The biology and genetics of curly hair

Experimental Dermatology ADF

The Biology and Genetics of Curly Hair

Journal:	*Experimental Dermatology*
Manuscript ID	Draft
Manuscript Type:	Review Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Westgate, Gillian; University of Bradford, Centre for Skin Sciences Ginger, Rebecca; Unilever R&D, Colworth Science Park Green, Martin; Unilever R&D, Colworth Science Park
Keywords:	human hair, curl, hair follicle, hair trait genetics

The Biology and Genetics of Curly Hair

Gillian E Westgate[1], Rebecca S Ginger[2] and Martin R Green[2]

[1]Centre for Skin Sciences, University of Bradford, Bradford, West Yorkshire, BD7 1DP, UK and [2]Unilever R&D Colworth Science Park, Sharnbrook, Bedfordshire, MK44 1LQ, UK

Hair fibers show wide diversity across and within all human populations suggesting that hair fiber form and its coloration has been subject to much adaptive pressure over many thousands of years. Human hair fibers typically have the same basic structure in all human hair types. However, the three dimensional shape of the entire fiber varies considerably depending on ethnicity and geography, with examples from very straight hair with no rotational turn about the long axis, to the tightly sprung coils of some African races. This review will introduce the reader to hair follicle formation, the hair growth cycle and basic hair follicle structure and will review the current understanding on how hair fibers are formed by follicles into a non-linear coiled form and which genetic and biological factors are thought to be responsible for hair shape. The creation of the highly complex biomaterials in hair fibre and follicle and how these confer mechanical functions on the fibre so formed is a topic that remains relatively unexplained thus far. We focus here on the links between genetics and protein expression and function in order to understand some of the molecular controls on formation of curly hair.

While it is true that across all mammals, the basic structure of the fiber is the same –a cuticle, cortex and medulla (in some)- how these structures are built by the hair follicle and shaped into the functional hair fiber for both an individual member of a species and the relevant body site, suggests that there is a level of 'fine control' on the process of hair fiber formation by the hair follicle.

The distribution of forms of curly hair is shown in Figure 1 and a closer inspection reveals that curly hair fibers are rarely a true coil but exhibit heterogeneity in the direction of the curl in all but the mildest cases. Curly hairs have an elliptical or 'D' shape in cross section. This enables bi-directional bending stiffness with fibres tending to bend most easily in the direction of the flattened axis. The relationship of the long and short diameter to the direction of hair growth also changes (unlike the eyelash where this relationship is maintained [1]. Therefore, at the level of the follicle, we need to understand how the arrangement of cells results in a fibre that is elliptical with the orientation of the ellipse changing with time during hair growth.

Hair fibres across all races and geographies show degrees of curl that are readily measurable [2-4]. Previous studies examining the classification of hair phenotypes reveal potentially important information about the biology of curly hair formation and the evolutionary and environmental drivers behind curly hair as a human trait. Both Hrdy and de la Mettrie [2, 5] studied various hair types sampled from countries and cultures across the world. The degree of curvature of a fiber in its natural state appears to account for most of the variation (87% [5]), which is as expected. The presence of a medulla is chiefly correlated with hair diameter. However 'twist' (as defined by the sudden natural constrictions in the fibre that produce a discontinuity in curvature and curvature variability); 'crimp' (change in direction of curvature); 'wave' – (the number of oscillations/coils per unit length) and 'kink' – (a sharp twist or bend) are also important in differentiating ethnicity and maybe also helpful in defining the genetic and functional origins of curly hair. Hrdy 1973 [2] showed that kinking and crimp was not always correlated with curvatures, and irregular curvature caused by kinking separated a population in the Solomon Islands (Melanesian) from African. Interestingly, the adaptation of highly curled hair separately in these two very distant populations achieved the same functional attribute of an intensely curled hair, suggesting the result of environmental pressures related to evaporation of sweat and scalp cooling.

The hair follicle is a self-sufficient and highly organized structure within the skin that has both proliferating (dividing) and differentiating (functional/specialized) cell compartments. The hair follicle comprises cells of epithelial, mesenchymal and neuronal (melanocyte) origin and is intimately connected to the surrounding dermis through blood and nerve supply and the interchange of individual cells associated with the follicle, including cells from the immune system such as mast cells. The hair follicle is an autonomous mini-organ in the skin, thus when considering how hair shape is controlled by the hair follicle, we must consider what is know about the first embryonic hair follicles, the diversity of hair shape within and between individuals and then drill down to investigate how the component parts of the follicle are arranged in order to make a fiber with low, moderate or high curl. Because the shapes of cells in the developing hair shaft are grossly altered during differentiation of the newly forming hair, it is also necessary to consider some of the biomechanical aspects that govern hair fiber shape. A global study of hair shape variability and racial classification reported no gender-based differences [5], suggesting that sex hormone influences are minimal in hair shape and curl determination.

Follicle anatomy, structure, size and relationship to hair shape

Hair follicles have a multi-layered structure with seven layers of specialized epithelial cells arranged in a concentric pattern (like an onion), with the hair fiber at the centre, (Figure 2). These concentric layers of epithelial cells all have unique differentiation pathways and properties. Most cells of the follicle are epithelial, however, a group of mesenchymal (fibroblast-like) cells lie right inside the lower follicle bulb called the dermal papilla (DP) and is continuous with the very outer layer of the follicle, the connective tissue sheath. The dermal papilla plays an essential role in directing the regulation of hair growth and the hair cycle.

In terms of the formation of the hair fiber inside the curly hair follicle, it is useful to consider activity in two compartments:-

i). *The mitotic region*, where cells of the lower bulb are undergoing rapid cell division and generating the 'force' behind hair formation. In curly hair the mitotic zone can be imagined as being organized in an asymmetrical arrangement around the dermal papilla. Studies in mice on zigzag hairs reveals a relationship between an asymmetric location of dermal papilla cells which results in the change in direction of hair growth generating the zig-zag [6]. More recent studies show that curved hair follicles emerge from wool follicles with asymmetric distribution of mitotic cells [7]. In human curly hair follicles there is some asymmetry in the proliferating pool of cells and this is described later.

ii). *The zone of differentiation*, where cells in the follicle inner root sheath and hair fiber become fully keratinized and confer rigidity to these structures. Both the mitotic 'force' and the subsequent hardening of the fiber and root sheath cells are considered important factors in establishing ultimate fiber shape, as described below.

In order to envision this arrangement of cells, it is helpful to view the hair follicle from a three dimensional viewpoint (Figure 3), in which the relationship of the fibre growth axis to the orientation of the dermal papilla is depicted. In Figure 3a there is full symmetry around the long axis of the follicle in a straight hair. In Figure 3b the axis is symmetrical through the bisected dermal papilla, however this does not match the long axis of hair growth and in Figure 3c the curved nature of the lower hair follicle bulb is shown as out of plane with the long axis of hair growth. The principle is now established that a curly follicle makes a curly hair [8] [9] and that some form of asymmetry in the follicle drives the formation of the coiled/curly hair. It has been shown in several studies that the shape and size of the follicle determines the shape and size of the hair and that curved/bent follicles produce curly hair fibers in all ethnicities [8-12] [13]. Thus while it is perhaps correct to assume that hair shape is defined in the follicle, the considered question for the hair biologist and biophysicist is how the follicle shape and associated cell distribution can set up a fixed or variable curl phenotype and generate features such as crimp (change in curl direction) and kink (discontinuity in curl) and also why these features may have been usefully inherited?

Hair follicle development and the hair cycle

Given that hair follicle structure is basically the same in human populations, we can propose that the development of the follicle structure is also very similar, even though this is poorly studied in embryonic tissues for reasons of ethics. The hair follicle forms as an organized involution of the epidermis during the early weeks of gestation in human scalp [14]. Firstly a hair placode forms that responds to the signals derived from the mesenchymal (dermal) cells immediately below the placode, which will eventually become the dermal papilla [15]. The epithelial cells expand in number and the epithelial hair peg extends into the dermis and cells of the newly forming hair matrix and upper hair canal start to differentiate [16]. Melanocytes, which are derived from the neuroectoderm (neuronal tissues of the embryo), arrive in the follicle during embryogenesis to provide hair with a source of melanin and so its colour. It is assumed that this process is identical for the formation of curly hair; however, this has not been studied in any detail. Children of African Negroid descent are born with loose 'silky' curls and that they may not attain the tight curls for another 12 months or so [17]. This suggests that the first hair cycle, or possibly the embryonic 'lanugo' hair has a different shape to subsequent hair cycles. The first hair cycle can be considered as somewhat different to subsequent hair cycles, given that it is governed by embryonic development processes, which are not all required in the post-natal hair cycles. Lanugo hair is normally shed very early in a child's life, much of it inter-uterine. Interestingly, when interrogating the internet for information on this topic, most African American or mixed race babies are reported to have rather straight hair at birth which curls a little more when wet and is replaced by very curly hair over the coming months and early years.

The cyclic behavior of hair follicles as the regulation of the hair cycle has led to an impressive amount of research into the molecular factors responsible for hair growth. Furthermore, because hair shape is reasonably fixed parameter in the adult (save for exogenous influences), the factors that control the re-growth of hair in each hair cycle must also maintain the shape characteristics of each new hair follicle formed during a lifetime. We understand that the hair follicle cycle

retains an element of developmental dynamics reflected in the interactions between the mesenchymal and epithelial elements as originally proposed by Sun et al [18] and Hardy (Hardy 1992) so each time a hair follicle goes through this 're-birth' in the hair cycle, all the factors that govern curl have to be re-established. The adult hair cycle has a growth phase (anagen) which also encompasses the very early stages of follicle re-formation, recently termed 'neogen' [19], a regression phase where the lower two thirds of the follicle undergoes programmed deletion (catagen) [20-22] and the resting and shedding phases telogen and exogen. [23] [24] [25]. These events occur in all hair-bearing species and the molecular dynamics of progression through the hair cycle has been the subject of much study [19, 26-29]. The factors controlling the progression through the cycle include genetic and epigenetic factors [30], [31] and the so-called hair follicle clock [32-34]. Anagen is maintained by growth factors such as VEGF and IGF1[35-37], which help maintain cell proliferation in the bulb matrix over several years in the case of scalp hair, but only just a few weeks in the case of eyebrow and eyelash. The signaling pathways that have been shown to be important in early anagen include Wnt/beta-catenin, bone morphogenic proteins (BMPs) and Sonic hedgehog (Shh) pathways and are all involved in hair follicle initiation in embryogenesis [38, 39] [40] and are governed by Hairless (HR) protein, the product of the hairless gene (*HR*) [41]. This strongly suggests that the program for hair shape is present in the hair follicle stem/progenitor cells and these may also govern follicle variation in size and function across the body. It is believed that the size of the hair follicle and subsequent fiber diameter is determined by the size (number of cells), and especially the maximum width of the dermal papilla [42] [43]. However the factors affecting the relationship between follicle size (fiber diameter), anagen duration (hair length), and curl are not, so far, understood.

We know that the follicle of curly hairs is also curved, but in two directions – retro-curvature, suggesting curl is set in the follicle [8, 9]. Key questions on formation of curly hair include 'what factors control the shape of the follicle?' 'what is the driving force for asymmetry?' does the follicle bend because of pre-

determined strain placed within the fibre owing to asymmetrical fixed protein structure, or, does the asymmetrical protein distribution arise because the follicle is curved? The retro-curved nature of the hair follicle in African scalp skin is shown in Figure 4 where both the bulb and the hair shaft shows curvature in the skin as indicated by the double bisection of the hair shaft (Figure 4b).

Studies (described in more detail below) suggest five sources of molecular control in conferring fibre shape:- 1) asymmetric expression of structural keratins in the pre-cortex; 2) variable cortical cell shape and keratin filament orientation in relation to the axis of hair growth; 3) asymmetric rates of proliferation in cells forming the inner and outer root sheaths, 4) polymorphic variation in the proteins of the IRS which (presumably) alters its ability to form a specific shape in the keratinizing zone of the hair shaft and 5) the asymmetry of the dermal papilla within the central 'core' of the hair bulb. The translation of these molecular 'settings' into curl also requires appreciation of biomechanics as the curvature of the fibre inside the follicle during growth and then outside when released, is very different.

Some of the most instructive studies on molecular factors for curl have come from examining the expression of a range of proteins in the asymmetric compartments of the curly hair follicle in relation to the structure and orientation of cells in the cortex. [1, 9, 12, 44]. These are summarised in Table1. Notable examples are hair keratin Ha8/K38 which is expressed earlier on the concave side of the follicle [12], insulin like growth factor binding protein 5 (IGFBP5) which shows elevated expression in the convex side of the outer root sheath (ORS) in curly hair follicles [44], keratin 71 which is only expressed in the inner root sheath, but when polymorphic leads to woolly hair syndromes, and the relationship between the cells of the DP and the bulb matrix. Ki67, which marks the proliferating compartment, shows asymmetric expression around the dermal papilla, [1].

So, how can compartmentalized expression of different proteins and protein functions influence curl? There are five possible mechanisms that are introduced below and will be referred to when discussing the genetic and developmental

origins of variation in hair fibre shape and curl identified through whole genomic screening studies of the curly hair trait.

1) Asymmetric expression of structural keratins in the pre-cortex.

The cortex of the hair fibre is most likely to structurally support curly hair characteristics (Figure 2). Thus, we would expect variation in the expression of the cortical keratins and keratin associate proteins. hHa8, K38 (gene KRT38) is a Type I acidic member of the hair keratins [45] and the only member of the complement of cortical hair keratins described as being asymmetrically distributed in the curly hair follicle [46]. There is no further research on the regulation of hHa8/K38 expression; this will be needed to help understand why this particular type I keratin associates with curly hair through its uneven expression. The cuticle keratin K82 (hHb1), was also shown to be expressed slightly later on the convex side of the follicle [9]. It is not yet known what regulates this differential expression pattern.

2) The cortex comprises three different types of cell (as judged microscopically); para-, ortho- and meso-cortex. Within the cortical cells, keratin filaments are formed in dense, almost crystalline arrays. Cortical cells are very long and aligned with the long axis of the hair. The main variation within these cells is the orientation of the long axis of the keratin filaments in relation to the long axis of the hair fibre and the ratio of keratin to keratin-associated-proteins. The distribution of these different cell types within the hair fibre cortex has been studied in wool follicles where a distinct microscopical arrangement into ortho- and para-cortex was originally thought to be associated with crimp [47], however, more recent studies looking at the orientation of the keratin filaments in human hair have failed to find such a relationship [48]. In human hair fibers, three different cortical cell types have been observed and it has been proposed that the distribution in the different cell types may be related to curl [11, 46].

3) Asymmetric proliferation in cells forming the inner and outer root sheaths.

Limited data exists to understand the role of asymmetric proliferation rates but it is likely that this is linked to the asymmetry in the shape and activity of the DP.

Ki67, a marker of proliferating cells, is distributed in an asymmetric pattern in both curly hair follicles and human eyelash follicles [1, 46]. The proliferating compartment extended higher up in the bulb on the convex side of the hair follicle. IFGBP5 is involved in the action of the growth factor IGF1 that is known to be required for hair growth [22, 35, 49] and the increased expression of IGFBP5 on the convex side of the follicle was also shown to impart asymmetric hair growth. This suggests that asymmetrical growth rate of cells forming the hair cortex influence curl degree. It should be considered whether the proliferating zone itself is mobile about the long axis of the follicle generating some form of curl force?

4) Inner Root Sheath links to variation on fibre shape

Perhaps the most compelling evidence for control on hair fibre shape supports the role of the IRS. Genetic studies (further described below) and hair diseases that give rise to Woolly hair, have revealed polymorphic variation in several proteins of the IRS and this is presumed to alter its ability to form specific shapes in the keratinizing zone of the hair shaft. The importance of the IRS was demonstrated by the namesakes for the two key layered structures; Jacob Henle and Thomas Huxley [50] and much molecular genetic evidence is emerging to support this view with several IRS protein polymorphisms associated with curliness. The inner root sheath in mammals is comprised of three layers; the cuticle that directly abuts the hair shaft cuticle; Huxley's layer and Henle's layer (Birbeck and Mercer 1957) Figure 2. Studies on monotremes reveal a simpler structure without a distinguishable Henle layer which could be the forbear of the more complex mammalian IRS and give a clue to how hairs arose from reptilian scales during evolution [51][52]. Henle's layer keratins are the first to fully keratinise or 'harden' in the follicle to support hair shape. Cells in Huxley's layer produce keratins that interact with the protein trichohyalin, the latter also specific to Huxleys layer. The cells of Huxley's layer are fully differentiated more distal to the bulb than Henle's layer and the interaction between these two layers forms the bulk of the IRS. Trichohyalin expression is only found where a hardened keratin structure is needed, such as hair, nail and the filiform papillae

on the tongue [53-56]. The interaction of trichohyalin with keratin is preceded by the enzymatic conversion of arginine to citrulline within the trichohyalin by the enzyme peptidyl arginine deiminase (PAD) that reduces its overall charge so facilitating stable interaction with the IRS keratins [57-60]. So it was very interesting to note that mutations in the trichohyalin gene were described for uncombable hair syndrome, which includes a curly hair phenotype [61]. In addition to TCHH, the genes for PAD (PADI3) and transglutaminase 3 (TGM3), both involved in the transformation of the IRS from 'soft' to 'hard' were also mutated in this rare genetic condition. On the inner side of the IRS, the IRS cuticle cells form a 'mirror' of the cuticle cells of the hair shaft holding the shaft very firmly into the follicle. Outside Henle's layer there is the companion layer (or innermost layer of the outer root sheath). These cells are intimately connected to the IRS and migrate with the shaft as it grows. Of special mention are so called flügelzellen, structures in Huxley's layer which project through Henle's layer to the companion layer. These structures are predicted to strengthen and stabilise the IRS [62, 63] with the latter already known to be highly influential on hair curl formation. Flügelzellen may be visualised by staining with antibodies to K74, directly linking this keratin with Flügelzellen. Further study of the spatial disposition of Flügelzellen in relation to hair curl is warranted. Thus, the current thinking is that the IRS is not merely a scaffold holding the shaft but is able to be programmed to confer properties on the shaft including shape.

5) Dermal Papilla asymmetry and links to curl

Little or no attention has been paid to whether the size and shape of the papilla contributes to the shape of the hair fiber and subsequent curliness in human hair, although the role of the DP generally in contributing to fibre type and shape was recently reviewed [64] and the links between DP size and fibre size are known [65]. Differences in DP shape are observed animals in relation to types of fibre produced [66] with the spiny mouse (*Acomys dimidiatus)* being a good example of how development of a crescent shaped DP influences the follicle proliferation and differentiation programs [67] to generate an unusually shaped fibre. A possible theory has been proposed in which the asymmetrical

distribution of proliferating cells in the hair follicle bulb matrix leads to a flattened hair fiber shape [68] and the asymmetrical control on matrix cell proliferation is assumed to be controlled by the dermal papilla, suggesting that it too has an asymmetry in relation to interactions with the surrounding hair matrix cells which could lead to the formation of a curly hair. Nissimov 2014 further hypothesized that the construction of certain features of curly hair had explanations within the construction of the hair follicle, proposing multiple papillary centres each autonomously influencing growth of adjacent bulb matrix cells, so building asymmetry [69].

Curly hair as a genetic trait: Identification of candidate genes and links to mechanistic factors involved in curly hair formation

Curly hair traits are straightforward if rather tedious to measure given that hair is easily sampled and good methods to quantify curl have been developed [3]. This has aided genetic studies (so called genome wide association studies (GWAS)) to try to identify the causative genes for hair traits and to explain their role in hair shape [70, 71].

Factors such as ethnicity and geographic variation must be controlled in these studies to minimise false positives. The advantage of GWAS investigations lies in the complete survey of the genome without prior hypothesis and the potential ability to identify unsuspected, novel genetic links to hair curl and shape. The most recent data to emerge from such studies is from the CANDELA cohort, a large (6630) admixed South American population with European, Native American and African ancestry [71]. In this study, hair shape was scored on a fairly simple four-point scale (straight, wavy, curly or frizzy) and was found to be associated with polymorphic variation in known curl associated genes (EDAR, Trichohyalin) and as yet unknown genes. PRSS53, Protease Serine S1 family member 53a is a serine protease expressed in the IRS and was shown by the authors to have a variant Q30R substitution causing a change in enzyme activity with recent evolutionary selection in East Asian populations. Its expression in the IRS adds weight to the hypothesis that shape of hair fibre is governed by the construction of the IRS; mechanism 4 as described above.

In a separate GWAS, designed to examine the curl variation only within South African populations, Unilever R&D studied 3 separate language (ethnic) groups; the black African Sotho/Tswana, Xhosa and Zulu peoples, for genetic links to hair curl variation within what is a largely similar African ancestral population. Prior observations in South Africa revealed wide variation in curl type and degree, lending weight to the hypothesis that curl was under polymorphic control; the key question was which proteins might be variable? The degree of curliness of hair samples from 2417 volunteers was measured accurately using a flat bed scanner and image analysis, with the overall curl variation observed shown in Figure 5. No significant differences in curl variation were seen between language groups; there was a trend for the Zulu language group to have less curly hair. DNA from the 25% highest curl and lowest curl subjects was compared using a DNA pooling strategy and assessing 1.6M single nucleotide polymorphic variants (SNPs). For general methods used see Stokowski RP et al [72]. A substantial genetic signal was detected comparing the two hair curl groups but overall there were no specific associations that passed a strict genome wide statistical test (5 X 10^{-8} after a Bonferroni multiple testing correction). These data suggest that black African hair curl variation is 'complex' in that many genes are involved each having a modest effect on hair curl. Never the less, 3 candidate genes were selected having suggestive links to curl based on a less strict statistical tests, follicle location and literature data (Table 2).

Two of the 3 genes listed in table 2 (KRT74 and TCHH) are located in the IRS, which again supports the hypothesis that the IRS strongly influences hair shape. (see also [73], [74], [7, 75], [76], [77] [61]. K74 (keratin 74 the protein product of KRT74) is found in Huxley's layer (Figure 2) and is also linked to woolly hair syndromes [76] a disorder manifest by fine curly hair. The role for the IRS in shaping hair curl is also supported by animal studies, for example Cadieu et al [78] demonstrated using pure bred dogs, that just three genes control the major coat attributes of length, curliness and facial hair such as long eyebrows and beard. In humans polymorphic variation in KRT71 also gives rise to woolly hair [79]. Thus both KRT71 and KRT74 variants underpin a curly hair phenotype most likely by altered structural behaviour (e.g. capacity to bend, flex or twist) of the K71 and K74 proteins.

Trichohyalin (the protein product of TCHH) is also expressed in Huxley's layer of the IRS and in the medulla. Trichohyalin is responsible for condensing the intermediate filaments as they change and harden. Electrostatic links to intermediate filaments are further stabilised by the action of peptide cross-linking enzymes called transglutaminases (TGase) [80] and, in particular, TGase3, appears to be very important in formation of important cross linkages in the hair fibre [81]; mouse TGase3 gene (TGM3) knock out studies show hair abnormalities as the major phenotype [82] and TGM3 gene is mutated in uncombable hair syndrome [61]. In terms of function, it is proposed that trichohyalin mechanically strengthens the hair follicle inner root sheath to subsequently contain and permit shape to be set into the hair fibre [59, 83]. Interestingly an independent study in people of western European descent living in Australia [84] suggests that trichohyalin polymorphisms are linked to the straightness of hair and therefore that, when combined with the observations reported here, trichohyalin might influence hair shape across more than one human population.

All 3 genes highlighted by the GWAS; KRT71, KRT74 and TCHH, are members of the so-called epidermal differentiation complex (EDC), a cluster of about 20 genes in chr1q21. A subset of EDC genes is therefore clearly involved in coordinating hair shape. It is known that the EDC is under epigenetic control in the epidermis [85, 86] with chromatin organisers key to epidermal differentiation. It is interesting to speculate that similar factors may also control genes in the EDC within the IRS, opening up the possibility for epigenetic regulation of hair shape.

The third gene listed in table 2 is CUTC (cutC copper transporter) with members of the family associated with copper homeostasis, namely the uptake, storage, delivery and efflux of copper. From animal studies copper is known to be associated with hair conditions including hair curl [87]. For example copper deficiency in lambs leads to poor quality wool that lacks crimp, an effect linked to the delayed differentiation of the IRS [7]. Menkes disease, which is associated with defects in hair traits including hair kinks, [88] is linked to another copper

transporter ATP7A, further supporting a role for copper homeostasis in affecting hair curl.

Curly hair follicle development is under the control of major developmental programmes

Genome-wide searches have also uncovered evidence that developmental genes are involved in shaping hair curl. The ectodysplasin receptor family (EDAR) are cell surface receptors of the tumour necrosis factor family (TNF) expressed in skin and hair follicles during hair follicle development, again at puberty and during the hair cycle [89]. Recently EDAR has been implicated in the control of hair shape and fibre thickness [90] [91]. Positive selection of a polymorphism in east Asian and native American populations about 10,000 years ago is believed to have affected both follicle size and fibre thickness as well as shovel-shaped incisor teeth and increased secretions of sebum and meiobian lipids in the eye and saliva [92] [93]. As indicated earlier, the shape of the hair follicle is set through embryogenesis and then curl manifests during childhood. Thus it is not surprising that factors involved in embryogenesis affect hair shape. The interesting question is why East Asians developed straight hair? One explanation is that the glandular changes may have been the driving force behind the penetrance of the new gene variant in East Asians, with straighter hair being a non-selective consequence of advantageous changes in tooth shape and gland activity. The hair phenotype maybe linked to higher Edar function which, through signalling via sonic hedgehog [94] may have led to greater symmetry in growth rates in the follicle bulb with straighter hair arising as a result.

A second developmental gene associated with hair morphology is suspected to be WNT10A (wingless-type MMTV integration site family, member 10A; [70]. WNT10A is upregulated at the beginning of the hair growth cycle and mutations in this gene are known to cause misformed hair [95] and appendage abnormalities in Hypohidrotic Ectodermal Dysplasia patients [96]. However, a known mutation in Wnt10A (rs7349332) in combination with mutations in TCHH (rs11803731and FRAS 1 (rs1268789) form a potential signature for straight hair of potential use in forensics [97].

In summary, the shape, type and colour of hair are determined not just during embryogenesis but also repeatedly in each hair growth cycle. Aside from pattern baldness, characteristic hair types are maintained in bodily patterns throughout life. Natural population variance in hair curl appears to have a largely genetic basis and environmental pressure selecting for specialised hair morphology may well have arisen when humans migrated out of Africa. There is evidence that trichohyalin (TCHH) may affect hair curl in most/all world populations and that other genes such as EDAR, WNT10A only affect specific populations. Hair curl variation in native Africans is very likely a complex trait with multiple genes influencing curl. The strongest evidence for the control of shape comes from the evidence of the role of the inner root sheath which appears to structurally mould hair fibre shape, including curl - but we are still a very long way from understanding the complete biological/biophysical mechanisms that produce such a wide range of curled, coiled, kinked and wavy hair fibres.

Funding Sources

MR Green and RS Ginger are employed by Unilever R&D.

Conflicts of Interest

The authors state no conflicts of interest.

References

1. Thibaut, S., et al., *Human eyelash characterization.* Br J Dermatol, 2010. **162**(2): p. 304-10.
2. Hrdy, D., *Quantitative hair form variation in seven populations.* Am J Phys Anthropol, 1973. **39**(1): p. 7-17.
3. Loussouarn, G., et al., *Worldwide diversity of hair curliness: a new method of assessment.* Int J Dermatol, 2007. **46 Suppl 1**: p. 2-6.

4. Loussouarn, G., et al., *Diversity in human hair growth, diameter, colour and shape. An in vivo study on young adults from 24 different ethnic groups observed in the five continents.* Eur J Dermatol, 2016. **26**(2): p. 144-54.
5. De la Mettrie, R., et al., *Shape variability and classification of human hair: a worldwide approach.* Hum Biol, 2007. **79**(3): p. 265-81.
6. Nagorcka, B.N., *Evidence for a reaction-diffusion system as a mechanism controlling mammalian hair growth.* Biosystems, 1983. **16**(3-4): p. 323-32.
7. Hynd, P.I., et al., *Wool fibre crimp is determined by mitotic asymmetry and position of final keratinisation and not ortho- and para-cortical cell segmentation.* Animal, 2009. **3**(6): p. 838-43.
8. Bernard, B.A., *Hair shape of curly hair.* J Am Acad Dermatol, 2003. **48**(6 Suppl): p. S120-6.
9. Thibaut, S., et al., *Human hair shape is programmed from the bulb.* Br J Dermatol, 2005. **152**(4): p. 632-8.
10. Thibaut, S. and B.A. Bernard, *The biology of hair shape.* Int J Dermatol, 2005. **44 Suppl 1**: p. 2-3.
11. Thibaut S, C.C., Langbein L, Schweizer J, Gautier B, Bernard BA, *Hair keratin pattern in human hair follicles grown in vitro.* Exp Dermatol, 2003. **12**: p. 160-164.
12. Sebastien Thibaut, P., Philippe Barbarat, PhD, Frederic Leroy, PhD, and Bruno A. Bernard, PhD, *Human hair keratin network and curvature.* Int J Dermatol. **46**(Suppl. 1): p. 7-10.
13. Mangelsdorf, S., et al., *Ethnic variation in vellus hair follicle size and distribution.* Skin Pharmacol Physiol, 2006. **19**(3): p. 159-67.
14. Kligman, A.M., *Neogenesis of human hair follicles.* Ann N Y Acad Sci, 1959. **83**: p. 507-11.
15. Robins, E.J. and A.S. Breathnach, *Fine structure of the human foetal hair follicle at hair-peg and early bulbous-peg stages of development.* J Anat, 1969. **104**(Pt 3): p. 553-69.
16. Akiyama, M., I. Matsuo, and H. Shimizu, *Formation of cornified cell envelope in human hair follicle development.* Br J Dermatol, 2002. **146**(6): p. 968-76.

17. Ajose, F.O., *Diseases that turn African hair silky.* Int J Dermatol, 2012. **51 Suppl 1**: p. 12-6, 14-9.
18. Sun, T.T., G. Cotsarelis, and R.M. Lavker, *Hair follicular stem cells: the bulge-activation hypothesis.* J Invest Dermatol, 1991. **96**(5): p. 77S-78S.
19. Bernard, B.A., *The human hair follicle, a bistable organ?* Exp Dermatol, 2012. **21**(6): p. 401-3.
20. Hibino, T. and T. Nishiyama, *Role of TGF-beta2 in the human hair cycle.* J Dermatol Sci, 2004. **35**(1): p. 9-18.
21. Philpott, M.P., M.R. Green, and T. Kealey, *Human hair growth in vitro.* J Cell Sci, 1990. **97 (Pt 3)**: p. 463-71.
22. Philpott, M.P., G.E. Westgate, and T. Kealey, *An in vitro model for the study of human hair growth.* Ann N Y Acad Sci, 1991. **642**: p. 148-64; discussion 164-6.
23. Kligman, A.M., *The human hair cycle.* J Invest Dermatol, 1959. **33**: p. 307-16.
24. Stenn, K., *Exogen is an active, separately controlled phase of the hair growth cycle.* J Am Acad Dermatol, 2005. **52**(2): p. 374-5.
25. Higgins, C.A., G.E. Westgate, and C.A. Jahoda, *From telogen to exogen: mechanisms underlying formation and subsequent loss of the hair club fiber.* J Invest Dermatol, 2009. **129**(9): p. 2100-8.
26. Stenn, K.S. and R. Paus, *Controls of hair follicle cycling.* Physiol Rev, 2001. **81**(1): p. 449-494.
27. Yang, C.C. and G. Cotsarelis, *Review of hair follicle dermal cells.* J Dermatol Sci, 2010. **57**(1): p. 2-11.
28. Millar, S.E., *Committing to a hairy fate: epigenetic regulation of hair follicle stem cells.* Cell Stem Cell, 2011. **9**(3): p. 183-4.
29. Murray, P.J., et al., *Modelling hair follicle growth dynamics as an excitable medium.* PLoS Comput Biol, 2012. **8**(12): p. e1002804.
30. Botchkarev, V.A., et al., *Epigenetic regulation of gene expression in keratinocytes.* J Invest Dermatol, 2012. **132**(11): p. 2505-21.
31. Beck, B. and C. Blanpain, *Mechanisms regulating epidermal stem cells.* EMBO J, 2012. **31**(9): p. 2067-75.

32. Paus, R., S. Muller-Rover, and V.A. Botchkarev, *Chronobiology of the hair follicle: hunting the " hair cycle clock".* J Investig Dermatol Symp Proc, 1999. **4**(3): p. 338-45.
33. Paus, R. and K. Foitzik, *In search of the "hair cycle clock": a guided tour.* Differentiation, 2004. **72**(9-10): p. 489-511.
34. Geyfman, M. and B. Andersen, *Clock genes, hair growth and aging.* Aging (Albany NY), 2010. **2**(3): p. 122-8.
35. Rudman, S.M., et al., *The role of IGF-I in human skin and its appendages: morphogen as well as mitogen?* J Invest Dermatol, 1997. **109**(6): p. 770-7.
36. Lachgar, S., et al., *Minoxidil upregulates the expression of vascular endothelial growth factor in human hair dermal papilla cells.* Br J Dermatol, 1998. **138**(3): p. 407-11.
37. Yano, K., L.F. Brown, and M. Detmar, *Control of hair growth and follicle size by VEGF-mediated angiogenesis.* J Clin Invest, 2001. **107**(4): p. 409-17.
38. Millar, S.E., et al., *WNT signaling in the control of hair growth and structure.* Dev Biol, 1999. **207**(1): p. 133-49.
39. Huelsken, J., et al., *beta-Catenin controls hair follicle morphogenesis and stem cell differentiation in the skin.* Cell, 2001. **105**(4): p. 533-45.
40. Lei, M.X., C.M. Chuong, and R.B. Widelitz, *Tuning Wnt signals for more or fewer hairs.* J Invest Dermatol, 2013. **133**(1): p. 7-9.
41. Abbasi, A.A., *Molecular evolution of HR, a gene that regulates the postnatal cycle of the hair follicle.* Sci Rep, 2011. **1**: p. 32.
42. Cannistra, C., et al., *New perspectives in the treatment of hidradenitis suppurativa: surgery and brewer's yeast-exclusion diet.* Surgery, 2013. **154**(5): p. 1126-30.
43. Van Scott, E.J. and T.M. Ekel, *Geometric relationships between the matrix of the hair bulb and its dermal papilla in normal and alopecic scalp.* J Invest Dermatol, 1958. **31**(5): p. 281-7.
44. Sriwiriyanont, P., et al., *Effects of IGF-binding protein 5 in dysregulating the shape of human hair.* J Invest Dermatol, 2011. **131**(2): p. 320-8.
45. Rogers, M.A., et al., *The human type I keratin gene family: characterization of new hair follicle specific members and evaluation of the chromosome 17q21.2 gene domain.* Differentiation, 2004. **72**(9-10): p. 527-40.

46. Thibaut, S., et al., *Human hair keratin network and curvature.* Int J Dermatol, 2007. **46 Suppl 1**: p. 7-10.
47. Kaplin, I.J. and K.J. Whiteley, *An electron microscope study of fibril: matrix arrangements in high- and low-crimp wool fibres.* Aust J Biol Sci, 1978. **31**(3): p. 231-40.
48. Harland, D.P., et al., *Arrangement of trichokeratin intermediate filaments and matrix in the cortex of Merino wool.* J Struct Biol, 2011. **173**(1): p. 29-37.
49. Philpott, M.P., D.A. Sanders, and T. Kealey, *Effects of insulin and insulin-like growth factors on cultured human hair follicles: IGF-I at physiologic concentrations is an important regulator of hair follicle growth in vitro.* J Invest Dermatol, 1994. **102**(6): p. 857-61.
50. Joshi, R.S., *The Inner Root Sheath and the Men Associated with it Eponymically.* Int J Trichology, 2011. **3**(1): p. 57-62.
51. Alibardi, L., *Comparative aspects of the inner root sheath in adult and developing hairs of mammals in relation to the evolution of hairs.* J Anat, 2004. **205**(3): p. 179-200.
52. Alibardi, L., *Fine structure and immunocytochemistry of monotreme hairs, with emphasis on the inner root sheath and trichohyalin-based cornification during hair evolution.* J Morphol, 2004. **261**(3): p. 345-63.
53. O'Keefe, E.J., et al., *Trichohyalin: a structural protein of hair, tongue, nail, and epidermis.* J Invest Dermatol, 1993. **101**(1 Suppl): p. 65S-71S.
54. Lee, S.C., et al., *Human trichohyalin gene is clustered with the genes for other epidermal structural proteins and calcium-binding proteins at chromosomal locus 1q21.* J Invest Dermatol, 1993. **100**(1): p. 65-8.
55. Fietz, M.J., et al., *Mapping of the trichohyalin gene: co-localization with the profilaggrin, involucrin, and loricrin genes.* J Invest Dermatol, 1992. **99**(5): p. 542-4.
56. Yamamoto, S., et al., *Molecular elements of the regulatory control of keratin filament modulator AHF/trichohyalin in the hair follicle.* Exp Dermatol, 2009. **18**(2): p. 152-9.

57. Rogers, G., et al., *Peptidylarginine deiminase of the hair follicle: characterization, localization, and function in keratinizing tissues.* J Invest Dermatol, 1997. **108**(5): p. 700-7.
58. Rogers, G., et al., *Hair follicle peptidylarginine deiminase.* Exp Dermatol, 1999. **8**(4): p. 362-3.
59. Steinert, P.M. and L.N. Marekov, *Multiple roles for trichohyalin in the inner root sheath.* Exp Dermatol, 1999. **8**(4): p. 331-2.
60. Nachat, R., et al., *Peptidylarginine deiminase isoforms are differentially expressed in the anagen hair follicles and other human skin appendages.* J Invest Dermatol, 2005. **125**(1): p. 34-41.
61. FB, U.B., et al., *Mutations in Three Genes Encoding Proteins Involved in Hair Shaft Formation Cause Uncombable Hair Syndrome.* Am J Hum Genet, 2016. **99**(6): p. 1292-1304.
62. Langbein, L., et al., *A novel epithelial keratin, hK6irs1, is expressed differentially in all layers of the inner root sheath, including specialized huxley cells (Flugelzellen) of the human hair follicle.* J Invest Dermatol, 2002. **118**(5): p. 789-99.
63. Langbein, L., et al., *K6irs1, K6irs2, K6irs3, and K6irs4 represent the inner-root-sheath-specific type II epithelial keratins of the human hair follicle.* J Invest Dermatol, 2003. **120**(4): p. 512-22.
64. Morgan, B.A., *The dermal papilla: an instructive niche for epithelial stem and progenitor cells in development and regeneration of the hair follicle.* Cold Spring Harb Perspect Med, 2014. **4**(7): p. a015180.
65. Elliott, K., T.J. Stephenson, and A.G. Messenger, *Differences in hair follicle dermal papilla volume are due to extracellular matrix volume and cell number: implications for the control of hair follicle size and androgen responses.* J Invest Dermatol, 1999. **113**(6): p. 873-7.
66. Chi, W., E. Wu, and B.A. Morgan, *Dermal papilla cell number specifies hair size, shape and cycling and its reduction causes follicular decline.* Development, 2013. **140**(8): p. 1676-83.
67. Montandon, S.A., et al., *Two waves of anisotropic growth generate enlarged follicles in the spiny mouse.* Evodevo, 2014. **5**: p. 33.

68. Claudine Pie´rard-Franchimont, M., PhD, Philippe Paquet, MD, PhD, Pascale Quatresooz, MD, PhD & Ge´rald E Pie´rard, MD, PhD, *Mechanobiology and cell tensegrity: the root of ethnic hair curling?* J Cosmetic Dermatol, 2011. **10**: p. 163-167.
69. Nissimov, J.N. and A.B. Das Chaudhuri, *Hair curvature: a natural dialectic and review.* Biol Rev Camb Philos Soc, 2014. **89**(3): p. 723-66.
70. Eriksson, N., et al., *Web-based, participant-driven studies yield novel genetic associations for common traits.* PLoS Genet, 2010. **6**(6): p. e1000993.
71. Adhikari, K., et al., *A genome-wide association scan in admixed Latin Americans identifies loci influencing facial and scalp hair features.* Nat Commun, 2016. **7**: p. 10815.
72. Stokowski, R.P., et al., *A genomewide association study of skin pigmentation in a South Asian population.* Am J Hum Genet, 2007. **81**(6): p. 1119-32.
73. Du, X., et al., *Velvet, a dominant Egfr mutation that causes wavy hair and defective eyelid development in mice.* Genetics, 2004. **166**(1): p. 331-40.
74. Raveh, E., et al., *Runx3 is involved in hair shape determination.* Dev Dyn, 2005. **233**(4): p. 1478-87.
75. Nahum, S., et al., *A large duplication in LIPH underlies autosomal recessive hypotrichosis simplex in four Middle Eastern families.* Arch Dermatol Res, 2009. **301**(5): p. 391-3.
76. Shimomura, Y., et al., *Autosomal-dominant woolly hair resulting from disruption of keratin 74 (KRT74), a potential determinant of human hair texture.* Am J Hum Genet, 2010. **86**(4): p. 632-8.
77. Westgate, G.E., N.V. Botchkareva, and D.J. Tobin, *The biology of hair diversity.* Int J Cosmet Sci, 2013.
78. Cadieu, E., et al., *Coat variation in the domestic dog is governed by variants in three genes.* Science, 2009. **326**(5949): p. 150-3.
79. Fujimoto, A., et al., *A missense mutation within the helix initiation motif of the keratin K71 gene underlies autosomal dominant woolly hair/hypotrichosis.* J Invest Dermatol, 2012. **132**(10): p. 2342-9.
80. Rogers, G.E., H.W. Harding, and I.J. Llewellyn-Smith, *The origin of citrulline-containing proteins in the hair follicle and the chemical nature of*

trichohyalin, an intracellular precursor. Biochim Biophys Acta, 1977. **495**(1): p. 159-75.

81. Thibaut, S., et al., *Transglutaminase-3 enzyme: a putative actor in human hair shaft scaffolding?* J Invest Dermatol, 2009. **129**(2): p. 449-59.
82. John, S., et al., *Epidermal transglutaminase (TGase 3) is required for proper hair development, but not the formation of the epidermal barrier.* PLoS One, 2012. **7**(4): p. e34252.
83. Steinert, P.M., D.A. Parry, and L.N. Marekov, *Trichohyalin mechanically strengthens the hair follicle: multiple cross-bridging roles in the inner root shealth.* J Biol Chem, 2003. **278**(42): p. 41409-19.
84. Medland, S.E., et al., *Common variants in the trichohyalin gene are associated with straight hair in Europeans.* Am J Hum Genet, 2009. **85**(5): p. 750-5.
85. Mardaryev, A.N., et al., *p63 and Brg1 control developmentally regulated higher-order chromatin remodelling at the epidermal differentiation complex locus in epidermal progenitor cells.* Development, 2014. **141**(1): p. 101-11.
86. Botchkarev, V.A., *Integration of the Transcription Factor-Regulated and Epigenetic Mechanisms in the Control of Keratinocyte Differentiation.* J Investig Dermatol Symp Proc, 2015. **17**(2): p. 30-2.
87. Marston, H.R., Lee, H.J., *Nutritional factors involved in wool production by merino sheep.* Aust J Scientific Res B, 1948. **1**: p. 376-387.
88. Danks, D.M., et al., *Menkes' kinky hair disease: further definition of the defect in copper transport.* Science, 1973. **179**(4078): p. 1140-2.
89. Fessing, M.Y., et al., *Involvement of the Edar signaling in the control of hair follicle involution (catagen).* Am J Pathol, 2006. **169**(6): p. 2075-84.
90. Medland, S.E., G. Zhu, and N.G. Martin, *Estimating the heritability of hair curliness in twins of European ancestry.* Twin Res Hum Genet, 2009. **12**(5): p. 514-8.
91. Tan, J., et al., *The adaptive variant EDARV370A is associated with straight hair in East Asians.* Hum Genet, 2013.
92. Kimura, R., et al., *A common variation in EDAR is a genetic determinant of shovel-shaped incisors.* Am J Hum Genet, 2009. **85**(4): p. 528-35.

93. Peng, Q., et al., *EDARV370A associated facial characteristics in Uyghur population revealing further pleiotropic effects.* Hum Genet, 2016. **135**(1): p. 99-108.

94. Mou, C., et al., *Enhanced ectodysplasin-A receptor (EDAR) signaling alters multiple fiber characteristics to produce the East Asian hair form.* Hum Mutat, 2008. **29**(12): p. 1405-11.

95. Adaimy, L., et al., *Mutation in WNT10A is associated with an autosomal recessive ectodermal dysplasia: the odonto-onycho-dermal dysplasia.* Am J Hum Genet, 2007. **81**(4): p. 821-8.

96. Zeng, B., et al., *Eight Mutations of Three Genes (EDA, EDAR, and WNT10A) Identified in Seven Hypohidrotic Ectodermal Dysplasia Patients.* Genes (Basel), 2016. **7**(9).

97. Pospiech, E., et al., *Evaluation of the predictive capacity of DNA variants associated with straight hair in Europeans.* Forensic Sci Int Genet, 2015. **19**: p. 280-8.

Figures and Tables

Figure 1. Variation in degree of curl in human hair fibres. Hair fibres were sampled from populations in South Africa (a) showing a range from high curl (left) to low curl (right). Magnified images of fibre samples from low (b) and high (c) curl reveal that both degree of curliness (tightness of the curl) and the change in direction of the curl contribute to overall curliness.

Figure 2. Diagrammatic representation of the lower hair follicle bulb. This figure shows the distribution of the 7 layers that comprise the fundamental hair follicle structure as well as the dermal papilla and connective tissue sheath. This diagram has been kindly provided by Dr. Claire Higgins.

Figure 3. Representation of the curly hair follicle in three dimensions. Figure 3a shows the line of symmetry around the long axis of the hair in a straight hair follicle. Figure 3b the long axis is shown as symmetrical through the dermal papilla of a curly hair follicle. Note that the hair shaft is no-longer in plane with the dermal papilla, Figure 3c shows the section through the bulb depicting the curved shape of the lower follicle.

Figure 4. A series of images showing the retrocurvature of the hair follicle when sectioning through a sample of scalp skin from an individual with very curly hair. The curly hair follicle is curved in two directions – left to right and fore and back. a) the upper bulb is out of the image, yet the shaft is sectioned through. b) the follicle curves twice (arrows). c) the bulb is sectioned through the mid dermal papilla d) the dermal papilla is almost out of section and e) the bend in the upper follicle reveals connective tissue sheath (arrow).

Figure 5. Population distribution of South African hair curl covering the main language groups (Sotho/Tswana, Xhosa and Zulu). Average Mean Curvature

(1/radius, AMC) was mathematically averaged over each fibre separately, before the mean value for 20 hairs from each volunteer was calculated. The range of hair AMC sampled was from 0.14 to 1.545. As illustrated in the figure, more curly hair has a higher AMC.

Table 1
Biomarkers associated with curl in the hair follicle

Biomarker	Localisation	Asymmetry	Reference
K38	Cortex	Earlier expression on concave side	[46]
K82	Cuticle	Later expression on convex side	[9]
Ki67	Bulb matrix	Proliferation is above the line of 'Auber' on the convex side	[9] [1]
K14	ORS	ORS is thicker on the concave side	[1, 9]
IGFBP-5	ORS	Greater expression on convex side	[44]
K74	IRS	Mutations give rise to wooly hair	[76]
K71	IRS	Mutations give rise to wooly hair and curly hair in dogs	[78, 79]
Trichohyalin	IRS	SNP associates with straighter hair in caucasians	[84]
Fibronectin	CTS	CTS is thicker on the concave side	[9]

Table 2
Candidate genetic associations influencing black African hair curl

Gene region	Possible function	SNPs	p-value
KRT74	IRS keratin linked to woolly hair syndrome. Adjacent to KRT71 which strongly affects hair curl in dogs	rs3912631	$<3x10^{-05}$
TCHH	Hair follicle specific protein also found in the IRS. Linked to hair curve in peoples of western European descent	Afd_1108920*	$<1x10^{-6}$
CUTC	Copper transport homologue. Copper changes linked to wool crimp in sheep and 'kinky hair' in Menkes disease	rs4919394 rs978554 rs7078602	$<5x10^{-7}$ $<9x10^{-7}$ $<1x10^{-6}$

p-values include a genome wide Bonferroni correction
* Afd_1108920 is a SNP used on the Perlegen genotyping platform, 7kb from rs11803731 identified by Medland et al [84]. The rs11803731 alternate allele is found only in populations of European descent and therefore is not informative for black African hair.

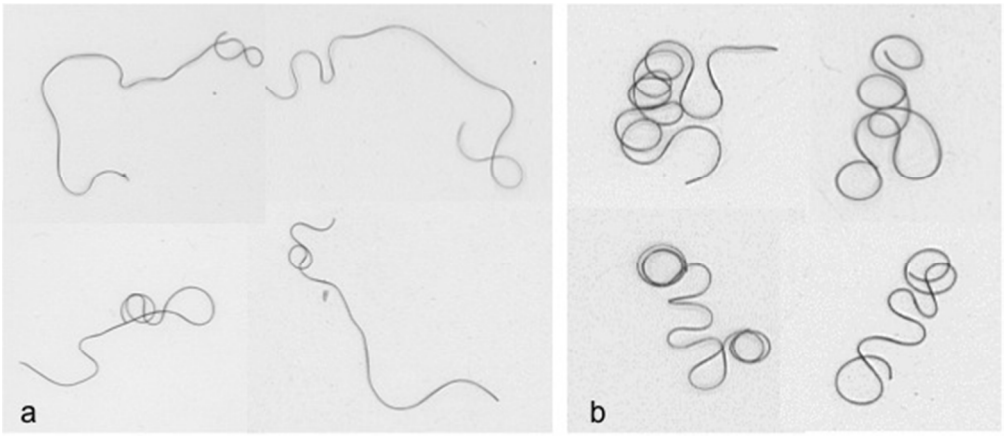

Figure 1. Variation in degree of curl in human hair fibres. Hair fibres were sampled from populations in South Africa (a) showing a range from high curl (left) to low curl (right). Magnified images of fibre samples from low (b) and high (c) curl reveal that both degree of curliness (tightness of the curl) and the change in direction of the curl contribute to overall curliness.

Figure 1
214x92mm (72 x 72 DPI)

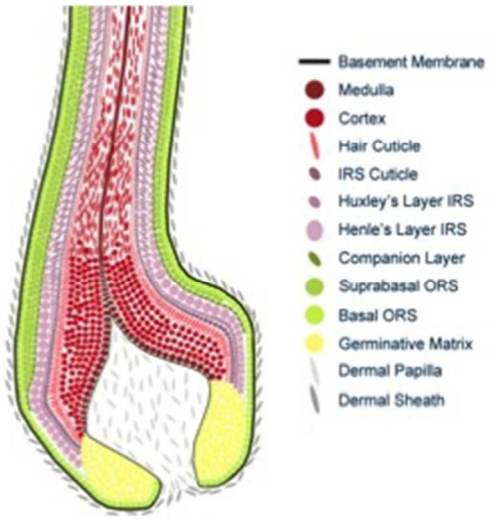

Figure 2. Diagrammatic representation of the lower hair follicle bulb. This figure shows the distribution of the 7 layers that comprise the fundamental hair follicle structure as well as the dermal papilla and connective tissue sheath. This diagram has been kindly provided by Dr. Claire Higgins.

Figure 2
103x97mm (72 x 72 DPI)

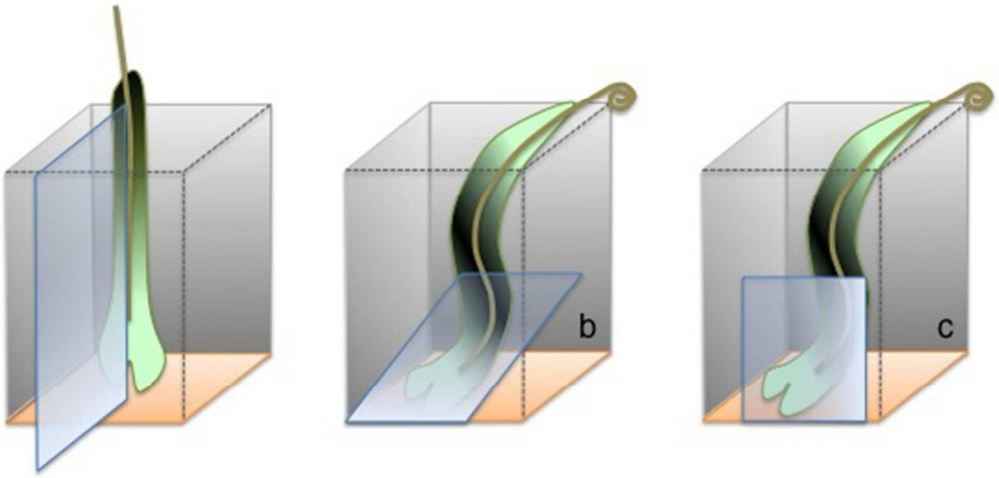

Figure 3. Representation of the curly hair follicle in three dimensions. Figure 3a shows the line of symmetry around the long axis of the hair in a straight hair follicle. Figure 3b the long axis is shown as symmetrical through the dermal papilla of a curly hair follicle. Note that the hair shaft is no-longer in plane with the dermal papilla, Figure 3c shows the section through the bulb depicting the curved shape of the lower follicle.

Figure 3
191x91mm (72 x 72 DPI)

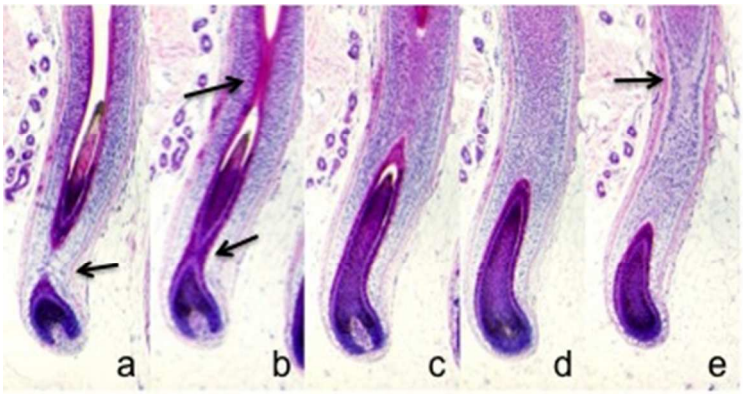

Figure 4. A series of images showing the retrocurvature of the hair follicle when sectioning through a sample of scalp skin from an individual with very curly hair. The curly hair follicle is curved in two directions – left to right and fore and back. a) the upper bulb is out of the image, yet the shaft is sectioned through. b) the follicle curves twice (arrows). c) the bulb is sectioned through the mid dermal papilla d) the dermal papilla is almost out of section and e) the bend in the upper follicle reveals connective tissue sheath (arrow).

Figure 4
130x68mm (72 x 72 DPI)

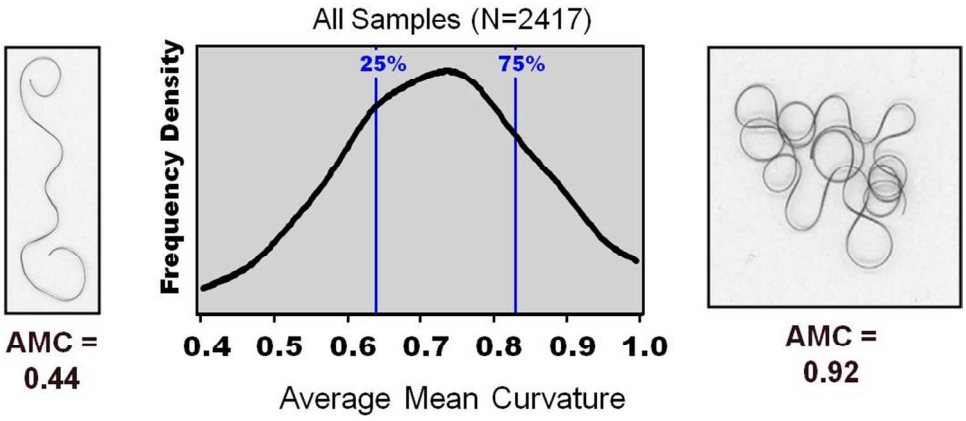

Figure 5. Population distribution of South African hair curl covering the main language groups (Sotho/Tswana, Xhosa and Zulu). Average Mean Curvature (1/radius, AMC) was mathematically averaged over each fibre separately, before the mean value for 20 hairs from each volunteer was calculated. The range of hair AMC sampled was from 0.14 to 1.545. As illustrated in the figure, more curly hair has a higher AMC.

Figure 5
235x104mm (120 x 120 DPI)