell'Acqua, M. (2006). cAMP-dependent protein kinase postsynaptic localization regulated by NMDA receptor activation through translocation of an A-kinase anchoring protein scaffold protein. *J. Neurosci.* 26, 2391–2402. doi: 10.1523/jneurosci.3092-05.2006
- Sun, H., Ma, C., Kelly, J., and Wu, S. (2006). GABA_B receptor-mediated presynaptic inhibition of glutamatergic transmission in the inferior colliculus. *Neurosci. Lett.* 399, 151–156. doi: 10.1016/j.neulet.2006.01.049
- Tabata, T., Araishi, K., Hashimoto, K., Hashimoto, Y., van der Putten, H., Bettler, B., et al. (2004). Ca²⁺ activity at GABA_B receptors constitutively promotes metabotropic glutamate signaling in the absence of GABA. *Proc. Natl. Acad. Sci. U S A* 101, 16952–16957. doi: 10.1073/pnas.0405387101
- Takahashi, T., Kajikawa, Y., and Tsujimoto, T. (1998). G-Protein-coupled modulation of presynaptic calcium currents and transmitter release by a GABA_B receptor. *J. Neurosci.* 18, 3138–3146.
- Terunuma, M., Revilla-Sanchez, R., Quadros, I., Deng, Q., Deeb, T., Lumb, M., et al. (2014). Postsynaptic GABA_B receptor activity regulates excitatory neuronal architecture and spatial memory. *J. Neurosci.* 34, 804–816. doi: 10.1523/JNEUROSCI.3320-13.2013
- Terunuma, M., Vargas, K., Wilkins, M., Ramírez, O., Jaureguiberry-Bravo, M., Pangalos, M., et al. (2010). Prolonged activation of NMDA receptors promotes dephosphorylation and alters postendocytic sorting of GABA_B receptors. *Proc. Natl. Acad. Sci. U S A* 107, 13918–13923. doi: 10.1073/pnas.1000853107
- Tunquist, B., Hoshi, N., Guire, E., Zhang, F., Mullendorff, K., Langeberg, L., et al. (2008). Loss of AKAP150 perturbs distinct neuronal processes in mice. *Proc. Natl. Acad. Sci. U S A* 105, 12557–12562. doi: 10.1073/pnas.0805922105

- Vargas, K., Terunuma, M., Tello, J., Pangalos, M., Moss, S., and Couve, A. (2008). The availability of surface GABA_B receptors is independent of gamma-aminobutyric acid but controlled by glutamate in central neurons. *J. Biol. Chem.* 283, 24641–24648. doi: 10.1074/jbc.M802419200
- Vigot, R., and Batini, C. (1997). GABA(B) receptor activation of Purkinje cells in cerebellar slices. *Neurosci. Res.* 29, 151–160. doi: 10.1016/s0168-0102(97)00087-4
- Wang, X., Zhang, Q., Zhang, Y., Liu, Y., Dong, R., Wang, Q., et al. (2011). Downregulation of GABA_B receptors in the spinal cord dorsal horn in diabetic neuropathy. *Neurosci. Lett.* 490, 112–115. doi: 10.1016/j.neulet.2010.12.038
- Watanabe, M., Maemura, K., Kanbara, K., Tamayama, T., and Hayasaki, H. (2002). GABA and GABA receptors in the central nervous system and other organs. *Int. Rev. Cytol.* 213, 1–47. doi: 10.1016/s0074-7696(02)13011-7
- Westphal, R., Tavalin, S., Lin, J., Alto, N., Fraser, I., and Langeberg, L. (1999). Regulation of NMDA receptors by an associated phosphatase-kinase signaling complex. *Science* 285, 93–96. doi: 10.1126/science.285.5424.93
- Wong, W., and Scott, J. (2004). AKAP signalling complexes: focal points in space and time. *Nat. Rev. Mol. Cell Biol.* 5, 959–970. doi: 10.1038/nrm1527
- Wu, L. G., and Saggau, P. (1995). GABA_B receptor-mediated presynaptic inhibition in guinea-pig hippocampus is caused by reduction of presynaptic Ca²⁺ influx. *J. Physiol.* 485, 649–657. doi: 10.1113/jphysiol.1995.sp020759

Conflict of Interest Statement: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Kantamneni. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution and reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.