

1 **The effect of photoperiod and high fat diet on the cognitive response in photoperiod-**
2 **sensitive F344 rats**

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4 Samantha L. McLean^a, Haesung Yun^b, Andrew Tedder^b and Gisela Helfer^{b*}

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6 ^aSchool of Pharmacy and Medical Sciences, Faculty of Life Sciences, University of Bradford,
7 Richmond Road, Bradford, BD7 1DP, UK

8 ^bSchool of Chemistry and Bioscience, Faculty of Life Sciences, University of Bradford,
9 Richmond Road, Bradford, BD7 1DP, UK

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11 **Running title:** Seasonal rhythms in cognition

12 ***Corresponding author:**

13 Gisela Helfer

14 School of Chemistry and Bioscience, University of Bradford

15 Richmond Road

16 Bradford

17 BD7 1DP, UK

18 E-mail: g.helfer@bradford.ac.uk

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20 **Highlights**

- 21 • **Diet and photoperiod have an impact on object recognition in photoperiod-sensitive**
22 **F344 rats.**
- 23 • Rats in long photoperiod on chow diet can perform a novel object recognition task
24 suggesting their short-term memory remains intact.
- 25 • **Our behavioral analysis suggests** that high fat diet induces an impairment in memory
26 independent of photoperiod.
- 27 • Changing rats to short photoperiod conditions impairs their ability to differentiate
28 between normal and familiar objects independent of diet.

29

1 **Abstract**

2 In many species, seasonal changes in day length (photoperiod) have profound effects on
3 physiology and behavior. In humans, these include cognitive function and mood. Here we
4 investigated the effect of photoperiod and high fat diets on cognitive deficits, as measured by
5 novel object recognition, in the photoperiod-sensitive F344 rat, which exhibits marked natural
6 changes in growth, body weight and food intake in response to photoperiod. 32 male juvenile
7 F344 rats were housed in either long or short photoperiod and fed either a high fat or
8 nutrient-matched chow diet. Rats were tested in the novel object recognition test before
9 photoperiod and diet intervention and re-tested 28 days after intervention. In both tests
10 during the acquisition trials there was no significant difference in exploration levels of the left
11 and right objects in the groups. Before intervention, all groups showed a significant increase
12 in exploration of the novel object compared to the familiar object. However, following the
13 photoperiod and diet interventions the retention trial revealed that only rats in the long
14 photoperiod-chow group explored the novel object significantly more than the familiar object,
15 whereas all other groups showed no significant preference. These results suggest that
16 changing rats to short photoperiod impairs their memory regardless of diet. The cognitive
17 performance of rats on long photoperiod-chow remained intact, whereas the high fat diet in
18 the long photoperiod group induced a memory impairment. **In conclusion, our study suggests**
19 **that photoperiod and high fat diet have an impact on object recognition in photoperiod-**
20 **sensitive F344 rats.**

21

22 **Key words:** photoperiod, seasonal, rhythms, F344, cognition, high fat diet, novel object
23 recognition

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1 Introduction

2 Seasonal animals have evolved diverse seasonal variations in physiology and behavior to
3 accommodate yearly changes in environmental and climatic conditions. These physiological
4 and behavioral changes are initiated by changes in day length (photoperiod) and include
5 annually occurring phenomena such as migration, hibernation, torpor and reproduction
6 (Helfer et al., 2019). The broad importance of seasonality in physiology and biomedical
7 research has increasingly been recognized in recent years (Stevenson et al., 2015).
8 Seasonal animals undergo pronounced cycles of weight gain and weight loss to precisely
9 control their energy stores as part of their natural physiology. Unlocking the basis of seasonal
10 body weight regulation is therefore important not only to our understanding of basic
11 physiology, but to understand the long-term impact of seasonal changes on the brain,
12 behavior and cognition in animals, including humans (Helfer and Dumbell, 2020).

13 The neuroendocrine mechanisms underlying seasonal energy balance regulation and
14 reproduction involve dynamic interactions across multiple central nervous system substrates
15 and hormonal messengers to provide system-wide orchestration (Helfer and Stevenson,
16 2020). Seasonal changes in photoperiod are mediated through the nocturnal secretion of
17 melatonin from the pineal gland which relays photoperiodic information to the pars tuberalis
18 of the pituitary gland to regulate the release of thyroid-stimulating hormone (TSH). In short
19 photoperiod (winter: short days and long nights), the increased duration of melatonin signal
20 inhibits the release of TSH whereas in long photoperiod (summer: long days and short
21 nights), the short duration of melatonin allows TSH release. The hypothalamus integrates the
22 TSH signal by increasing the expression of deiodinase enzymes to catalyze the conversion
23 of inactive thyroid hormone T4 to biological active thyroid hormone T3. Increased T3
24 regulates key downstream pathways, such as retinoic acid signaling, resulting in appropriate
25 seasonal phenotypes (Helfer et al., 2019; Helfer and Dumbell, 2020). This process also
26 involves the photoperiodic regulation of neuropeptides localized in discrete appetite-
27 regulating centers of the hypothalamus to regulate food intake and body composition (Helfer
28 and Stevenson, 2020).

29 To study the interactions of the mechanisms involved in the regulation of body weight and
30 growth with those involved in diet-induced obesity, we previously investigated the effect of
31 photoperiod and high fat diet on physiology (body weight, body composition, food intake),
32 circulating levels of hormones that regulate feeding status and hypothalamic gene
33 expression in Fisher F344 rats (Ross et al., 2015). The study showed that photoperiod and
34 high fat diet regulate body weight and body composition through independent pathways, with
35 photoperiod primarily effecting growth and lean mass accretion and high fat diet effecting
36 adipose tissue accretion. **Anecdotal findings from this study suggested that photoperiod and**

1 high fat diet had an effect on rats' behavior, insofar we noticed that animals housed in long
2 photoperiod demonstrated increased explorative behavior and seemed generally more alert
3 (unpublished data). It is well established that high fat diet impairs cognitive function and
4 induces cognitive deficits in rodent models (Buie et al., 2019; Del Olmo and Ruiz-Gayo, 2018;
5 Kanoski and Davidson, 2011; McLean et al., 2018; Noble and Kanoski, 2016). Much less
6 common are reports on the effect of photoperiod on cognitive performance, yet seasonal
7 environmental changes are expected to influence cognition to meet the ecological needs of
8 animals (Buchanan et al., 2013).

9 The best studied examples of seasonal changes in cognitive processes come from songbirds
10 where seasonal changes in song production and learning are accompanied by changes in
11 the brain regions controlling song (Tramontin and Brenowitz, 2000). Interestingly, a recent
12 study has shown that these seasonal changes are not under photoperiod control (Pozner et
13 al., 2018). Additionally, the hippocampus undergoes seasonal changes in food-storing birds
14 and brood parasites, but similarly this seems independent of photoperiod (Sherry and
15 MacDougall-Shackleton, 2015). In some seasonal breeders, such as cowbirds, deer mice
16 and voles, spatial memory is enhanced prior to the breeding season (Clayton et al., 1997;
17 Galea et al., 1996; Galea et al., 1994). Male African striped mice show increased spatial
18 performance and attention during winter and it has been suggested that this might be due to
19 greater dispersal motivation before breeding season (Maille et al., 2015). Interestingly,
20 female striped mice do not show the same seasonal variation (Maille et al., 2015; Maille and
21 Schradin, 2016). These studies provide compelling evidence for seasonal patterns of
22 cognitive performance, however, studies designed to examine photoperiod control of
23 cognition are limited.

24 Interestingly, in non-seasonal animals, for example the non-photoperiodic Wistar rat, there is
25 some evidence indicating that photoperiod has an impact on animal behavior, including
26 depression, anxiety and stress (Barnes et al., 2017). This provides an exciting opportunity for
27 novel treatment strategies using photoperiod that may be applicable to humans. Indeed,
28 short photoperiod has been suggested as an approach in the management of central
29 nervous system injuries (Subhadeep et al., 2020) and therapeutic interventions targeting
30 photoperiodic regulated dopamine signaling could help patients suffering from seasonal
31 affective disorders (Okimura et al., 2021).

32 Here we investigated the effect of photoperiod and diet on cognition in the photoperiod-
33 sensitive F344 rat. The inbred F344 rat strain is one of the few rat strains that shows
34 pronounced photoperiod-induced changes in its metabolic phenotype, growth and
35 reproductive status (Tavolaro et al., 2015). In the laboratory, a simple switch in photoperiod
36 induces large scale changes in body composition, food intake, reproduction and

1 hypothalamic gene expression within 2-4 weeks (Helfer et al., 2013; Helfer et al., 2012;
2 Helfer et al., 2016; Ross et al., 2015). Furthermore, the F344 rat strain is one of the strains
3 preferred in behavioral tests due to low level activation of the hypothalamic-pituitary-adrenal
4 axis and low open field defecation (Glowa et al., 1992; Harrington, 2013; Van Der Staay and
5 Blokland, 1996). Thus, the F344 rat is the ideal model to study photoperiod regulation of
6 cognitive flexibility. To test photoperiod control of cognition, we used the novel object
7 recognition (NOR) task, a spontaneous and ethologically relevant behavioral paradigm that is
8 based on rodents' natural tendency to explore novel stimuli and environments (Ennaceur and
9 Delacour, 1988). This is a robust, well-characterized behavioral task which is routinely used
10 to assess cognition and natural behavior in an open field arena (Cohen and Stackman, 2015;
11 Grayson et al., 2015; McLean et al., 2016; McLean et al., 2017). In line with previous studies,
12 we predicted that high fat diet would decrease cognitive performance in F344 rats
13 independent of photoperiod. Given that F344 rats breed during long photoperiod (Tavolaro et
14 al., 2015), we hypothesized that cognitive performance would be enhanced in long
15 photoperiod in these rats.

16 **Methods**

17 **Ethics statement**

18 All animal procedures were performed according to the Animals (Scientific Procedures) Act,
19 1986 and approved by the Animal Welfare Ethical Review Body at the University of Bradford.
20 Animal experiments were licensed by the UK Home Office under the project license number
21 P0D6AA50D.

22 **Animal experiment**

23 32 male Fischer F344/NHsd (F344) rats from barrier 208A were obtained from Envigo (Oxon,
24 UK) at 4-5 weeks old (weight range 75-100g). Initially, rats were acclimatized for 10 days
25 under 12h light:12h dark photoperiod in groups of four with *ad libitum* access to water and
26 standard chow diet (2018 Teklad global 18% protein rodent diet, Envigo). Rats were then
27 randomly assigned to four groups containing eight rats each and transferred to different
28 photoperiod rooms (Table 1). Group sizes were selected based on power calculations
29 performed on results from previous studies (Ross et al., 2015; Tavolaro et al., 2015). Two
30 groups (n=8/group) were switched to a short photoperiod (SP; 8h light:16h dark) and two
31 groups were switched to a long photoperiod (LP; 16h light:8h dark). Photoperiods were
32 changed by intermittently shortening or lengthening the light-off time with the original light-on
33 time remaining the same in all rooms. One group in each photoperiod room was provided *ad*
34 *libitum* access to water and either a high fat diet (45% fat by kcal, TD.06415, Envigo) or a
35 nutrient-matched chow diet (10% fat by kcal, TD.06416, Envigo) (Figure 1). In photoperiod
36 rooms, rats were housed in pairs to avoid effects of isolation on behavior (McLean *et al.*,

1 2010). Rats were housed in standard rat cages (Type III rat caging, 2017cm² floor area,
2 Arrowmicht, Hereford, UK) with soft woodchip bedding (EC06 chips, Datesand, Manchester,
3 UK) and a red plastic tunnel and shredded paper (Sizzle Nest, Datesand) for enrichment.
4 Apart from photoperiod, all other environmental conditions were kept constant with a
5 temperature of 21±1°C, humidity of 50±5% and average light intensity of 150 lux. Body
6 weight of individual rats and food intake of the cage was measured three times weekly within
7 two hours after lights on. Food intake of the cage was measured by subtracting the
8 remaining amount of pellets in the dispenser and cage from the pre-weighed amount
9 provided. Food intake of the cage was then divided by two to obtain food intake values for
10 each individual rat. Health checks were carried out twice daily and no welfare-related issues
11 were observed.

12 After 28 days under experiment conditions, rats were assigned random numbers before
13 killing to ensure blind analyses to grouping. Rats were killed by terminal anesthesia using
14 isoflurane followed by decapitation at Zeitgeber Time 3 (=3h after lights on). Testes were
15 dissected and paired testes weight was recorded to confirm photoperiod responses
16 (Tavolaro et al., 2015).

17 **Behavioral testing**

18 A total of 32 rats were tested in the NOR task as described in detail previously (McLean et al.,
19 2011). Briefly, rats were habituated to the test box for 20 min on three consecutive days.
20 Following a 3 min habituation session on the day of testing each rat was placed in the NOR
21 chamber (52 cm wide × 40 cm high × 52 cm long) and exposed to two identical objects for a
22 period of 3 min. The rats were then returned to their home cage for an inter-trial interval (ITI)
23 of 1 min; the box was cleaned with 70% ethanol, both objects removed, and one replaced
24 with an identical familiar copy and one with a novel object. Following the ITI, rats were
25 returned to explore the familiar and a novel object in the test box for a 3 min retention trial.
26 The location of the novel object in the retention trial was randomly assigned for each rat
27 using a Gellerman schedule. All experiments were video recorded (Home Guard - CCTV
28 Home Security Kit) for subsequent behavioral analysis by an experimenter blind to the
29 treatments. Exploration time (sec) for each object in each trial was recorded manually using
30 two stopwatches. Locomotor activity was recorded by scoring the number of line crossings
31 by the rat in both acquisition and retention trials (Figure 1).

32 Rats were then randomly assigned to one of the 4 intervention groups (as described above,
33 Figure 1) and were re-tested after 4 weeks in the NOR task. Care was taken to use new
34 objects for the second experiment to ensure rats still had a natural propensity to explore
35 based on novelty.

36 **Statistics**

1 Body weight, food intake and energy intake data were analyzed by three-way repeated
2 measures (RM) ANOVA (photoperiod x diet x time interaction) followed by Tukey's multiple
3 comparisons test. Testes weight were analyzed by one-way ANOVA followed by Tukey's
4 honestly significant difference post-hoc test for pairwise comparison. The NOR data are
5 expressed as mean exploration time \pm SD. Data passed normality (Shapiro-Wilk) and
6 student's two-tailed paired t-test was performed to compare time spent exploring the familiar
7 versus the novel object. The locomotor activity and total exploration levels were analyzed
8 using one-way ANOVA followed by Tukey's post-hoc test. Differences were considered
9 statistically significant if $P < 0.05$. Data are presented as mean \pm SD, n refers to the number
10 of animals.

11 Results

12 The effect of photoperiod and high fat diet on body weight, food and energy intake and 13 paired testes mass

14 A three way RM ANOVA revealed a significant interaction between photoperiod, diet and
15 time for body weight ($F_{(12,336)}=2.276$; $P = 0.009$) comparable to our previous study (Ross et
16 al., 2015). After 28 days of high fat feeding, body weight of rats on short photoperiod was
17 11% lower relative to rats on long photoperiod ($P = 0.006$; Figure 2A). The difference
18 between chow fed rats was less pronounced with body weight of rats on short photoperiod
19 being 7% lower compared to rats on long photoperiod ($P = 0.075$). This might be due to one
20 rat in the long photoperiod-chow group failing to respond to photoperiod and gaining less
21 weight than the remainder of the group, however Grubb's test did not reveal this rat as an
22 outlier. The effects of photoperiod and high fat diet on testes size were in proportion to body
23 weight as previously shown (Tavolaro et al., 2015). One-way ANOVA revealed a significant
24 difference in paired testes weight ($F_{(3,28)}=5.166$, $P = 0.0057$; Figure 2B). A pairwise
25 comparison showed that paired testes weight of short photoperiod-HF and short photoperiod-
26 chow rats was significantly lower compared to long photoperiod-HF rats (Tukey's test, $P =$
27 0.009 and $P = 0.013$, respectively). Consistent with the difference in body weights, three-way
28 RM ANOVA showed a significant interaction between photoperiod, diet and time for food
29 intake ($F_{(3,84)}=10.31$; $P < 0.001$) and energy intake ($F_{(3,84)}=9.335$; $P < 0.001$). Food intake was
30 highest in the long photoperiod-chow rats and these intakes were higher than the intakes of
31 the long photoperiod-HF rats (Tukey's test, $P < 0.001$) (Figure 2C). In addition, food intake of
32 chow diet rats was significantly higher than high fat diet rats independent of photoperiod (LD:
33 $P < 0.0001$; SD: $P = 0.0031$). There was also a significant difference in food intake between
34 long photoperiod-chow and short photoperiod-chow rats ($P = 0.004$). After 28 days, energy
35 intake was higher in long photoperiod-HF than in short photoperiod chow rats ($P = 0.0002$),
36 but all other pairwise comparisons did not reveal a significant difference (Figure 2D).

1 **Novel object recognition before intervention**

2 A total of three rats were excluded from the analysis as they failed to explore both objects in
3 one or both trials of the task. There was no significant difference in time spent exploring the
4 two identical objects during the acquisition trial in any of the treatment groups (short
5 photoperiod-chow, $P = 0.364$; short photoperiod-HF, $P = 0.921$; long photoperiod-chow $P =$
6 0.625 ; long photoperiod-HF, $P = 0.699$; Figure 3A). In the retention trial, all four groups
7 explored the novel object significantly more than the familiar object (short photoperiod-chow,
8 $P = 0.016$; short photoperiod-HF, $P = 0.047$; long photoperiod-chow, $P = 0.033$; long
9 photoperiod-HF, $P = 0.009$; Figure 3B). A one-way ANOVA revealed no significant difference
10 in locomotor activity between the groups $F_{(3,28)} = 0.2033$, $P = 0.8931$ (Figure 3C). There was
11 also no significant differences in total exploration time in either the acquisition ($F_{(3,28)} =$
12 0.7881 , $P = 0.5119$) or retention ($F_{(3,28)} = 1.303$, $P = 0.2956$) trials (Table 2).

13 **Novel object recognition following intervention**

14 Rats were re-tested in the NOR task following four weeks of photoperiod and high fat diet
15 intervention. A total of eight rats were excluded from the analysis as they failed to explore
16 both objects in one or both trials of the task, therefore the final group numbers were short
17 photoperiod-chow ($n=5$), short photoperiod-HF ($n=6$), long photoperiod-chow ($n=7$) and long
18 photoperiod-HF ($n=6$). There was no significant difference in time spent exploring the two
19 identical objects during the acquisition trial in any of the treatment groups (short photoperiod-
20 chow, $P = 0.834$; short photoperiod-HF, $P = 0.883$; long photoperiod-chow, $P = 0.516$; long
21 photoperiod-HF, $P = 0.072$; Figure 4A). In the retention trial, only the long photoperiod-chow
22 group explored the novel object significantly more than the familiar object ($P = 0.040$; Figure
23 4B), whereas all other groups showed no preference (short photoperiod-chow, $P = 0.173$;
24 short photoperiod-HF, $P = 0.397$ and long photoperiod-HF, $P = 0.493$). A one-way ANOVA
25 revealed no significant difference in locomotor activity between the groups $F_{(3,23)} = 0.4207$, P
26 $= 0.7402$ (Figure 4C). There was also no significant differences in total exploration time in
27 either the acquisition ($F_{(3,23)} = 1.595$, $P = 0.222$) or retention ($F_{(3,23)} = 0.2262$, $P = 0.8.771$)
28 trials (Table 3).

29 **Discussion**

30 In this study, we examined the effect of photoperiod and high fat feeding on the cognitive
31 abilities of photoperiod-sensitive F344 rats. Our results show that short photoperiod results in
32 cognitive impairment **in the NOR test** in young male F344 rats independent of diet. In
33 contrast, rats in long photoperiod on chow diet are able to perform the object recognition task
34 suggesting their short-term recognition memory remains intact. However, a high fat diet
35 induces impairment in memory. This result was not unexpected, given that there is a well-
36 established link between obesity and a decline in cognitive performance (Sellbom and

1 Gunstad, 2012). High fat diets have previously been shown to cause cognitive decline in
2 animal models (Garcia-Mesa et al., 2012; McNeilly et al., 2011; Sobesky et al., 2014). There
3 is evidence that high fat diets can also promote plaque and tangle pathology in a mouse
4 model of Alzheimer's disease (Julien et al., 2010) suggesting a link between high fat diet and
5 neurodegenerative diseases. Previous work has established a link between altered energy
6 intake, insulin sensitivity and poor cognitive performance (McNeilly et al., 2012; McNeilly et
7 al., 2011). However, the precise molecular mechanisms that underlies the observed
8 decrease in cognitive performance remains to be established. Our results give us a first
9 insight to understand the impact of photoperiod and high fat diet on cognition in the 'healthy'
10 vs the 'obese' brain and might help to untangle the mechanisms involved in the regulation of
11 body weight (related to growth) from those involved in diet-induced obesity.

12 As anticipated, F344 rats on long photoperiod showed no impairment in the NOR task
13 indicating that increased cognitive performance helps them to anticipate changes in food
14 availability during breeding season (Buchanan et al., 2013). Maintaining (or even improving)
15 cognitive performance during long photoperiod will maximize their reproduction and survival.
16 In contrast, cognitive performance might be impaired in short photoperiod to reduce energy-
17 demanding physiological processes associated with the development and maintenance of
18 the underlying enlarged and complex neural structures (Buchanan et al., 2013). Cognitive
19 processing is energetically costly and requires neurogenesis and brain plasticity. Indeed, we
20 recently proposed that seasonal changes in neurogenesis serve as the basis of
21 photoperiodic control of energy balance and reproduction in seasonal animal models (Helfer
22 et al., 2019). Interestingly, here we found that changing rats to short photoperiod conditions
23 impairs their ability to differentiate between normal and familiar objects independent of diet.
24 We show that this is under direct photoperiod control because a simple switch in photoperiod
25 induces cognitive impairment in the F344 rats housed in short photoperiod.

26 In line with our previous studies, juvenile F344 rats housed in long photoperiod increase their
27 body weight compared to rats housed in short photoperiod (Ross et al., 2011; Ross et al.,
28 2015; Tavolaro et al., 2015) and are susceptible to mild obesity after four weeks of high fat
29 feeding (Ross et al., 2015). Similarly, the results of food intake are consistent with our
30 previous reports and extend them to show that young F344 rats, irrespective of single or pair
31 housing, have higher food intake in long photoperiod relative to short photoperiod. While
32 there is evidence that food intake is higher in single housed rats, relative to socially housed
33 rats, body weight is not effected by housing conditions (Schipper et al., 2018). Given that
34 photoperiodic response is predominantly assessed by changes in body weight which is
35 accompanied by changes in food intake, our results suggest that in future photoperiod

1 studies pair housing can be utilized to avoid emotional stress in the study animals (McLean
2 et al., 2010).

3 A limitation of the current study is the low number of animals used in the NOR task. Previous
4 studies from our laboratory suggest robust and reproducible effects in NOR when using 8 to
5 10 rats (McLean et al., 2011; McLean et al., 2016; McLean et al., 2017). In this study we
6 started with 8 rats per group but due to the application of our exclusion criteria final group
7 numbers ranged between 5 and 8. Nevertheless, statistically significant differences were
8 observed between the groups.

9 While the present study does not address molecular mechanisms underlying the effect of
10 photoperiod on cognition, recent studies have implicated that mechanisms may involve
11 insulin, leptin, BDNF, inflammatory pathways and blood brain barrier dysfunction (Cordner
12 and Tamashiro, 2015). Considering that cognitive decline after high fat feeding is a
13 pathophysiological response, it may be expected that different mechanisms are in place to
14 regulate cognition in the 'healthy' seasonal brain. Interestingly, photoperiod-regulated
15 changes in inflammatory signals have been proposed to be involved in the photoperiodic
16 response in seasonal mammals (Helfer et al., 2019). Microarray analysis of F344 rat
17 hypothalami identified a range of inflammatory chemokines and cytokines in response to
18 photoperiod (Helfer et al., 2016; Ross et al., 2011). NF- κ B, a major transcription factor
19 regulating the innate and adaptive immune response, has been investigated in more detail
20 and was found to be upregulated in the hypothalamus of F344 rats housed in short
21 photoperiod and regulated by TSH (Stoney et al., 2017). This is noteworthy, because it has
22 been suggested that the immune system regulates cognitive function in seasonal animals
23 (Buchanan et al., 2013). In support, short photoperiod Siberian hamster injected with
24 lipopolysaccharide exhibit diminished fever response with decreased locomotor inactivity
25 (Fonken et al., 2012). However, it is currently unknown whether this effect is mediated by
26 direct communication or a trade-off between the nervous and immune systems (Buchanan et
27 al., 2013). In future work, it will be interesting to investigate whether photoperiodic regulation
28 of the immune system and inflammatory pathways mediates cognitive function.

29 In conclusion, our study demonstrates for the first time an association between short
30 photoperiod and cognitive impairment and provides the basis for mechanistic studies into
31 how photoperiod effects cognition in the healthy brain. Recent evidence indicates that
32 photoperiod has an impact on animal behavior even in non-photoperiodic Wistar rats (Barnes
33 et al., 2017), thus a better understanding of how photoperiod effects cognition might lead to
34 novel preventive and therapeutic strategies in humans.

35

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8 **References**

- 9 Barnes, A.K., Smith, S.B., Datta, S., 2017. Beyond emotional and spatial processes:
10 Cognitive dysfunction in a depressive phenotype produced by long photoperiod exposure.
11 PLoS One 12, e0170032.
- 12 Buchanan, K.L., Grindstaff, J.L., Pravosudov, V.V., 2013. Condition dependence,
13 developmental plasticity, and cognition: implications for ecology and evolution. Trends Ecol.
14 Evol. 28, 290-296.
- 15 Buie, J.J., Watson, L.S., Smith, C.J., Sims-Robinson, C., 2019. Obesity-related cognitive
16 impairment: The role of endothelial dysfunction. Neurobiol. Dis. 132, 104580.
- 17 Clayton, N.S., Reboresda, J.C., Kacelnik, A., 1997. Seasonal changes of hippocampus
18 volume in parasitic cowbirds. Behav. Processes 41, 237-243.
- 19 Cohen, S.J., Stackman, R.W., Jr., 2015. Assessing rodent hippocampal involvement in the
20 novel object recognition task. A review. Behav. Brain Res. 285, 105-117.
- 21 Cordner, Z.A., Tamashiro, K.L., 2015. Effects of high-fat diet exposure on learning & memory.
22 Physiol. Behav. 152, 363-371.
- 23 Del Olmo, N., Ruiz-Gayo, M., 2018. Influence of high-fat diets consumed during the juvenile
24 period on hippocampal morphology and function. Front. Cell. Neurosci. 12, 439.
- 25 Ennaceur, A., Delacour, J., 1988. A new one-trial test for neurobiological studies of memory
26 in rats. 1: Behavioral data. Behav. Brain Res. 31, 47-59.
- 27 Fonken, L.K., Bedrosian, T.A., Michaels, H.D., Weil, Z.M., Nelson, R.J., 2012. Short
28 photoperiods attenuate central responses to an inflammogen. Brain. Behav. Immun. 26, 617-
29 622.
- 30 Galea, L.A., Kavaliers, M., Ossenkopp, K.P., 1996. Sexually dimorphic spatial learning in
31 meadow voles *Microtus pennsylvanicus* and deer mice *Peromyscus maniculatus*. J. Exp. Biol.
32 199, 195-200.
- 33 Galea, L.A., Kavaliers, M., Ossenkopp, K.P., Innes, D., Hargreaves, E.L., 1994. Sexually
34 dimorphic spatial learning varies seasonally in two populations of deer mice. Brain Res. 635,
35 18-26.

1 Garcia-Mesa, Y., Gimenez-Llort, L., Lopez, L.C., Venegas, C., Cristofol, R., Escames, G.,
2 Acuna-Castroviejo, D., Sanfeliu, C., 2012. Melatonin plus physical exercise are highly
3 neuroprotective in the 3xTg-AD mouse. *Neurobiol. Aging* 33, 1124 e1113-1129.

4 Glowa, J.R., Geyer, M.A., Gold, P.W., Sternberg, E.M., 1992. Differential startle amplitude
5 and corticosterone response in rats. *Neuroendocrinology* 56, 719-723.

6 Grayson, B., Leger, M., Piercy, C., Adamson, L., Harte, M., Neill, J.C., 2015. Assessment of
7 disease-related cognitive impairments using the novel object recognition (NOR) task in
8 rodents. *Behav. Brain Res.* 285, 176-193.

9 Harrington, G.M., 2013. Strain differences in open-field behavior of the rat. *Psychon. Sci.* 27,
10 51-53.

11 Helfer, G., Barrett, P., Morgan, P.J., 2019. A unifying hypothesis for control of body weight
12 and reproduction in seasonally breeding mammals. *J. Neuroendocrinol.* 31, e12680.

13 Helfer, G., Dumbell, R., 2020. Endocrine drivers of photoperiod response. *Curr Opin*
14 *Endocrine Metabol Res* 11, 49-54.

15 Helfer, G., Ross, A.W., Morgan, P.J., 2013. Neuromedin U partly mimics thyroid-stimulating
16 hormone and triggers Wnt/ β -Catenin signalling in the photoperiodic response of F344 rats. *J.*
17 *Neuroendocrinol.* 25, 1264-1272.

18 Helfer, G., Ross, A.W., Russell, L., Thomson, L.M., Shearer, K.D., Goodman, T.H.,
19 McCaffery, P.J., Morgan, P.J., 2012. Photoperiod regulates Vitamin A and Wnt/ β -Catenin
20 signaling in F344 rats. *Endocrinology* 153, 815-824.

21 Helfer, G., Ross, A.W., Thomson, L.M., Mayer, C.D., Stoney, P.N., McCaffery, P.J., Morgan,
22 P.J., 2016. A neuroendocrine role for chemerin in hypothalamic remodelling and
23 photoperiodic control of energy balance. *Sci. Rep.* May 26, 26830.

24 Helfer, G., Stevenson, T.J., 2020. Pleiotropic effects of proopiomelanocortin and VGF nerve
25 growth factor inducible neuropeptides for the long-term regulation of energy balance. *Mol.*
26 *Cell. Endocrinol.* 514, 110876.

27 Julien, C., Tremblay, C., Phivilay, A., Berthiaume, L., Emond, V., Julien, P., Calon, F., 2010.
28 High-fat diet aggravates amyloid-beta and tau pathologies in the 3xTg-AD mouse model.
29 *Neurobiol. Aging* 31, 1516-1531.

30 Kanoski, S.E., Davidson, T.L., 2011. Western diet consumption and cognitive impairment:
31 links to hippocampal dysfunction and obesity. *Physiol. Behav.* 103, 59-68.

32 Maille, A., Pillay, N., Schradin, C., 2015. Seasonal variation in attention and spatial
33 performance in a wild population of the African striped mouse (*Rhabdomys pumilio*). *Anim.*
34 *Cogn.* 18, 1231-1242.

35 Maille, A., Schradin, C., 2016. Survival is linked with reaction time and spatial memory in
36 African striped mice. *Biol. Lett.* 12.

1 McLean, F.H., Grant, C., Morris, A.C., Horgan, G.W., Polanski, A.J., Allan, K., Campbell,
2 F.M., Langston, R.F., Williams, L.M., 2018. Rapid and reversible impairment of episodic
3 memory by a high-fat diet in mice. *Sci. Rep.* 8, 11976.

4 McLean, S., Grayson, B., Harris, M., Protheroe, C., Woolley, M., Neill, J., 2010. Isolation
5 rearing impairs novel object recognition and attentional set shifting performance in female
6 rats. *J Psychopharmacol* 24, 57-63.

7 McLean, S.L., Grayson, B., Idris, N.F., Lesage, A.S., Pemberton, D.J., Mackie, C., Neill, J.C.,
8 2011. Activation of alpha7 nicotinic receptors improves phencyclidine-induced deficits in
9 cognitive tasks in rats: implications for therapy of cognitive dysfunction in schizophrenia. *Eur.*
10 *Neuropsychopharmacol.* 21, 333-343.

11 McLean, S.L., Grayson, B., Marsh, S., Zarroug, S.H., Harte, M.K., Neill, J.C., 2016. Nicotinic
12 alpha7 and alpha4beta2 agonists enhance the formation and retrieval of recognition memory:
13 Potential mechanisms for cognitive performance enhancement in neurological and
14 psychiatric disorders. *Behav. Brain Res.* 302, 73-80.

15 McLean, S.L., Harte, M.K., Neill, J.C., Young, A.M., 2017. Dopamine dysregulation in the
16 prefrontal cortex relates to cognitive deficits in the sub-chronic PCP-model for schizophrenia:
17 A preliminary investigation. *J Psychopharmacol* 31, 660-666.

18 McNeilly, A.D., Williamson, R., Balfour, D.J., Stewart, C.A., Sutherland, C., 2012. A high-fat-
19 diet-induced cognitive deficit in rats that is not prevented by improving insulin sensitivity with
20 metformin. *Diabetologia* 55, 3061-3070.

21 McNeilly, A.D., Williamson, R., Sutherland, C., Balfour, D.J., Stewart, C.A., 2011. High fat
22 feeding promotes simultaneous decline in insulin sensitivity and cognitive performance in a
23 delayed matching and non-matching to position task. *Behav. Brain Res.* 217, 134-141.

24 Noble, E.E., Kanoski, S.E., 2016. Early life exposure to obesogenic diets and learning and
25 memory dysfunction. *Curr Opin Behav Sci* 9, 7-14.

26 Okimura, K., Nakane, Y., Nishiwaki-Ohkawa, T., Yoshimura, T., 2021. Photoperiodic
27 regulation of dopamine signaling regulates seasonal changes in retinal photosensitivity in
28 mice. *Sci. Rep.* 11, 1843.

29 Pozner, T., Vistoropsky, Y., Moaraf, S., Heiblum, R., Barnea, A., 2018. Questioning
30 Seasonality of Neuronal Plasticity in the Adult Avian Brain. *Sci. Rep.* 8, 11289.

31 Ross, A.W., Helfer, G., Russell, L., Darras, V.M., Morgan, P.J., 2011. Thyroid hormone
32 signalling genes are regulated by photoperiod in the hypothalamus of F344 rats. *PLoS One* 6,
33 e21351.

34 Ross, A.W., Russell, L., Helfer, G., Thomson, L.M., Dalby, M.J., Morgan, P.J., 2015.
35 Photoperiod regulates lean mass accretion, but not adiposity, in growing F344 rats fed a high
36 fat diet. *PLoS One* 10, e0119763.

1 Schipper, L., Harvey, L., van der Beek, E.M., van Dijk, G., 2018. Home alone: a systematic
2 review and meta-analysis on the effects of individual housing on body weight, food intake
3 and visceral fat mass in rodents. *Obes. Rev.* 19, 614-637.

4 Sellbom, K.S., Gunstad, J., 2012. Cognitive function and decline in obesity. *J. Alzheimers Dis.*
5 30, S89-95.

6 Sherry, D.F., MacDougall-Shackleton, S.A., 2015. Seasonal change in the avian
7 hippocampus. *Front. Neuroendocrinol.* 37, 158-167.

8 Sobesky, J.L., Barrientos, R.M., De May, H.S., Thompson, B.M., Weber, M.D., Watkins, L.R.,
9 Maier, S.F., 2014. High-fat diet consumption disrupts memory and primes elevations in
10 hippocampal IL-1beta, an effect that can be prevented with dietary reversal or IL-1 receptor
11 antagonism. *Brain. Behav. Immun.* 42, 22-32.

12 Stevenson, T.J., Visser, M.E., Arnold, W., Barrett, P., Biello, S., Dawson, A., Denlinger, D.L.,
13 Dominoni, D., Ebling, F.J., Elton, S., Evans, N., Ferguson, H.M., Foster, R.G., Hau, M.,
14 Haydon, D.T., Hazlerigg, D.G., Heideman, P., Hopcraft, J.G., Jonsson, N.N., Kronfeld-Schor,
15 N., Kumar, V., Lincoln, G.A., MacLeod, R., Martin, S.A., Martinez-Bakker, M., Nelson, R.J.,
16 Reed, T., Robinson, J.E., Rock, D., Schwartz, W.J., Steffan-Dewenter, I., Tauber, E.,
17 Thackeray, S.J., Umstatter, C., Yoshimura, T., Helm, B., 2015. Disrupted seasonal biology
18 impacts health, food security and ecosystems. *Proc. Biol. Sci.* 282, 20151453.

19 Stoney, P.N., Rodrigues, D., Helfer, G., Khatib, T., Ashton, A., Hay, E.A., Starr, R.,
20 Kociszewska, D., Morgan, P., McCaffery, P., 2017. A seasonal switch in histone deacetylase
21 gene expression in the hypothalamus and their capacity to modulate nuclear signaling
22 pathways. *Brain. Behav. Immun.* Mar, 340-352.

23 Subhadeep, D., Srikumar, B.N., Shankaranarayana Rao, B.S., Kutty, B.M., 2020. Short
24 photoperiod restores ventral subicular lesion-induced deficits in affective and socio-cognitive
25 behavior in male Wistar rats. *J. Neurosci. Res.*

26 Tavolaro, F.M., Thomson, L.M., Ross, A.W., Morgan, P.J., Helfer, G., 2015. Photoperiodic
27 effects on seasonal physiology, reproductive status and hypothalamic gene expression in
28 young male F344 rats. *J. Neuroendocrinol.* 27, 79-87.

29 Tramontin, A.D., Brenowitz, E.A., 2000. Seasonal plasticity in the adult brain. *Trends*
30 *Neurosci.* 23, 251-258.

31 Van Der Staay, J.F., Blokland, A., 1996. Behavioral differences between outbred Wistar,
32 inbred Fischer 344, Brown Norway, and hybrid Fischer 344 × Brown Norway rats. *Physiol.*
33 *Behav.* 60, 97-109.

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1 **Tables**

2 **Table 1: Baseline data for each experimental group prior to testing.** All rats were healthy
 3 and test naïve. SP, short photoperiod; LP, long photoperiod; HF, high fat diet; n=8/group.

Treatment Group	Weight mean (g)	Weight SD	Weight median (g)
SP Chow	126.8	10.3	126.4
SP High fat	132.6	6.6	132.2
LP Chow	127.2	10.0	130.6
LP High fat	132.4	9.1	130.4

4
 5 **Table 2: Total exploration times in the novel object recognition test before random**
 6 **assignment to intervention.** Data shown are the mean total exploration time \pm SD. SP,
 7 short photoperiod; LP, long photoperiod; HF, high fat diet; n=7-8/group.

Total Exploration Time (sec)		
Treatment Group	Acquisition	Retention
SP Chow	17.1 \pm 7.0	17.4 \pm 7.2
SP High fat	22.4 \pm 9.5	19.1 \pm 11.7
LP Chow	22.1 \pm 6.2	13.4 \pm 6.9
LP High fat	20.1 \pm 6.0	11.9 \pm 5.4

8
 9 **Table 3: Total exploration times in the novel object recognition test following 4 weeks**
 10 **of intervention.** Data shown are the mean total exploration time \pm SD. SP, short photoperiod;
 11 LP, long photoperiod; HF, high fat diet; n=5-7/group.

Total Exploration Time (sec)		
Treatment Group	Acquisition	Retention
SP Chow	16.0 \pm 5.7	13.6 \pm 9.6
SP High fat	9.5 \pm 4.2	12.3 \pm 6.2
LP Chow	14.6 \pm 4.1	11.9 \pm 10.4
LP High fat	13.7 \pm 7.0	15.8 \pm 10.4

1 **Figures and legends**

2 **Figure 1: Schematic study design and timeline.** Rats were acclimatized in 12:12h
3 photoperiod on chow diet. Before photoperiod and diet intervention, rats were tested in the
4 novel recognition test. Rats were then randomly assigned to four groups. Two groups were
5 switched to short photoperiod and two groups to long photoperiod. One group in each
6 photoperiod was provided *ad libitum* with either a high fat diet or nutrient-matched chow diet.
7 Rats in photoperiod and diet groups were re-tested after 28 days of intervention. SP, short
8 photoperiod; LP, long photoperiod; HF, high fat diet.

9 **Figure 2: Effect of photoperiod and high fat diet on body weight, testes weight, food**
10 **intake and energy intake. (A)** Body weight was significantly lower in short photoperiod-HF
11 compared to long photoperiod-HF **(B)** Paired testes weight of male F344 rats after 4 weeks
12 exposure to photoperiod and high fat diet. Data were analysed by One-way ANOVA,
13 Significance is shown compared to the long photoperiod-chow group. * $P < 0.05$; ** $P < 0.01$
14 **(C)** Food intake in grams was highest in the long photoperiod-chow group and **(D)** energy
15 intakes in Kcal was highest in the long photoperiod-HF group. For (A), (C) and (D), data were
16 analysed by three-way RM ANOVA followed by Tukey's multiple comparisons test. Data are
17 presented as mean \pm SD (n=8/group). SP, short photoperiod; LP, long photoperiod; HF, high
18 fat diet.

19 **Figure 3: Novel object recognition before intervention.** Performance in the novel object
20 recognition (NOR) task before groups were exposed to short/long photoperiod chow/high fat
21 interventions. Data are expressed as the mean \pm SD. (n=7-8 per group). **(A)** Mean
22 exploration time of identical objects in the acquisition phase. **Data were analyzed by**
23 **Student's paired t-test, no significant difference observed between left and right object. (B)**
24 Mean exploration time of a familiar object and a novel object in the retention trial following.
25 Data were analyzed by Student's paired t-test. * $P < 0.05$, ** $P < 0.01$; Significant difference
26 between time spent exploring the familiar and novel object. **(C)** Mean total number of line
27 crossings in the acquisition and retention trials. **One-way ANOVA revealed no significant**
28 **difference in locomotor activity between the groups.** SP, short photoperiod; LP, long
29 photoperiod; HF, high fat diet

30 **Figure 4: Effect of photoperiod and high fat diet on novel object recognition.** The effect
31 of short photoperiod/long photoperiod chow/HF intervention on the novel object recognition
32 task. Data are expressed as the mean \pm SD. (n=5-7 per group). **(A)** Mean exploration time of
33 identical objects in the acquisition phase. **Data were analyzed by Student's paired t-test, no**
34 **significant difference observed between left and right object. (B)** Mean exploration time of a
35 familiar object and a novel object in the retention trial following. Data were analyzed by

1 Student's paired t-test. * $P < 0.05$; Significant difference between time spent exploring the
2 familiar and novel object. **(C)** Mean total number of line crossings in the acquisition and
3 retention trials. **One-way ANOVA revealed no significant difference in locomotor activity**
4 **between the groups.** SP, short photoperiod; LP, long photoperiod; HF, high fat diet

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