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Development of a Mathematical Model for Apple Juice Compounds Rejection in a Spiral-wound Reverse Osmosis Process

Al-Obaidi M. A., Kara-Zaitri C. and Mujtaba I. M. *

School of Engineering, Faculty of Engineering and Informatics. University of Bradford, Bradford, West Yorkshire BD7 1DP, UK

*Corresponding author, Tel.: +44 0 1274 233645

E-mail address: I.M.Mujtaba@bradford.ac.uk

Abstract

The use of Reverse Osmosis (RO) membrane processes for the clarification and the concentration of apple juice is proposed as an alternative to the conventional concentration technique, which is based on evaporation and freezing. Several models have been published on RO process models relying on different assumptions that predict the permeate flux and aroma compounds rejections for aqueous solutions apple juice. The solution-diffusion model (Lumped model) has been applied for the previous models. The main instrument of this study is the use of the gPROMS software to develop a new distributed steady state model that will relax a number of earlier assumptions.

The model has been validated with an observational data of apple juice filtration derived from the literature by analysing the permeate flux and the performance of membrane rejection at different concentrations, temperatures and pressures for a laboratory scale of spiral-wound RO module. Simulated results corroborate with experimental and model predictions.

Keywords: Apple Juice Concentration; Spiral-wound Reverse Osmosis; One Dimensional Distributed Model; gPROMS software.

1. Introduction

The concentration of fruit juices is achieved by reducing the water content. This has many advantages, including easier and cheaper conservation, storage, transportation and distribution of the extracted juice. Conventional methods of fruit juices concentration are usually conducted using a high temperature multi-stage vacuum evaporation process. This process usually results in significant losses of nutritional compounds, such as vitamin C, as well as associated thermal effects (Pozderovic´ et al. 2006). As a result, RO has become an alternative process to the conventional methods for removing water from fruit juices and other liquid foods (Girard and Fukumoto, 2000). However, one of the main disadvantages of using RO is related to lower concentration of the yield in comparison to the thermal process due to high osmotic pressure limitation. Having said this, the RO process has affirmed its

35 potential as the prominent process for retaining the aroma compounds (Jiao et al., 2004).
36 Therefore, the RO process is a well-recognized technique for concentrating aqueous solutions
37 within a limit of 25 to 30 °Brix, which is quite below the typical value of 45 to 60 °Brix gain
38 by the evaporation process (Pepper, 1990). With the aim to improve the concentration and yet
39 retain the critical aroma feature, this research proposes a RO process but one that is used as a
40 pre-concentration step with other technologies (Sheu and Wiley, 1983).

41 Due to the obvious high commercial implications, several attempts can be found in the
42 literature to improve the concentration of apple juice. Many such models are based on the RO
43 process in order to measure permeate flux and to elucidate the rejection of one component
44 and multi-component fruit solutions. A critique on the current literature is discussed in the
45 following section.

46 Nabetani et al. (1992a) have proposed a new correlation to estimate the osmotic pressure of
47 sucrose and glucose solution using thermodynamic definition of the osmotic pressure. The
48 proposed equations assume that the osmotic pressure can be expressed in terms of solute
49 concentration. Accordingly, Nabetani et al. (1992b) have developed a model using the
50 combination of sucrose and glucose osmotic pressure developed in Nabetani et al. (1992a)
51 with the solution-diffusion model equations. The model can predict the permeation of apple
52 juice solution considering the solution physical properties of both one component and a
53 binary solute solution. The model has been validated for a tubular RO module type (ZF 99)
54 supplied by PCI (Paterson Candy International, England) and shows a good agreement
55 between experimental RO data and those calculated on the basis of the solution-diffusion
56 model. However, this particular model considered only sucrose and glucose solute
57 concentration in the bulk retentate with ignored the permeate concentration.

58 Álvarez et al. (1997) have used the solution-diffusion model and the film theory with the
59 proposed osmotic pressure of Nabetani et al. (1992b) to predict the permeate flux in apple
60 juice concentration. This was done by using a tubular polyamide RO membrane type (AFC
61 99) supplied by PCI (Paterson Candy International, England). The model incorporates the
62 physico-chemical correlations to evaluate the characteristics of concentrated apple juice.
63 However, the model ignored the solute concentration at the permeate side and degraded the
64 osmotic pressure caused by fructose and sorbitol in spite of taking into account the
65 contribution of sucrose, glucose and malic acid to the osmotic pressure. Furthermore, Álvarez
66 et al. (1998) used the procedure developed by Matsuura et al. (1974) to calculate the solute

67 transport parameter for each aroma compound for a spiral-wound RO aromatic polyamide
68 membrane type (MSCB 2521 R99) supplied by Separem Spa. (Biella, Italy). Table 1 reports
69 the values of free energy parameter, steric Taft number and solute transport parameter for
70 each aroma compound. Then, the solution-diffusion model is used to estimate the rejection of
71 hydrophilic aroma compounds by considering the average value of inlet feed and retentate as
72 the concentration of bulk solution, but overlooked the osmotic pressure. After that, Álvarez et
73 al. (2001) studied the rejection of aroma compounds using experimental data of solute
74 transport parameter for each aroma compounds calculated for the same above module of RO
75 membrane in the solution-diffusion model. Similarly, this work shows the influence of
76 temperature and feed flow rate on pure water permeability coefficient. The model studied was
77 used to predict the impact of operating conditions such as feed flow rate and concentration on
78 the permeate flux and aroma compound rejections. However, feed osmotic pressure is
79 referred only to glucose.

80 Álvarez et al. (2002) have used the same procedure developed in the work shown above to
81 predict the aroma compounds rejection and the permeate flux during the reverse osmosis
82 concentration of apple juice at laboratory and pilot-scales of MSCB 2521 R99 and MSCE
83 4040 R99 spiral wound membrane supplied by Separem Spa. (Biella, Italy) respectively. The
84 model can predict the influence of operating conditions on permeate flux and aroma
85 compounds rejection. However, this work not only ignored the contribution of fructose and
86 sorbitol in feed osmotic pressure, but also neglected the concentration at the permeate side.

87 To the best of authors' knowledge, all the published RO process modelling for the
88 concentration of apple juice has been carried out using the entire arrangement as a black box
89 and simply taking average inputs and outputs parameter values. In this paper, the finite
90 difference approach is used instead as it gives more accurate results because it takes into
91 account the variation of the operating parameters along the entire system. Also, the above
92 studies have been proposed to describe flux behaviour and compounds retention by relying
93 on the assumption that the osmotic pressure of sugar is only caused by glucose, sucrose and
94 malic acid, and thus ignoring fructose and sorbitol. Pereira et al. (1976) confirmed that
95 glucose and fructose have greater mass transfer and solute transport characteristics than that
96 of sucrose. To systematically resolve this problem, the scope of this paper is to present the
97 development of a distributed one dimensional steady state model and define and assess the
98 variation of the operating conditions as a function of position along the x-axis of the spiral-
99 wound module in the process of apple juice filtration. As well as this, the contribution of all

100 sugar species in the feed osmotic pressure will be taken into consideration. The validation of
101 this model will be based on a comparison between simulation and experimental results
102 derived from the literature. The process model developed can be used later to assess the
103 operating conditions and system design in order to optimize the concentration of apple juice
104 and the retention of aroma compounds.

105 **2. The Main Principles**

106 Reverse Osmosis can be defined as a process of removing undesirable species (salts,
107 pollutants, etc.) from liquid solutions (seawater, wastewater, etc.) by pumping the solution at
108 a hydraulic pressure greater than the osmotic pressure within a closed vessel to move water
109 from high solute side to the diluted side (Jain et al., 2004). There does not appear to be a
110 widespread agreement on the mechanisms of water and solute transport through RO
111 membrane for aqueous solutes (Girard and Fukumoto, 2000). However, the most accepted
112 approaches in this respect are the Solution-diffusion and Preferential sorption theories. The
113 first theory assumes that solvent and solute dissolve in the membrane and pass through by
114 diffusion, while the second theory assumes that solvent and solute are adsorbed at the
115 membrane surface and then pass through the membrane pores.

116 Generally, as the water is removed and the solute is rejected and accumulated at the
117 membrane surface, the water flux drops due to an increase in the osmotic pressure of the feed
118 and concentration polarization impact. There are considered as the main factors causing flux
119 deterioration. These impediments can be fixed by altering the operating condition such as,
120 feed pressure, concentration, temperature and cross-flow velocity and also by turbulence
121 promotion, back flushing/washing and pulsed flow.

122 Normally, a solution treated by RO in food industries is considered as a multi-component
123 solution, which contains a number of solutes at different concentrations. Specifically, apple
124 juice comprises two groups of organic compounds; sugar and aroma compounds, which are
125 categorized as esters (the main compound), aldehydes and alcohols. Also, aroma is one of the
126 most appreciated fresh fruit juice flavor characteristics and is of great importance by
127 consumers. Aroma is due to a large number of volatile organic compounds present in
128 different concentrations, which play a key role in customer perception and satisfaction
129 (Cheong et al., 2010).

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135 **2. Materials and Methods**

136 **2.1 Apple Juice Characteristics**

137 Table 1 shows the composition and concentration of the apple juice (sugar and aroma
138 compounds) used by Álvarez et al. (2002) in all of the experiments as the feed with
139 concentration 10.5 ± 0.5 °Brix. The solution was prepared from 72 °Brix concentrated apple
140 juice supplied by the apple processing company Valle, Ballina and Fernández, S. A.
141 (Villaviciosa, Spain), by adding distilled water.

142 **2.2 Membrane Module Characteristics and Operating Conditions**

143 The RO system used in all of the experiments conducted by Álvarez et al. (2002) was
144 designed in a laboratory scale experiment consisting of a MSCB 2521 R99 spiral-wound
145 aromatic polyamide membrane module supplied by Sparem Spa. (Biella, Italy). The detail of
146 the manufacturer's specification membrane module is presented in Table 2. Experiments are
147 carried out using a batch operation mode where the standard flow configuration of the feed
148 volume is plug flow (passes once time through the system) and the concentrate retentate is
149 recycled back to the feed tank to achieve high system recovery. In addition, permeate was
150 recycled back to the feed tank to maintain a constant concentration and then removed from
151 the equipment which concentration was increased. Experiments are implemented at three
152 different trans-membrane pressures of 14.8, 24.673 and 34.542 atm within 20 to 30 °C , while
153 the used inlet feed flow rate are $5.5556E-5$, $1.111E-4$ and $1.6667E-4$ m³/s respectively.

154

155 **3. Model Rationale and Development**

156 **3.1 Assumptions**

157 A number of reasonable assumptions and simplifications are used in order to develop this
158 model. They include:

- 159 1. The module is made up of porous flat sheet with spacers and negligible leaf curvature.
- 160 2. Validity of the solution-diffusion model for the transport of the solvent and solutes
161 through the membrane.

- 162 3. Validity of the Darcy's law for the feed and permeate channels, which assumes that
 163 the pressure drop is proportional to the feed and permeate flow rate and the friction
 164 parameter is applied to characterize the pressure drop.
- 165 4. Validity of the film model theory to estimate the concentration polarization impact.
- 166 5. The feed osmotic pressure is caused by the impact of all the species found in sugar
 167 and not restricted to only sucrose, glucose and malic acid.
- 168 6. Constant pressure of 1 atm on the permeate side.
- 169 7. Complete mixing in the y-axis of the feed channel due to the existence of a network of
 170 spacers.
- 171 8. The underlying process is assumed to be isothermal.

172

173 **3.2 Model Structure**

174 A computational one dimensional model was developed in this study to predict the variation
 175 of operating parameters and permeate flux in the x-axis during the reverse osmosis
 176 concentration of apple juice at laboratory scale spiral-wound RO process.

177 The details of model development, the equations and the physic-chemical properties of apple
 178 juice used throughout the simulation study are described below.

179 Based on Assumption 2, the solution-diffusion model is valid to predict the permeate flux
 180 through the membrane at any point along the x-axis as expressed by the following equation
 181 (Lonsdale et al., 1965).

$$182 J_{w(x)} = A_w \left(\Delta P_{b(x)} - \Delta \pi_{Total,(x)} \right) \quad (1)$$

183 A_w , $J_{w(x)}$ (m/s, m/ s atm) are the water flux at any point along the x-axis and the membrane
 184 permeability coefficient respectively. A_w was experimentally determined for the spiral-
 185 wound module type (MSCB 2521 R99) using pure water and accounts for the pore
 186 distribution of the membrane, porosity and membrane thickness. Álvarez et al. (2001)
 187 introduce the following correlation to show the impact of feed flow rate and operating
 188 temperature on A_w .

$$189 A_{w,T_b} = 9.059 \times 10^{-7} \left(\frac{T_b}{25} \right)^{0.62} \left(\frac{36.0 \times 10^5 F_{b(0)}}{400} \right)^{-0.1447} \quad (2)$$

190 The above equation confirms that the water permeability coefficient slightly decreased with
 191 inlet feed flow rate and increased with temperature.

192 $(\Delta P_{b(x)} - \Delta \pi_{\text{Total},(x)})$ is the quantity of force per unit area required to handle the osmotic
 193 pressure and release pure water from the feed solution. $\Delta P_{b(x)}$ (atm) is the trans-membrane
 194 pressure defined in Eq. (3).

$$195 \quad \Delta P_{b(x)} = (P_{b(x)} - P_p) \quad (3)$$

196 $P_{b(x)}$, P_p are the feed pressure at any point along the x-axis and the constant permeate pressure
 197 (Assumption 6) respectively. While, the overall trans-membrane pressure (TMP) (atm) for
 198 each run is calculated as:

$$199 \quad TMP = \frac{P_{b(0)} + P_{b(L)}}{2} - P_p \quad (4)$$

200 Where $P_{b(0)}$ and $P_{b(L)}$ (atm) are the pressure into the membrane and the pressure at the outlet
 201 of the membrane respectively.

202 Because the aroma compounds concentration is very small compared to the sugar compounds
 203 in apple juice (Table 1), $\Delta \pi_{\text{Total},(x)}$ can only refer to the summation of the osmotic pressure
 204 difference of sugar compounds along the length of the membrane. The osmotic pressure
 205 difference of each sugar species can be defined as:

$$206 \quad \Delta \pi_{(x)} = \pi_{C_w(x)} - \pi_{C_p(av)} \quad (5)$$

207 $\pi_{C_w(x)}$, $\pi_{C_p(av)}$ (atm) are the osmotic pressure of any sugar compound at the membrane wall
 208 and permeate channel respectively.

209 Eq. (5) can be written as:

$$210 \quad \Delta \pi_{(x)} = R T_b (C_w(x) - C_p(av)) \quad (6)$$

211 C_w and $C_{p(av)}$ (kmol/m³) are the solute concentration of sugar or aroma compounds at any
 212 point along the x-axis at the membrane wall and average solute concentration at the permeate
 213 side respectively. Also, R , T_b (atm L/K mol, K) are the gas constant and the apple juice
 214 temperature respectively.

215 The solute flux $J_{S(x)}$ (kmol/m² s) of any sugar or aroma compounds along the x-axis can be
 216 calculated as:

$$217 \quad J_{S(x)} = B_s (C_w(x) - C_p(av)) \quad (7)$$

218 B_s (m/s) is the solute transport parameter of the determined species (sugar or aroma), which
 219 is assumed as a constant along the length of the membrane and treated as unknown
 220 parameters.

221 The solute flux is lower than volumetric solvent flux, and so $J_{S(x)}$ can be expressed as:

$$222 \quad J_{S(x)} = J_w(x) C_p(av) \quad (8)$$

223 Eq. (6) can be written as:

$$224 \quad \Delta\pi_{(x)} = R T_b \frac{J_s(x)}{B_s} \quad (9)$$

225 Substituting Eq. (8) in Eq. (9) and combining the result in Eq. (1) with re-arrangements yields
226 to Eq. (10).

$$227 \quad J_w(x) = \frac{A_w B_s \Delta P_{b(x)}}{B_s + A_w R T_b C_{p(av)}} \quad (10)$$

228 Based on Assumption 6, Eq. (11) can readily be derived as:

$$229 \quad \frac{d \Delta P_{b(x)}}{dx} = \frac{d P_{b(x)}}{dx} \quad (11)$$

230 Following on, Darcy's law can be used to express the feed pressure drop along the x-axis
231 according to Assumption 3.

$$232 \quad \frac{d P_{b(x)}}{dx} = -b F_{b(x)} \quad (12)$$

233 $F_{b(x)}$, b (m^3/s , $\text{atm s}/\text{m}^4$) are the feed flow rate at any point along the x-axis and the friction
234 parameter respectively. Taking the total mass balance based on the flow rate, gives:

$$235 \quad F_{b(0)} = F_{b(x)} + F_{p(x)} \quad (13)$$

236 $F_{b(0)}$, $F_{p(x)}$ (m^3/s) are the inlet feed flow rate and the permeate flow rate respectively. Also,
237 taking the total mass balance across a small section in the feed channel of the unit gives:

$$238 \quad \frac{d F_{b(x)}}{dx} = -W J_w(x) \quad (14)$$

239 W (m) is the width of the membrane.

240 Furthermore, taking the derivative of Eq. (13) yields Eq. (15) to express the variation of
241 permeated flow rate along the x-axis as:

$$242 \quad \frac{d F_{b(x)}}{dx} = -\frac{d F_{p(x)}}{dx} = W J_w(x) \quad (15)$$

243 Now, Dividing Eq. (12) and Eq. (14), yields:

$$244 \quad \frac{d \Delta P_{b(x)}}{d F_{b(x)}} = \frac{b F_{b(x)}}{W J_w(x)} \quad (16)$$

245 The above equation can be written in the form of Eq. (17) by putting the value of solvent flux
246 from Eq. (10).

$$247 \quad F_{b(x)} d F_{b(x)} = \frac{W A_w B_s \Delta P_{b(x)}}{b (B_s + A_w R T_b C_{p(av)})} d \Delta P_{b(x)} \quad (17)$$

248 Further simplification, yields the following expression:

$$249 \quad F_{b(x)} d F_{b(x)} = \emptyset \Delta P_{b(x)} d \Delta P_{b(x)} \quad (18)$$

$$250 \quad \text{Where, } \emptyset = \frac{W A_w B_s}{b (B_s + A_w R T_b C_{p(av)})} \quad (19)$$

251 ϕ is a parameter that can be calculated for all the sugar and aroma compounds and then the
 252 average value will be considered as $\phi_{(mix)}$ for the rest of calculations.

$$253 \quad \phi_{(mix)} = \frac{\sum_{i=1}^n \frac{W A_w B_s}{b (B_s + A_w R T_b C_p(av))}}{n} \quad (20)$$

254 where, n = Total number of sugar and aroma compounds

255 Re-arrangement with integration of Eq. (18) gives a correlation to calculate the feed flow rate
 256 at any point along the x-axis as follows:

$$257 \quad F_{b(x)} = F_{b(0)} + \phi_{(mix)}^{0.5} \left(\Delta P_{b(x)} - \Delta P_{b(0)} \right) \quad (21)$$

258 Substituting Eq. (21) into Eq. (12) and taking the integration facilitates the calculation of the
 259 trans-membrane pressure in any point along the x-axis.

$$260 \quad \Delta P_{b(x)} = \Delta P_{b(0)} - b x F_{b(0)} - b x \Delta P_{b(x)} \left(\phi_{(mix)} \right)^{0.5} + b x \Delta P_{b(0)} \left(\phi_{(mix)} \right)^{0.5} \quad (22)$$

261 Substituting Eqs. (20) and (22) into Eq. (10) with re-arrangement gives:

$$262 \quad J_{w(x)} = \frac{\phi_{(mix)} b}{W} \left(\Delta P_{b(0)} - b x F_{b(0)} - b x \Delta P_{b(x)} \left(\phi_{(mix)} \right)^{0.5} + b x \Delta P_{b(0)} \left(\phi_{(mix)} \right)^{0.5} \right) \quad (23)$$

263 Also, another equation for solvent flux can be derived by taking the derivative of Eq. (11)
 264 with respect to the x-axis as follows:

$$265 \quad \frac{dJ_{w(x)}}{dx} = \frac{\phi_{(mix)} b}{W} \left(\frac{d\Delta P_{b(x)}}{dx} \right) \quad (24)$$

266 Substituting Eq. (12) into Eq. (24) gives:

$$267 \quad \frac{dJ_{w(x)}}{dx} = \frac{\phi_{(mix)} b}{W} \left(-b F_{b(x)} \right) \quad (25)$$

268 Then, the variation of solvent flux in the x-axis can be calculated by the following equation:

$$269 \quad J_{w(x)} = J_{w(0)} - \left(\frac{\phi_{(mix)}}{W} b^2 x F_{b(0)} \right) + \left(\frac{\phi_{(mix)}^2 b^3}{W} \Delta P_{b(0)} \left(\frac{x^2}{2} \right) \right) - \left(\frac{\phi_{(mix)}^2 b^4}{W} F_{b(0)} \left(\frac{x^3}{6} \right) \right) - \\
 270 \quad \left(\frac{\phi_{(mix)}^{2.5} b^4}{W} \Delta P_{b(x)} \left(\frac{x^3}{6} \right) \right) + \left(\frac{\phi_{(mix)}^{2.5} b^4}{W} \Delta P_{b(0)} \left(\frac{x^3}{6} \right) \right) \quad (26)$$

271 Where, $J_{w(0)}$ (m/s) is the water flux at the inlet edge of the membrane.

272 Here, it is assumed that the osmotic pressure is caused by the impact of all the species found
 273 in sugar (Assumption 5) in contrary to the statement of Álvarez et al. (2001) who neglects
 274 both fructose and sorbitol. Therefore, the solvent flux at $x = 0$ is calculated using Eq. (27)
 275 regarding the osmotic pressure, which is caused by sugar compounds (sucrose, glucose, malic
 276 acid, fructose and sorbitol).

$$277 \quad J_{w(0)} = A_w \left(\Delta P_{b(0)} - \left(\pi_{su(0)} + \pi_{g(0)} + \pi_{m(0)} + \pi_{f(0)} + \pi_{so(0)} \right) \right) \quad (27)$$

278 Where, $\pi_{su(0)}, \pi_{g(0)}, \pi_{m(0)}, \pi_{f(0)}$ and $\pi_{so(0)}$ are the osmotic pressure (atm) of sucrose,
 279 glucose, malic acid, fructose and sorbitol respectively. The estimation of the osmotic pressure
 280 caused by sucrose, glucose and malic acid at any point along the x-axis is carried out using
 281 the empirical equation derived by Nabetani et al. (1992b) as can be seen in Eq. (28).

$$282 \quad \pi_{su(x)} + \pi_{g(x)} + \pi_{m(x)} = -\frac{R T_b}{V_w} \ln \left\{ \frac{\left[\frac{(1000 - C_{w(su)}(x) - C_{w(g)}(x))}{M_{ww}} \right] - \left[\frac{(4 C_{w(su)}(x))}{M_{su}} \right] - \left[\frac{(2 C_{w(g)}(x))}{M_g} \right]}{\left[\frac{(1000 - C_{w(su)}(x) - C_{w(g)}(x))}{M_{ww}} \right] - \left[\frac{(4 C_{w(su)}(x))}{M_{su}} \right] - \left[\frac{(2 C_{w(g)}(x))}{M_g} \right]} \right\} +$$

$$283 \quad \frac{R T_b C_{w(m)}(x)}{M_m} \quad (28)$$

284 While, the contribution of fructose and sorbitol to osmotic pressure is calculated by Eqs. (29)
 285 and (30).

$$286 \quad \pi_{f(x)} = \frac{R T_b C_{w(f)}(x)}{M_f} \quad (29)$$

$$287 \quad \pi_{so(x)} = \frac{R T_b C_{w(so)}(x)}{M_{so}} \quad (30)$$

288 Note, all the concentrations expressed in Eqs. (28), (29) and (30) are referred to the
 289 concentration of the species at the wall membrane and expressed in (kg/m³).

290 The concentration of the sugar and aroma compounds at the wall membrane was estimated
 291 based on Assumption 4, which in turn is based on the validity of the film model theory where
 292 the solvent flux is linked to concentration polarization and mass transfer coefficient, k by the
 293 following equation:

$$294 \quad \frac{(C_{w(x)} - C_{p(av)})}{(C_{b(x)} - C_{p(av)})} = \exp \left(\frac{J_w(x)}{k(x)} \right) \quad (31)$$

295 $C_{b(x)}, k(x)$ (kmol/m³, m/s) are the concentration in the feed channel of any species and the
 296 mass transfer coefficient respectively.

297 The feed velocity along the x-axis $U_{b(x)}$ (m/s) is calculated by:

$$298 \quad U_{b(x)} = \frac{F_{b(x)}}{W t_f} \quad (32)$$

299 The feed pressure equation can be derived from integration of Eq. (12).

$$300 \quad P_{b(x)} = P_{b(0)} - [b F_{b(0)} x] + \left[\phi_{(mix)} b^2 \left(\frac{x^2}{2} \right) \Delta P_{b(0)} \right] - \left[\phi_{(mix)} b^3 F_{b(0)} \left(\frac{x^3}{6} \right) \right] -$$

$$301 \quad \left[\phi_{(mix)}^{1.5} b^3 \Delta P_{b(x)} \left(\frac{x^3}{6} \right) \right] + \left[\phi_{(mix)}^{1.5} b^3 \Delta P_{b(0)} \left(\frac{x^3}{6} \right) \right] \quad (33)$$

302 The sugar or aroma compounds concentration at the feed channel and at any point along the
 303 x-axis is calculated using Eq. (34) as proposed by Lee et al. (2010).

$$304 \quad \frac{d \left(\frac{C_{b(x)} F_{b(x)}}{t_f W} \right)}{dx} = -\frac{J_w(x) c_{p(av)}}{t_f} + \frac{J_w(x) C_{b(x)}}{t_f} + \frac{d}{dx} \left(D_{(x)} \frac{dC_{b(x)}}{dx} \right) \quad (34)$$

305 Then, substituting Eq. (31) and Eq. (8) into Eq. (7) with re-arrangement gives a correlation to
 306 calculate the concentration of any sugar or aroma compound at the permeate side. This
 307 equation will be used twice at $x=0$ and $x=L$ as can be shown in Eqs. (35) and (36), and the
 308 average solute permeate concentration $C_{p(av)}$ (kmol/m^3) is calculated using Eq. (37) as
 309 follows:

$$310 \quad C_{p(0)} = \frac{B_s C_{b(0)} e^{\frac{J_{w(0)}}{k(0)}}}{J_{w(0)} + B_s e^{\frac{J_{w(0)}}{k(0)}}} \quad (35)$$

$$311 \quad C_{p(L)} = \frac{B_s C_{b(L)} e^{\frac{J_{w(L)}}{k(L)}}}{J_{w(L)} + B_s e^{\frac{J_{w(L)}}{k(L)}}} \quad (36)$$

$$312 \quad C_{p(av)} = \frac{C_{p(0)} + C_{p(L)}}{2} \quad (37)$$

313 The volumetric permeated flow rate along the x-axis in the permeate channel can be
 314 calculated using Eq. (38):

$$315 \quad F_{p(x)} = W \int_{x=0}^{x=L} J_w(x) dx \quad (38)$$

316 Also, the model facilitates the evaluation of the performance of the unit by calculating the
 317 rejection of organic compounds as can be seen in the counter of Eq. (39).

$$318 \quad Rej = \frac{C_{b(L)} - C_{p(av)}}{C_{b(L)}} \times 100 \quad (39)$$

319 The recovery of the module calculated using Eq. (40) is the fraction of the feed that is
 320 recovered as permeate at the permeate channel. Note that, for a laboratory scale apparatus,
 321 the permeate flow rate is small in comparison to feed flow rate.

$$322 \quad Rec_{(Total)} = \frac{F_{p(Total)}}{F_{b(0)}} \times 100 \quad (40)$$

323

324 **3.3 The Physical Properties Equations**

325 The mass transfer coefficient is a function of pressure, concentration, flow rate and
 326 temperature, which means that k will vary with the membrane length. Schock and Miquel's
 327 (1987) correlation is used to estimate the mass transfer coefficient along the x-axis for any
 328 species of sugar or aroma compounds as can be depicted in Eq. (41).

$$329 \quad k_{(x)} = 0.065 \left(\frac{D_{(x)}}{d_h} \right) Re_{b(x)}^{0.875} Sc_{(x)}^{0.25} \quad (41)$$

330 $D_{(x)}$, $Re_{b(x)}$, $Sc_{(x)}$ (m^2/s , dimensionless) are the diffusion coefficient of any sugar or aroma
 331 compound, the Reynolds number and the Schmidt number of any sugar or aroma compound
 332 at any point along the x-axis respectively. The terms can be calculated as follows:

333 $Re_{b(x)} = \frac{\rho_{b(x)} d_h U_{b(x)}}{\mu_{b(x)}} \quad (42)$

334 $Sc_{(x)} = \frac{\mu_{b(x)}}{\rho_{b(x)} D_{(x)}} \quad (43)$

335 Where, $\rho_{b(x)}$, $\mu_{b(x)}$ and d_h (kg/m³, kg/m s, m) are the apple juice density, viscosity and the
336 hydraulic diameter respectively.

337 The apple juice viscosity can be calculated as a function of concentration in °Brix and
338 temperature using Eq. (44) (Constenla et al., 1989).

339 $\frac{\mu_{b(x)}}{\mu_w} = \exp\left(\frac{A \text{ } ^\circ\text{Brix}_{(x)}}{100 - B \text{ } ^\circ\text{Brix}_{(x)}}\right) \quad (44)$

340 μ_w and $^\circ\text{Brix}_{(x)}$ are the viscosity of water (8.94E-4 kg/m s) and the concentration of apple
341 juice in °Brix. A and B are parameters related to the temperature and can be estimated using
342 Eqs. (45) and (46).

343 $A = -0.25801 + \frac{817.11}{T_b} \quad (45)$

344 $B = 1.8909 - 3.0212 \times 10^{-3} T_b \quad (46)$

345 T_b is the absolute temperature.

346 Eq. (47) can be used to calculate the variation of apple juice concentration in °Brix along the
347 length of membrane regarding the concentration of the mixture in kg/m³.

348 $^\circ\text{Brix}_{(x)} = 0.099198 \left(\sum_{i=1}^n C_{(x,n)}\right) \quad (47)$

349 where, n = Total number of sugar and aroma compounds

350 Where, $C_{(x)}$ (kg/m³) is the concentration of sugar and aroma compounds at any point along
351 the x-axis and calculated using Eq. (48).

352 $C_{(x,i)} = C_{b(x,i)} M_{wt(i)} \quad (48)$

353 where, i represents the particular species of any sugar or aroma compounds

354 $M_{wt(i)}$ (kg/kmol) is the molecular weight of any species under consideration.

355 The apple juice density is calculated using Eq. (49) as a function of concentration in °Brix
356 and temperature (Constenla et al., 1989).

357 $\rho_{b(x)} = 0.8272 + 0.34708 \exp(0.01 \text{ } ^\circ\text{Brix}_{(x)}) - 5.479 \times 10^{-4} T_b \quad (49)$

358 Then, the diffusion coefficient for any sugar species $D_{SU(x)}$ (m²/s) and aroma compounds
359 $D_{AR(x)}$ (m²/s) along the x-axis can be calculated using the empirical equation proposed by
360 Gladdon and Dole (1953) as can be seen in Eqs. (50) and (51) respectively.

361 $D_{SU(x)} = D_s \left(\frac{\mu_w}{\mu_{b(x)}}\right)^{0.45} \quad (50)$

362
$$D_{AR(x)} = D_a \left(\frac{\mu_w}{\mu_b(x)} \right)^{0.45} \quad (51)$$

363 D_s , D_a (m^2/s) are referred to the diffusion coefficient of any species of sugar and aroma
 364 compounds respectively in a very dilute solution. These coefficients have been calculated
 365 using the proposed correlation of Wilke and Chang (1955).

366
$$D_a = \left(\frac{7.4 \times 10^{-8} (2.6 M_w)^{0.5} (T_b + 273.15)}{(1000 \mu_b(x)) (1000 V_{bp,A})^{0.6}} \right) \times 10^{-4} \quad (52)$$

367 The above equation is correlated to be compatible with the units used. M_w and $V_{bp,A}$
 368 ($kg/kmol$, $m^3/kmol$) are the molecular weight of water and the molar volume of the solute at
 369 its normal boiling point. $V_{bp,A}$ values for all sugar and aroma compounds are shown in Table
 370 1.

371 Finally, the model presented in this section is built within gPROMS (general Process
 372 Modelling System) Model builder, which provides a modelling platform for steady state and
 373 dynamic simulation, optimisation, experiment design and parameter estimation of any
 374 process.

375

376 **4. Determination of Friction and Transport Parameters**

377 **4.1 Determination of Friction Parameter**

378 Unknown friction parameter of the membrane elements and the operating conditions should
 379 be determined before solving the model equations. In the simulation study, experimental data
 380 will be used to predict the best values of unknown feed channel friction parameter. These are
 381 then used with the known parameters to assess the behaviour of the unit with the variance of
 382 operating variables.

383 In this work, the friction parameter has been estimated using an optimization methodology of
 384 the gEST parameter estimation tool developed in gPROMS (Process System Enterprise Ltd.,
 385 2001). This method has been used on the experimental data of Álvarez et al. (2002) in order
 386 to optimize the value of friction parameter. The starting point was based on an initial guess,
 387 which was subsequently used to solve the model equations. The preferred value of the
 388 friction parameter is reached by continuously varying the predicted value until close fit with
 389 experimental data is reached. The registered value of friction parameters for the membrane
 390 type MSCB 2521 R99 of effective area $1.03 m^2$ is $90 (atm s/m^4)$.

391

392

393

394 **4.2 Determination of Solute Transport Parameters**

395 The solute flux of sugar and aroma compounds through the membrane is given by the product
396 of solute transport parameter and the solute concentration difference at the two channels of
397 the unit as expressed in Eq. (7). So, for calculation purposes, a separate value of the solute
398 transport parameter is required for each species for multiple solutes feed.

399

400 **4.2.1 Solute Parameters of Aroma Compounds**

401 The solute parameters of aroma compounds B_s (m²/s) for the reverse osmosis module
402 consisting of a spiral-wound aromatic polyamide membrane type (MSCB 2521 R99) were
403 calculated using the equation of Álvarez et al. (2001).

$$404 B_{s i} = B_{s i, Ref.} \exp^{0.098(T_b - T_{Ref.})} \quad (53)$$

405 i represents the particular species under consideration. $B_{s i}$, $B_{s i, Ref.}$ and $T_{Ref.}$ are the solute
406 parameter of any aroma compounds at operating temperature (T_b) and the reference
407 temperature of 25 °C ($T_{Ref.}$). Eq. (53) was obtained for a temperature range of 15 °C to 30 °C.
408 The estimated values of solute parameter for each aroma compounds at 25 °C are shown in
409 Table 1.

410

411 **4.2.2 Solute Parameters of Sugar Compounds**

412 The solute transport parameters B_s (m²/s) of sugar compounds were calculated using the
413 correlation of Matsuura et al. (1976), which assumed the concept of free energy parameter
414 ($-\Delta\Delta G/RT$) governing non-ionized polar organic solutes in aqueous solution reverse osmosis
415 separation. Eq. (54) shows the general form of this correlation.

$$416 \ln B_{s i} = \ln C_{NaCl}^* + \left(-\frac{\Delta\Delta G}{RT}\right)_i + \delta^* E_s^* \quad (54)$$

417 i represents the particular species under consideration. $\ln C_{NaCl}^*$ is a constant depending on the
418 chemical nature of the membrane and the effective pore size where NaCl as the reference
419 solute. While, the steric Taft number ($\delta^* E_s^*$) is characteristic of each solute in the bulk
420 solution and represents the properties of the solute on the membrane-solution interface and
421 relates to the membrane type.

422 For the aromatic polyamide membrane type, Matsuura et al. (1976) have found the quantity
423 $\ln C_{NaCl}^*$ using the experimental solute transport parameter data $B_{s NaCl}$ for a completely
424 ionized inorganic solute taken NaCl as a reference, and the known values of ($-\Delta\Delta G/RT$) for
425 both Na^+ and Cl^- ions as can be shown in Eq. (55).

$$\ln B_{sNaCl} = \ln C_{NaCl}^* + \left[\left(-\frac{\Delta\Delta G}{RT} \right)_{cation} + \left(-\frac{\Delta\Delta G}{RT} \right)_{anion} \right] \quad (55)$$

Then, the numerical value of $(-\Delta\Delta G/RT)$ for several monovalent inorganic cations and anions in aqueous solutions used in conjunction with aromatic polyamide membrane in reverse osmosis has been obtained using Eq. (56) (Matsuura et al., 1975; Dickson et al., 1975).

$$\ln B_{sNaCl} = \ln C_{NaCl}^* + \left[\left(-\frac{\Delta\Delta G}{RT} \right)_i \right] \quad (56)$$

While, the free energy parameter of each sugar species $(-\Delta\Delta G/RT)$ and the steric Taft number (δ^*Es^*) of each species of sugar has been calculated by Matsuura et al. (1976) and shown in Table 1.

Finally, the transport parameter for each species of sugar for the aromatic polyamide membrane type (MSCB 2521 R99) at 25 °C can be calculated using Eq. (54) as reported in Table 1. However, the transport parameter of malic acid was taken from Malalyandi et al. (1982).

438

439 **5. Model Validation, Results and Discussion**

The model described in Section 3 has been validated by comparing the model predictions results with those obtained from actual experimentation for a MSCB 2521 R99 spiral-wound RO aromatic polyamide membrane module carried out by Álvarez et al. (2002). The comparison between the model predictions and experiments is shown in the following section.

Figure 1 shows the model rejections of two selected aroma compounds, Isopentyl acetate and *trans*-2-hexanal at two different inlet feed flow rates versus the operating temperature and against experimental results. While, Figures 2 and 3 show the experimental and theoretical results of outlet water flux and feed flow rate versus the operating trans-membrane pressure for different inlet feed flow rates at operating temperature 20 °C.

The expectation that increasing inlet feed temperature would increase the solute rejection is validated here as it decreases the viscosity of apple juice as expressed in Eq. (44). This accelerates the flux of water through the membrane and reduces the concentration polarization impact. Interestingly, Figure 1 also shows a slight reduction of Isopentyl acetate and *trans*-2-hexanal rejections with operating temperature for different inlet feed flow rates. The probable explanation for this can be that by increasing the feed temperature, the solute concentration over the membrane wall will increase and causes an increase in solute flux accompanied by the penetrated water that causes an increase in the permeate solute

458 concentration at the permeate channel. As a result, the solute rejection will decrease as
459 expressed in Eq. (39).

460 Furthermore, Figure 1 shows that the model tends to only underestimate the rejection of
461 *trans*-2-hexanal at lower operating temperatures and inlet feed flow rate. This might be
462 attributed to the inaccurate estimation of the transport membrane parameter of *trans*-2-
463 hexanal at such conditions.

464 To illustrate the impact of operating trans-membrane pressure and inlet feed flow rate on
465 solute rejection, Figure 2 shows the variation of Isopentyl acetate rejection versus the
466 operating trans-membrane pressure at three different inlet feed flow rates with comparative
467 data between the model and experiments results. It is expected that the retention of any
468 species will increase due to the increase in operating pressure in turn due to an increase in the
469 water flux passing the membrane. Moreover, the increase of the inlet feed flow rate causes an
470 increase in the Isopentyl acetate rejection due to a reduction in solute flux through the
471 membrane. The increased feed flow rate reduces the wall membrane concentration and causes
472 a decrease of osmotic pressure along the membrane length. Also, an increase in the feed flow
473 rate causes a specific impact on the solute retention by decreasing the amount of accumulated
474 salt on the membrane wall. Consequently, the increasing applied pressure for the same inlet
475 feed flow rate will increase the accumulated salt on the membrane by increasing the water
476 flux.

477 Figure 3 illustrates the effect of operating pressure and inlet feed flow rate in the outlet water
478 flux. The water flux increases due to increase in the operating pressure in line with Eq. (1),
479 which shows that the feed pressure has a substantial impact by bringing up the diffusion rate
480 of water passing through the membrane. Also, it can be noticed that the impact of the inlet
481 feed flow rate is significantly greater at higher operating pressures due to a higher reduction
482 in concentration polarization caused by combining the concurrent impacts of the two feed
483 flow rate and pressure parameters.

484 Similarly, Figure 3 shows that the model predicts the water flux within an accepted error,
485 except at high inlet feed flow rate and operating pressure. This is due to the use of a constant
486 value of water permeability coefficient in all the calculations. It can be argued that this
487 coefficient decreases exponentially with the operating trans-membrane pressure as a result to
488 membrane compaction. At the same time, the water permeability coefficient increases due to
489 an increase in the operating temperature, that causes a reduction in water viscosity. It is
490 expected that these reasons contribute to the slight discrepancy between the outputs of the
491 model and experiments at these conditions.

492 The clear corroboration with experimental data readily shows the suitability of the model to
493 measure the observed retention and water flux parameters with an accepted error over the
494 operating ranges of trans-membrane pressures and temperatures.

495 Figure 4 shows the consistence between the model prediction and experiments results for the
496 outlet feed flow rate versus the operating trans-membrane pressure using three different inlet
497 feed flow rates.

498 Figure 5 shows the effect of operating trans-membrane pressure in the outlet °Brix for
499 different inlet feed flow rates. It is expected that the concentration in °Brix will increase due
500 to an increase in the operating pressure. This is due to the increase in water flux by increasing
501 the operating pressure. The concentration in °Brix that can be obtained is limited to the range
502 10.55 – 11.32 of used pressure and this might be attributed to the use of small specific area of
503 membrane module.

504 It is also interesting to notice that the outlet concentration in °Brix is almost the same for all
505 three inlet feed flow rates at lower inlet operating pressure. However, there is a noted
506 discrepancy at higher operating pressures. Overall, the concentration in °Brix decreases due
507 to an increase in the operating feed flow rate, especially when using higher operating
508 pressures in spite of increasing water flux with increasing inlet feed flow rate, as more
509 specifically illustrated in Figure 3. The reason for this phenomenon is that increasing inlet
510 feed flow rate results in increasing the mass transfer coefficient and decreasing the
511 concentration polarization. Also, the increased feed flow rate reduces the wall membrane
512 concentration and causes a decrease of osmotic pressure, which is followed by decreasing
513 sugar and aroma compounds concentration along the membrane due to a better mixing in the
514 feed channel.

515 Figure 6 displays the variation of operating temperature within the permissible limits of the
516 manufacturer's specifications of the module as a function of apple juice concentration
517 measured in °Brix. It can be observed that the concentration increases as a result to increase
518 in the operating temperature. In line with Eq. (2), the water permeability coefficient increases
519 with increasing the operating temperature, which causes an increase in water flux that raises
520 the apple juice concentration in the feed side.

521 Finally, a 10% maximum error agreement is obtained in a comparison between experimental
522 and calculated rejection for all aroma compounds as presented in Figure 7. The relatively
523 small discrepancy can be attributed to two reasons. Firstly, the actual experiments of Álvarez
524 et al. (2002) is carried out using apple juice concentration of 11 °Brix not 10.5 °Brix.

525 Secondly, constant solute transport parameters of aroma compounds are used in the
526 calculation of solute flux through the membrane as can be seen in Eq. (7).

527

528 **Conclusions**

529 A mathematical one dimensional steady model applicable for apple juice concentration
530 process using a spiral-wound reverse osmosis process has been developed with a simulation
531 study of permeate flux and aroma compounds rejection. The model can predict the variation
532 of the feed flow rate, sugar and aroma compounds concentration in both the feed and
533 permeate channels, membrane wall concentration, feed pressure and water and solute fluxes
534 in each point along the membrane length. A number of differential equations have been
535 developed based on the solution-diffusion model, which takes into account the impact of all
536 sugar species in the calculation of osmotic pressure. Besides, the model estimates the
537 physical properties of apple juice using the empirical equations that shows the impact of
538 concentration and temperature derived from the literature. Also, the solute transport
539 parameters of sugar species were determined based on the concept of free energy parameter.
540 The model has been validated against an experimental data set derived from an apple juice
541 concentration process and shows accepted relative errors between theoretical and
542 experimental results for most operating parameters. The model has been used for further
543 simulation to study the influence of various operating conditions on permeate flux and aroma
544 compounds rejection. Further work is planned to optimize the apple juice concentration and
545 aroma compound retention by assessing the impact of module area and operating variables.

546 **References**

547 Álvarez V., Álvarez S., Riera F., and Álvarez R., 1997. Permeate flux prediction in apple
548 juice concentration by reverse osmosis. *Journal of Membrane Science*, 127, 25-34.

549 Álvarez S., Riera F., Álvarez R. and Coca J., 1998. Permeation of apple juice compounds in
550 reverse osmosis. *Separation Purification Technology*, 14, 209-220.

551 Álvarez S., Riera F., Álvarez R. and Coca J., 2001. Prediction of flux and aroma compounds
552 rejection in a reverse osmosis concentration of apple juice model solutions. *Ind. Eng.
553 Chem. Res.*, 40, 4925-4934.

554 Álvarez S., Riera F., Álvarez R. and Coca J., 2002. Concentration of apple juice by reverse
555 osmosis at laboratory and pilot-plant scales. *Ind. Eng. Chem. Res.*, 41, 6156-6164.

556 Cheong K. W., Tan C. P., Mirhosseini H., Abdul Hamid N. S., Osman A. and Basri M., 2010.
557 Equilibrium headspace analysis of volatile flavor compounds extracted from soursop
558 (*Annona muricata*) using solid-phase microextraction. *Food Research International*,
559 43(5), 1267-1276.

560 Dickson J. M., Matsuura T., Blais P. and Sourirajan S., 1975. Reverse osmosis separation in
561 some organic and inorganic solutes in aqueous solutions using aromatic polyamide
562 membranes. *Journal of Applied Polymer Science*, 19, 801-819.

563 Gladdon J. K. and Dole M., 1953. Diffusivity determination of sucrose and glucose solutions.
564 *J. Am. Chem. Soc.*, 75, 3900-3904.

565 Constenla D. T., Lozano J. E. and Crapiste G. H., 1989. Thermo-physical properties of
566 clarified apple juice as a function of concentration and temperature. *J. Food Sci.*, 54(3),
567 663-668.

568 Girard B. and Fukumoto L. R., 2000. Membrane processing of fruit juices and beverages: A
569 review. *Critical Reviews in Food Science and Nutrition*, 40(2), 91-157.

570 Jain S. and Gupta S. K., 2004. Analysis of modified surface force pore flow model with
571 concentration polarization and comparison with Spiegler–Kedem model in reverse
572 osmosis systems. *Journal of Membrane Science*, 232(1-2), 45-62.

573 Jiao B., Cassano A. and Drioli E., 2004. Recent advance on membrane processes for the
574 concentration of fruit juices: A review. *Journal of Food Engineering*, 63, 303-324.

575 Lee C., Chen Y. and Wang G., 2010. A dynamic simulation model of reverse osmosis
576 desalination systems. *The 5th International Symposium on Design, Operation and
577 Control of Chemical Processes, PSE Asia, Singapore.*

578 Lonsdale H. K., Merten U. and Riley R. L., 1965. Transport properties of cellulose acetate
579 osmotic membranes. *Journal of Applied Polymer Science*, 9, 1341-1362.

580 Malalyandi P., Matsuura T. and Sourirajan S., 1982. Predictability of membrane performance
581 for mixed solute reverse osmosis systems. System cellulose acetate membrane-D-
582 Glucose-D,L-Malic acid-water. *Ind. Eng. Chem. Process. Dev.*, 21, 277-282.

583 Matsuura T., Bednas M. E., Dickson J. M. and Sourirajan S., 1974. Polar and steric effects in
584 reverse osmosis. *Journal of Applied Polymer Science*, 18, 2829-2846.

585 Matsuura T., Bednas M. E., Dickson J. M. and Sourirajan S., 1975. Reverse osmosis
586 separations of aldehydes, ketones, and ethers in aqueous solutions using porous
587 cellulose acetate membranes. *J. Appl. Polym. Sci.*, 19(9), 2473-2484.

588 Matsuura T., Dickson J. M. and Sourirajan S., 1976. Free energy parameters for reverse
589 osmosis separations of undissociated polar organic solutes in dilute aqueous solutions.
590 *Ind. Eng. Chem. Process Des. Dev.*, 15(1), 149-161.

591 Nabetani H., Nakajima M., Watanabe A., Ikeda S., Nakao S. and Kimura S., 1992a.
592 Development of a new type of membrane osmometer. *Journal of Chemical Engineering*
593 *of Japan*, 25, 269-274.

594 Nabetani H., Nakajima M., Watanabe A., Nakao S. and Kimura S., 1992b. Prediction of the
595 flux for the reverse osmosis of a solution containing sucrose and glucose. *Journal of*
596 *Chemical Engineering of Japan*, 25(5), 575-580.

597 Pereira E. N., Matsuura T. and Sourirajan S., 1976. Reverse osmosis separations and
598 concentrations of food sugars. *J. Food Sci.*, 41, 672.

599 Pepper D., 1990. RO for improved products in the food and chemical industries and water
600 treatment. *Desalination*, 77, 55-71.

601 Pozderovic´ A., Moslavac T. and Pichler A., 2006. Concentration of aqueous solutions of
602 organic components by reverse osmosis II. Influence of transmembrane pressure and
603 membrane type on concentration of different alcohol solutions by reverse osmosis.
604 *Journal of Food Engineering*, 77, 810-817.

605 Process System Enterprise Ltd, 2001. gPROMS Introductory User Guide. Process System
606 Enterprise Ltd., London.

607 Sheu M. and Wiley R., 1983. Preconcentration of apple juice by reverse osmosis. *Journal of*
608 *Food Science*, 48(2), 422-429.

609 Schock G. and Miquel A., 1987. Mass transfer and pressure loss in spiral wound modules.
610 *Desalination*, 64, 339-352.

611 Wilke C. R. and Chang P., 1955. Correlation of diffusion coefficients in dilute solutions.
612 *A.I.Ch.E. Journal*, 1(2), 264-270.

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615

616 Table 1: Characteristics of the sugar and aroma compounds and their inlet concentration in the model solution of
 617 10.5 °Brix (Matsuura et al., 1976; Álvarez et al., 1998; Malalyandi et al., 1982; Álvarez et al., 2002)

Compound	Molecular weight M_{wt} , (kg/kmol)	Concentration $C_{b(0)}$, (kmol/m ³)	Molar volume, $V_{bp,A}$ (m ³ /kmol)	Free energy parameter, $(-\frac{\Delta\Delta G}{RT})_{25\text{ }^{\circ}C}$	Steric Taft number, $(\delta^*Es^*)_{25\text{ }^{\circ}C}$	Solute transport parameter, $B_{s,25\text{ }^{\circ}C}$ (m/s)
sucrose	342	0.035555	0.215689	-1.76	-7.42	2.32996E-10
glucose	180	0.138000	0.116987	1.81	-5.42	6.11461E-8
malic acid	134	0.029104	0.083337	---	---	5.40E-8
fructose	180	0.340722	0.106351	1.59	-5.56	4.26602E-8
sorbitol	182	0.018406	0.122343	1.82	-5.57	5.31579E-8
ethyl acetate	88.11	0.000566	0.097683	2.11	-0.07	4.818E-6
ethyl butanoate	116.16	0.000129	0.132150	1.54	-0.43	1.739E-6
ethyl-2-methyl butanoate	130.19	5.37E-05	0.150508	1.47	-1.20	0.223E-6
isopentyl acetate	130.19	0.000130	0.148618	1.47	-0.35	0.387E-6
Hexyl acetate	144.22	6.926E-05	0.166274	1.85	-0.40	1.564E-6
trans-2-hexenal	98.143	0.000712	0.116004	---	---	4.574E-6
hexanal	100.2	0.000149	0.123095	2.19	-0.40	2.084E-6
isobutanol	74.12	0.000269	0.092421	2.42	-0.93	0.302E-6
butanol	74.12	0.000269	0.091506	2.17	-0.39	1.905E-6
isopentanol	88.15	0.000169	0.108771	2.12	-0.35	0.297E-6
hexanol	102.18	0.000293	0.125522	2.81	-0.40	1.556E-6

618

619 Table 2: Specifications of the spiral-wound membrane element (Álvarez et al., 2002)

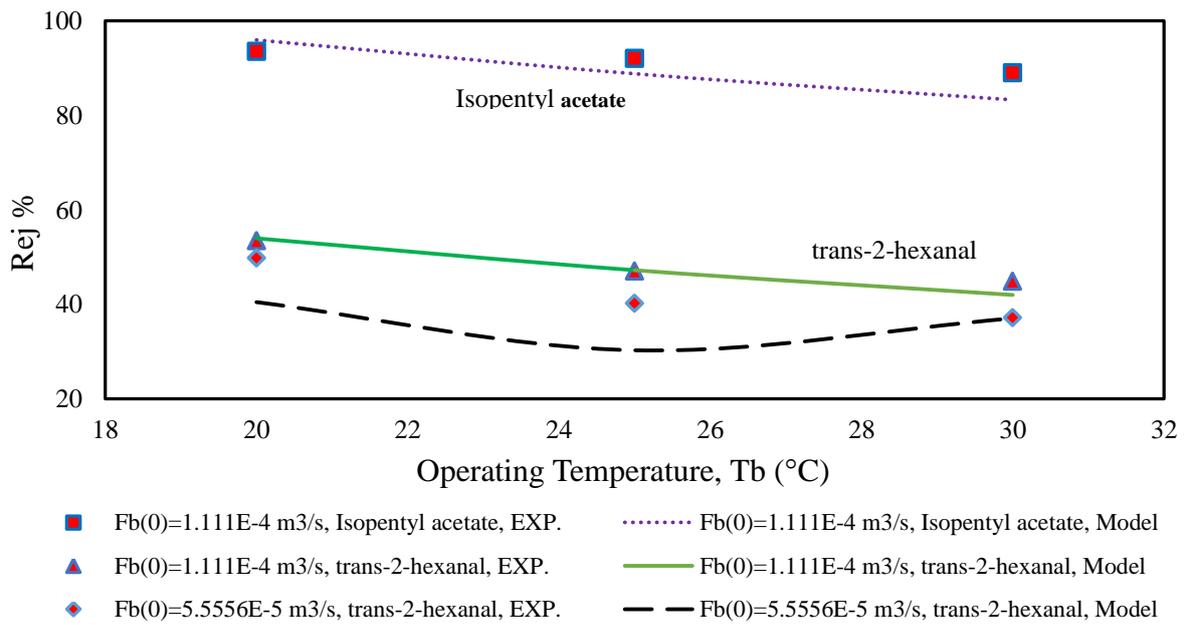
<i>Make</i>	<i>Sparem Spa. (Biella, Italy)</i>
Membrane type and configuration	MSCB 2521 R99, Spiral-wound, Polyamide membrane
Active surface area (m ²)	1.03
Feed and permeate spacer thickness (t_f) and (t_p) (m)	0.0007 and 0.00055
Membrane sheet length (L) and width (W) (m)	0.44 and 2.3409
Hydraulic diameter (m)	0.00096
Max. operating pressure (atm)	41.4508
Max. operating temperature (°C)	50

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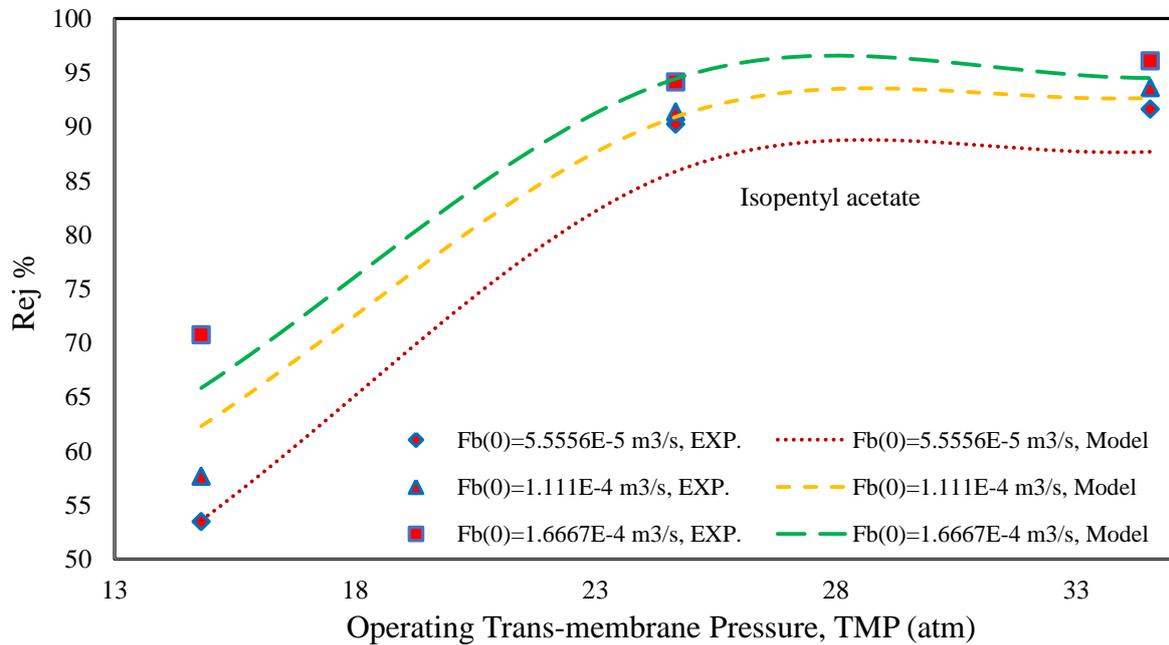


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625 Figure 1: Experimental and model rejections of the two selected aroma compounds versus average operating
 626 temperature for two different inlet feed flow rates at inlet conditions ($^{\circ}\text{Brix} = 10.5$, $TMP = 34.542$ atm)

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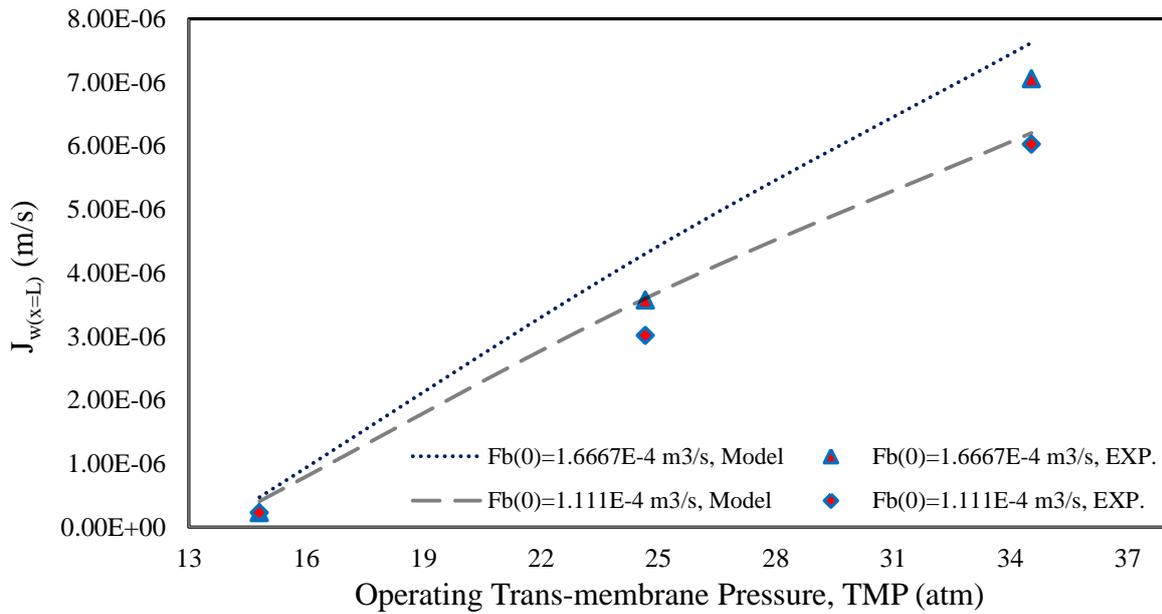
630 Figure 2: Experimental and model Isopentyl acetate rejection versus operating trans-membrane pressure for
 631 three different inlet feed flow rates at inlet conditions ($^{\circ}\text{Brix} = 10.5$, $T_b = 20$ $^{\circ}\text{C}$)

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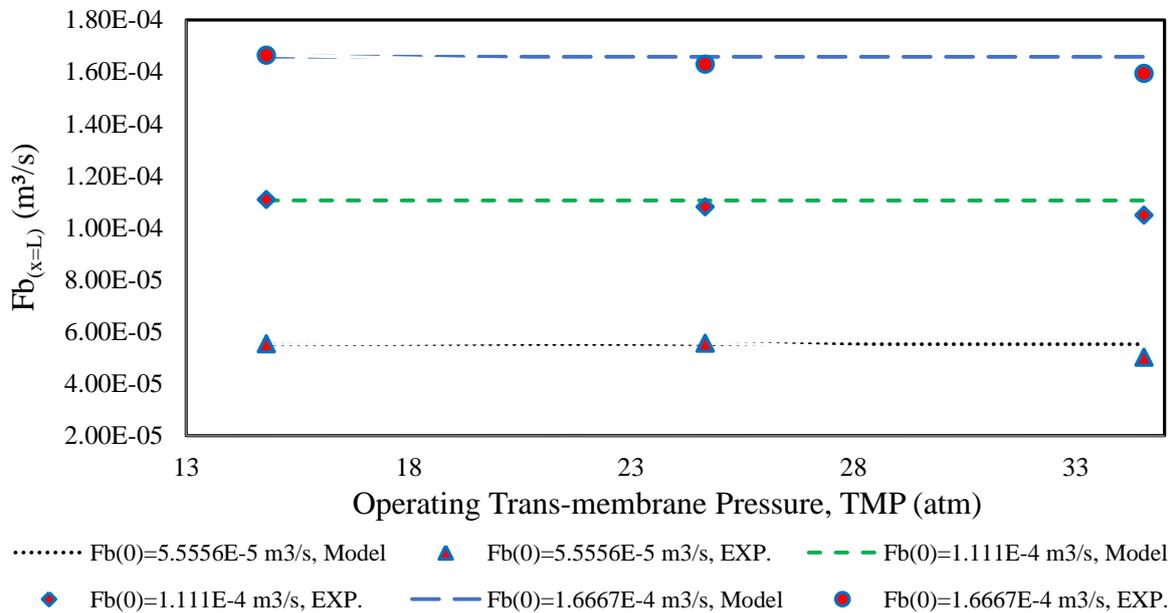
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637 Figure 3: Experimental and model outlet water flux versus operating trans-membrane pressure for two different
638 inlet feed flow rates at inlet conditions ($^{\circ}$ Brix = 10.5, $T_b = 20$ $^{\circ}$ C)
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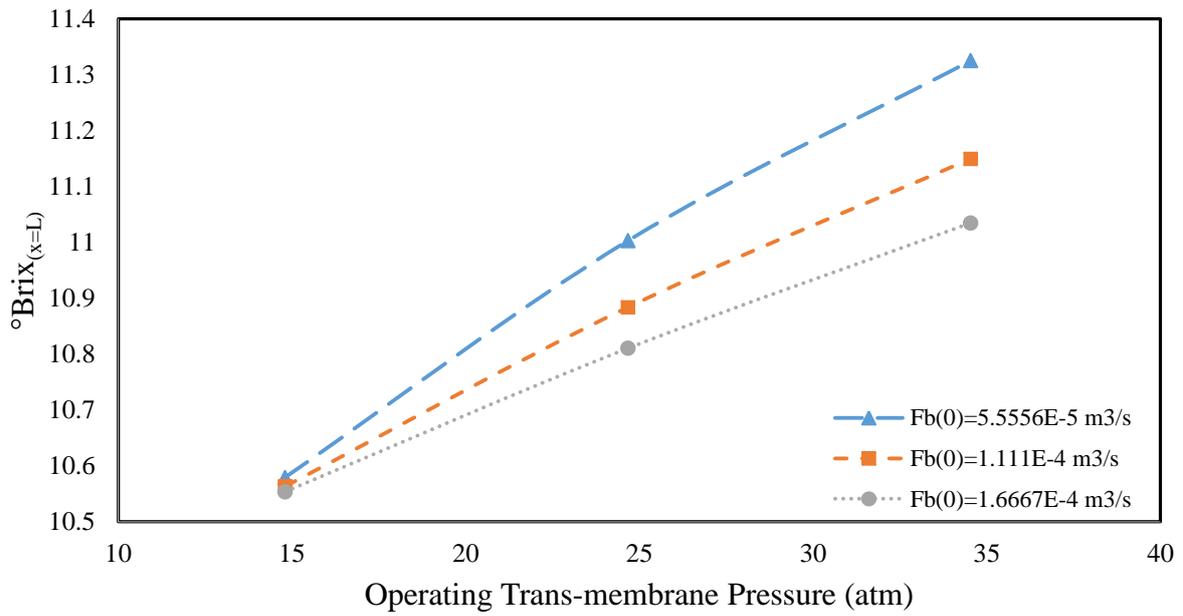
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642 Figure 4: Experimental and model outlet feed flow rate versus operating trans-membrane pressure for three
643 different inlet feed flow rates at inlet conditions ($^{\circ}$ Brix = 10.5, $T_b = 20$ $^{\circ}$ C)
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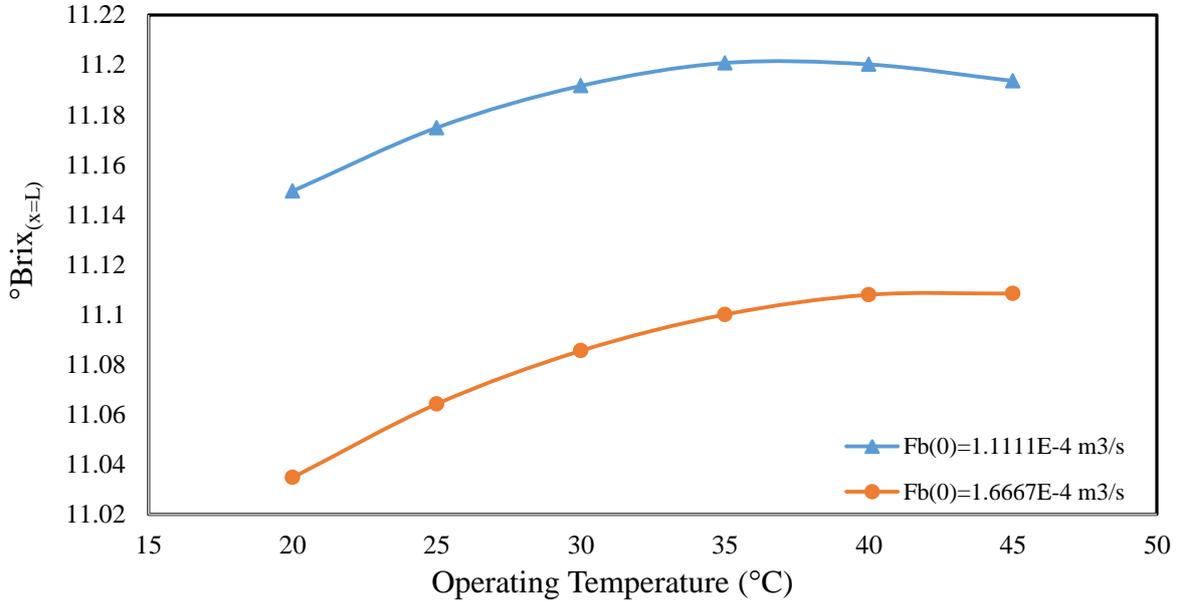


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649 Figure 5: Outlet Brix variation as a function of operating trans-membrane pressure at different inlet feed flow
650 rates at inlet conditions ($^{\circ}\text{Brix} = 10.5, T_b = 20^{\circ}\text{C}$)
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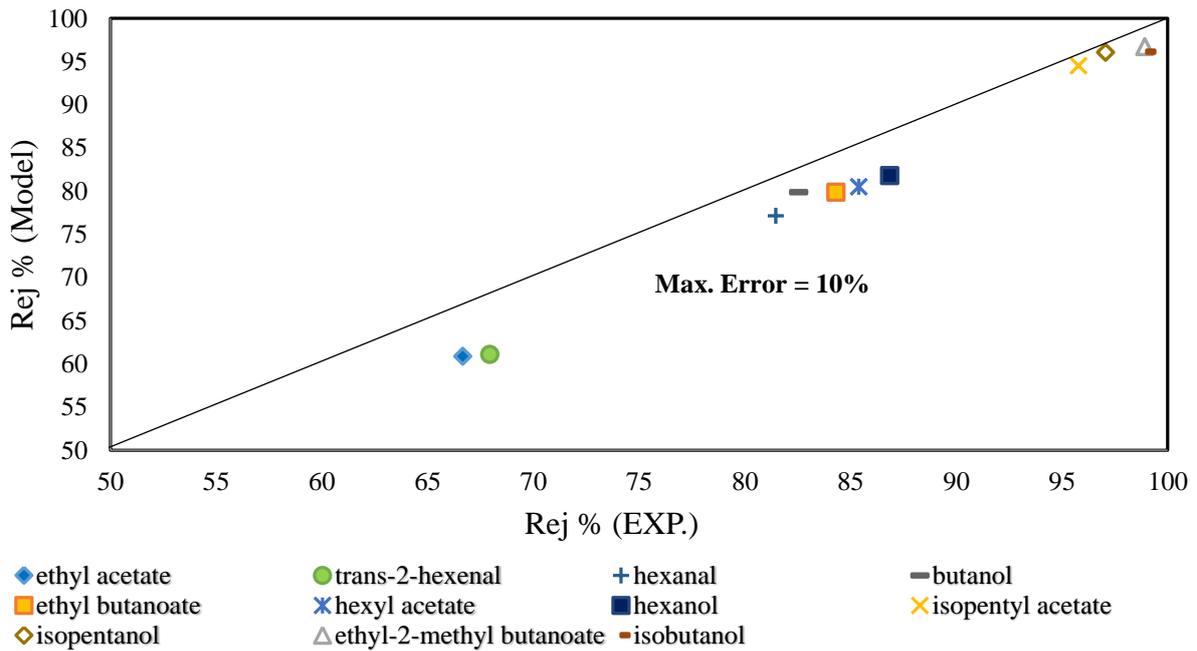
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654 Figure 6: Outlet Brix variation as a function of operating temperature at different inlet feed flow rates at inlet
655 conditions ($^{\circ}\text{Brix} = 10.5, TMP = 34.542\text{ atm}$)
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660

661 Figure 7: Experimental and predicted aroma compounds rejection at inlet conditions ($^{\circ}\text{Brix} = 10.5, T_b = 20\text{ }^{\circ}\text{C},$
 662 $TMP = 34.542\text{ atm}, F_{b(0)} = 1.6667\text{E-}4\text{ m}^3/\text{s}$)

663

664